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| (54) Title: A BIFUNCTIONAL OR BIVALENT ANTIBODY FRAGMENT ANALOGUE | | | |
| (57) Abstract <p>The invention relates to a bispecific or bivalent antibody fragment analogue comprising a binding complex containing two polypeptide chains, whereby one polypeptide chain comprises two heavy chain variable domains (V_H) in series and the other polypeptide chain comprises two light chain variable domains (V_L) in series, the binding complex further containing two pairs of variable domains (V_H-A/V_L-A and V_H-B/V_L-B). The two V_H's and/or the two V_L's are connected directly or via an intermediate peptide linker. Also a production method for such antibody fragment analogues is disclosed.</p> | | | |

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Title: A bifunctional or bivalent antibody fragment analogue

The invention relates to new bispecific or bivalent antibody fragment analogues, a process for preparing such antibody fragment analogues and various uses of such antibody fragment analogues.

Background of the invention and prior art

1. Antibody structure

Antibody molecules typically are Y-shaped molecules whose basic unit consist of four polypeptides, two identical heavy chains and two identical light chains, which are covalently linked together by disulfide bonds. Each of these chains is folded in discrete domains. The C-terminal regions of both heavy and light chains are conserved in sequence and are called the constant regions, also known as C-domains. The N-terminal regions, also known as V-domains, are variable in sequence and are responsible for the antibody specificity. The antibody specifically recognizes and binds to an antigen mainly through six short complementarity-determining regions located in their V-domains (see Figure 1).

In this specification abbreviations are used having the following meaning.

| | |
|-------------------|--|
| C-domain: | Constant domain |
| V-domain: | Variable domain |
| V _L | : Variable domain of the light chain |
| 30 V _H | : Variable domain of the heavy chain |
| Fv | : dual chain antibody fragment containing both a V _H and a V _L |
| scFv | : single-chain Fv (V _H and V _L genetically linked either directly or via a peptide linker) |
| 35 CDR | : Complementarity-determining region |
| ELISA | : Enzyme Linked Immuno Sorbent Assay |
| PCR | : Polymerase Chain Reaction |

IPTG : IsoPropyl- β -ThioGalactopyranoside
PBS : Phosphate Buffered Saline
PBST : Phosphate Buffered Saline with 0.15% Tween
TMB : 3,3',5,5'-TetraMethylBenzidine

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It is generally known that proteolytic digestion of an antibody with papain yields three fragments. The fragment containing the CH₂ and CH₃ domains of the two heavy chains connected by the complete hinge (see Figure 1) crystallises very easily and was therefore called Fc fragment. The two other fragments are identical and were called Fab fragments, as they contained the antigen-binding site. Digestion with pepsin is such that the two Fab's remain connected via the hinge, forming only two fragments: Fc' and Fab₂.

The Fv is the smallest unit of an antibody which still contains the complete binding site (see Figure 1) and full antigen binding activity. It consists of only the V-domains of the heavy and light chains thus forming a small, heterodimeric variable fragment or Fv. Fv's have a molecular weight of about 25 kD, which is only one sixth of the parent whole antibody (in the case of an IgG). Previously Fv's were only available by proteolysis in a select number of cases (Givol, 1991). The production of Fv's can now be achieved more routinely using genetic engineering methods through cloning and expressing DNA encoding only the V-domains of the antibody of interest. Smaller fragments, such as individual V-domains (Domain Antibodies or dABs, Ward et al., 1989), and even individual CDR's (Williams et al., 1989; Taub et al., 1989) were shown to retain the binding characteristics of the parent antibody. However, this is not achievable on a routine basis: most naturally occurring antibodies need both a V_H and a V_L to retain full immunoreactivity. For example, in the case of V_H D1.3 (Ward et al., 1989), although it still binds hen egg lysozyme (HEL) with an affinity close to that of the parent antibody, it was shown that loss of

specificity was observed in that it can no longer distinguish turkey lysozyme from HBL, whereas the Fv can (Berry and Davies, 1992). Although murine dABs can be obtained more routinely from spleen libraries (Ward et al., 1989), the approach is unsustainable because of the many problems associated with their production and physical behaviour: expression is extremely poor, affinity tends to be low, stability and solubility in water is low, and non-specific binding is usually very high. According to the literature a possible explanation of these undesirable characteristics is the exposure of the hydrophobic residues which are normally buried in the V_H - V_L interface. The exposed hydrophobic patches are thought to contribute to aggregation of the protein inside the cells and/or in the culture medium, leading to poor expression and/or poor solubility (Anthony et al., 1992; Ward et al., 1989). The hydrophobic patches can also explain the high non-specific binding described by Berry and Davies, 1992. These problems clearly limit the usefulness of these molecules.

Most of the Camelid antibodies appear to be an exception to this rule in that they only need one V-domain, namely V_H , to specifically and effectively bind an antigen (Hamers-Castermans et al., 1993). In addition, preliminary data indicate that they seem not to suffer from the disadvantages of mouse dABs, as these camelid antibodies or fragments thereof are soluble and have been shown to express well in yeast and *Aspergillus* moulds. These observations can have important consequences for the production and exploitation of antibody-based products, see patent application WO 94/25591 (UNILEVER et al., first priority date 29.04.93).

2. Production of antibody fragments

Several microbial expression systems have already been developed for producing active antibody fragments, e.g. the production of Fab in various hosts, such as *E. coli* (Better et al., 1988, Skerra and Plückthun, 1988, Carter et al.,

1992), yeast (Horwitz et al., 1988), and the filamentous fungus *Trichoderma reesei* (Nyyssönen et al., 1993) has been described. The recombinant protein yields in these alternative systems can be relatively high (1-2 g/l for Fab secreted to the periplasmic space of *E. coli* in high cell density fermentation, see Carter et al., 1992), or at a lower level, e.g. about 0.1 mg/l for Fab in yeast in fermenters (Horwitz et al., 1988), and 150 mg/l for a fusion protein CBHI-Fab and 1 mg/l for Fab in *Trichoderma* in fermenters (Nyyssönen et al., 1993) and such production is very cheap compared to whole antibody production in mammalian cells (hybridoma, myeloma, CHO). Although the latter can give yields of the order of 1 g/l in high cell density fermentation, it is a time-consuming and very expensive manufacturing method resulting in a cost price of about 1000 £/gram of antibody. It was further demonstrated that plants can be used as hosts for the production of both whole antibodies (Hiatt et al., 1989) and scFv's (Owen et al., 1992, Firek et al., 1993), whereby yields of upto 0.5% of the total soluble protein content in tobacco leaves were mentioned.

The fragments can be produced as Fab's or as Fv's, but additionally it has been shown that a V_H and a V_L can be genetically linked in either order by a flexible polypeptide linker, which combination is known as an scFv (Bird et al. (1988), Huston et al. (1988), and granted patent EP-B-0281604 (GENEX/ENZON LABS INC.; first priority date 02-09-1986).

3. Bivalent and bispecific antibodies and antibody fragments

The antibody fragments Fab, Fv and scFv differ from whole antibodies in that the antibody fragments carry only a single antigen-binding site. Recombinant fragments with two binding sites have been made in several ways, for example, by chemical cross-linking of cysteine residues introduced at the C-terminus of the V_H of an Fv (Cumber et al., 1992),

or at the C-terminus of the V_L of an scFv (Pack and Plückthun, 1992), or through the hinge cysteine residues of Fab's (Carter et al., 1992). Another approach to produce bivalent antibody fragments is described by Kostelny et al. (1992) and Pack and Plückthun (1992) and is based on the inclusion of a C-terminal peptide that promotes dimerization.

When two different specificities are desired, one can generate bispecific antibody fragments. The traditional approach to generate bispecific whole antibodies was to fuse two hybridoma cell lines each producing an antibody having the desired specificity. Because of the random association of immunoglobulin heavy and light chains, these hybrid hybridomas produce a mixture of up to 10 different heavy and light chain combinations, only one of which is the bispecific antibody (Milstein and Cuello, 1983).

Therefore, these bispecific antibodies have to be purified with cumbersome procedures, which considerably decrease the yield of the desired product.

Alternative approaches include in-vitro linking of two antigen specificities by chemical cross-linking of cysteine residues either in the hinge or via a genetically introduced C-terminal Cys as described above. An improvement of such in vitro assembly was achieved by using recombinant fusions of Fab's with peptides that promote formation of heterodimers (Kostelny et al., 1992). However, the yield of bispecific product in these methods is far less than 100%.

A more efficient approach to produce bivalent or bispecific antibody fragments, not involving in vitro chemical assembly steps, was described by Holliger et al. (1993).

This approach takes advantage of the observation that scFv's secreted from bacteria are often present as both monomers and dimers. This observation suggested that the V_H and V_L of different chains can pair, thus forming dimers and larger complexes. The dimeric antibody fragments, also named "diabodies" by Hollinger et al., in fact are small

bivalent antibody fragments that assembled in vivo. By linking the V_H and V_L of two different antibodies 1 and 2, to form "cross-over" chains $V_{H1}V_{L2}$ and $V_{H2}V_{L1}$ (see Figure 2B), the dimerisation process was shown to reassemble both antigen-binding sites. The affinity of the two binding sites was shown to be equal to the starting scFv's, or even to be 10-fold increased when the polypeptide linker covalently linking V_H and V_L was removed, thus generating two proteins each consisting of a V_H directly and covalently linked to a V_L not pairing with the V_H (see Figure 2C). This strategy of producing bispecific antibody fragments was also described in several patent applications. Patent application WO 94/09131 (SCOTGEN LTD; priority date 15.10.92) relates to a bispecific binding protein in which the binding domains are derived from both a V_H and a V_L region either present at two chains or linked in an scFv, whereas other fused antibody domains, e.g. C-terminal constant domains, are used to stabilise the dimeric constructs. Patent application WO 94/13804 (CAMBRIDGE ANTIBODY TECHNOLOGY / MEDICAL RESEARCH COUNCIL; first priority date 04.12.92) relates to a polypeptide containing a V_H and a V_L which are incapable of associating with each other, whereby the V-domains can be connected with or without a linker.

Mallender and Voss, 1994 (also described in patent application WO 94/13806; DOW CHEMICAL CO; priority date 11.12.92) reported the in vivo production of a single-chain bispecific antibody fragment in *E. coli*. The bispecificity of the bivalent protein was based on two previously produced monovalent scFv molecules possessing distinct specificities, being linked together at the genetic level by a flexible polypeptide linker. The thus formed V_{H1} -linker- V_{L1} -linker- V_{H2} -linker- V_{L2} fragment (see Figure 2A) was shown to contain both antigen binding specificities 1 and 2. (1= anti-fluorescein, 2= anti-single-stranded DNA).

Traditionally, whenever single-chain antibody fragments are referred to, a single molecule consisting of one heavy chain linked to one (corresponding) light chain in the presence or absence of a polypeptide linker is implicated.

- 5 When making bivalent or bispecific antibody fragments through the 'diabody' approach (Holliger et al., (1993) and patent application WO 94/09131) or by the 'double scFv' approach (Mallender and Voss, 1994 and patent application WO 94/13806), again the V_H is linked to a (the
10 corresponding) V_L .

- It is realised that claims 32 and 33 of patent application WO 93/11161 (ENZON INC.; priority date 25.11.91) and the corresponding passages in that specification on page 22, lines 1-10 may read on a polypeptide comprising two V_L 's
15 fused together via a flexible polypeptide linker, and on a polypeptide comprising two V_H 's fused together via a flexible polypeptide linker, respectively. However, no examples were given to substantiate this approach, thus it was in fact a hypothetical possibility instead of an
20 actually produced compound.

- A skilled person would not have expected that such approach would be viable for at least three reasons. Firstly, it is widely recognised that immunoglobulin heavy chains (excluding the above described camel immunoglobulins) have
25 very limited solubility and spontaneously precipitate out of aqueous solution when isolated from their light chain partners. Secondly, several groups have shown (Ward et al., 1989, Berry and Davies, 1992, and Anthony et al., 1992) that expression of V_H 's in the absence of V_L 's is hampered
30 by extremely poor yields of unstable product with many undesirable properties, e.g. non-specific binding. Thirdly in patent application WO 94/13804 it was described on page 31 lines 10-12, that in computer modelling experiments they could not model as heterodimers V_H - V_H and V_L - V_L given the
35 constraints of short linkers.

Thus the simple suggestion given in patent application WO 93/11161 is not an enabling disclosure leading a skilled

person to try with a reasonable expectation of success whether such suggestion would work; therefore, that patent application should not be considered as relevant prior art for the present invention.

5

Summary of the invention

The present invention provides a bispecific or bivalent antibody fragment analogue, which comprises a binding complex containing two polypeptide chains, one of which
10 comprises two times a variable domain of a heavy chain (V_H) in series and the other comprises two times a variable domain of a light chain (V_L) in series.

In one aspect of the invention one chain of the antibody fragment analogue comprises a first V_H (V_H -A) connected to a
15 second V_H (V_H -B) and the other chain comprises a first V_L (V_L -A) connected to a second V_L (V_L -B). In a preferred embodiment of this aspect one chain comprises a first V_H (V_H -A) followed by a second V_H (V_H -B), thus [V_H -A * V_H -B], and the other chain comprises a first V_L (V_L -A) preceded by
20 a second V_L (V_L -B), thus [V_L -B * V_L -A]. For some embodiments of this aspect the two V_H 's are directly connected to each other, but for other embodiments of this aspect of the invention the two V_L 's are directly connected to each other. According to another embodiment of this aspect of the
25 invention the two V_H 's are connected to each other by a linker and also the two V_L 's are connected to each other by a linker. Such a linker usually comprises at least one amino acid residue.

According to a special embodiment of this aspect of the
30 invention one chain comprises a first V_H (V_H -A) followed by a second V_H (V_H -B), thus [V_H -A * V_H -B], and the other chain comprises a first V_L (V_L -A) followed by a second V_L (V_L -B), thus [V_L -A * V_L -B], and in which the two V_H 's are connected to each other by a linker and also the two V_L 's are
35 connected to each other by a linker, whereas each linker comprises at least 10 amino acid residues.

According to the above aspect of the invention with A being different from B there are provided bispecific antibody fragment analogues.

- According to another aspect of the invention the
- 5 specificities A and B are the same resulting in a bivalent antibody fragment.

According to a further aspect of the invention the bispecific or bivalent antibody fragment analogues can be used in a diagnostic technique or for immunoassays, in a

10 purification method, for therapy, or in other methods in which immunoglobulins or fragments thereof are used. Such uses are well-known in the art.

The invention also provides a process for producing the antibody fragments of the invention in that a host is

15 transformed by incorporating into that host a DNA encoding the two V_H 's with or without a connecting linker and a DNA encoding the two V_L 's with or without a connecting linker. Preferably the two DNA's are placed in a dicistronic arrangement.

- 20 It is also possible that the two linked V_H 's and the two linked V_L 's are produced separately by different hosts, after which the linked V_H 's produced by one host can be combined with the linked V_L 's produced by the other host. The hosts can be selected from the group consisting of
- 25 prokaryotic bacteria of which examples are Gram-negative bacteria, e.g. *E. coli*, and Gram-positive bacteria, e.g. *B. subtilis* or lactic acid bacteria, lower eukaryotes examples of which are yeasts, e.g. belonging to the genera *Saccharomyces*, *Kluyveromyces*, or *Trichoderma*, moulds, e.g.
- 30 belonging to the genera *Aspergillus* and *Neurospora*, and higher eukaryotes, examples of which are plants, e.g. tobacco, and animal cells, examples of which are myeloma cells and CHO cells. The techniques to transform a host by genetic engineering methods in order to have a desirable
- 35 polypeptide produced by such host are well-known to persons skilled in the art as is evident from the literature

mentioned above under the heading "Background of the invention and prior art".

Brief description of the drawings

- 5 Figure 1 depicts in schematic form the structure of a typical antibody (immunoglobulin) molecule.
- Figure 2 shows a schematic representation of published arrangements of heavy and light chain V-domain gene fragments that have been proven to
- 10 produce bispecific antibody fragments.
- Figure 3 shows in diagrammatic form the suggested arrangement of the V-domains of a double head antibody fragment according to the invention with the V-domains in the following order:
- 15 $V_HA-V_HB + V_LB-V_LA$.
- Figure 4 shows the nucleotide sequence of the *EcoRI*-*HindIII* insert of pUR.4124 containing DNA (see SEQ ID NO: 1) encoding V_L Lys-Linker- V_H Lys (see SEQ ID NO: 2).
- 20 Figure 5 shows the nucleotide sequence of the *HindIII*-*EcoRI* insert of plasmid Fv.3418 (see SEQ ID NO: 3) containing DNA encoding pelB leader- V_H 3418 (see SEQ ID NO: 4) and DNA encoding pelB leader- V_L 3418 (see SEQ ID NO: 5).
- 25 Figure 6 shows the nucleotide sequence of the *HindIII*-*EcoRI* insert of plasmid Fv.4715-myc (see SEQ ID NO: 6) containing DNA encoding pelB leader- V_H 4715 (see SEQ ID NO: 7) and DNA encoding pelB leader- V_L 4715-Myc tag (see SEQ ID NO: 8).
- 30 Figure 7 shows the nucleotide sequence of the *HindIII*-*EcoRI* insert of scFv.4715-myc containing DNA (see SEQ ID NO: 9) encoding pelB leader- V_H 4715-Linker- V_L 4715-Myc tag (see SEQ ID NO: 10).
- 35 Figure 8 a/b shows the nucleotide sequence of the *HindIII*-*EcoRI* insert of pGOSA.E (see SEQ ID NO: 11) containing DNA encoding pelB leader- V_H 4715-

Linker-V_L3418 (see SEQ ID NO: 12) and DNA encoding pelB leader-V_L3418-Linker-V_H4715 (see SEQ ID NO: 13).

- 5 Figure 9 gives an overview of the oligonucleotides and their positions in pGOSA.E that can be used to replace V-domain gene fragments.
- 10 Figure 10 illustrates the amino acid sequence of the V_H-V_H and V_L-V_L domain junctions in fusion polypeptides GOSA.E (see amino acids 114-145 in SEQ ID NO: 12 and amino acids 102-128 in SEQ ID NO: 13), GOSA.V (see SEQ ID NO: 30 and amino acids 102-128 in SEQ ID NO: 13), GOSA.S (see amino acids 114-145 in SEQ ID NO: 12 and SEQ ID NO: 31) and GOSA.T (see SEQ ID NO: 30 and SEQ ID NO: 31).
- 15 Figure 11 shows the specificity of *Streptococcus* binding of scFv.4715-myc.
- 20 Figure 12 shows the specificity of glucose oxidase targeting onto the surface of various *Streptococcus* strains by GOSA.E.
- 25 Figure 13 shows the specificity of glucose oxidase targeting onto the surface of various *Streptococcus* strains by GOSA.V.
- 30 Figure 14 shows the specificity of glucose oxidase targeting onto the surface of various *Streptococcus* strains by GOSA.S.
- 35 Figure 15 shows the specificity of glucose oxidase targeting onto the surface of various *Streptococcus* strains by GOSA.T.
- 30 Figure 16 shows the results of an ELISA. Individual fractions of a gelfiltration experiment using partially purified GOSA.E as feedstock were tested for glucose oxidase and *Streptococcus sanguis* bispecific binding activity.
- 35 Figure 17 shows the results of an ELISA. Individual fractions of a gelfiltration experiment using partially purified GOSA.V as feedstock were

- tested for glucose oxidase and *Streptococcus sanguis* bispecific binding activity.
- Figure 18 shows the results of an ELISA. Individual fractions of a gelfiltration experiment using partially purified GOSA.S as feedstock were tested for glucose oxidase and *Streptococcus sanguis* bispecific binding activity.
- Figure 19 shows the results of an ELISA. Individual fractions of a gelfiltration experiment using partially purified GOSA.T as feedstock were tested for glucose oxidase and *Streptococcus sanguis* bispecific binding activity.
- Figure 20 shows the source of fragment PCR.I *BstEII/SacI*
- Figure 21 shows the source of fragment PCR.II *SfiI/EcoRI*
- Figure 22 shows the source of fragment PCR.III *NheI/SacI*
- Figure 23 shows the source of fragment PCR.IV *XhoI/EcoRI*
- Figure 24 shows the source of fragment PCR.V *Sall/EcoRI*
- Figure 25 shows the source of fragment PCR.VI *SfiI/NheI*
- Figure 26 shows the source of fragment PCR.VII *BstEII/NheI*
- Figure 27 shows the source of fragment PCR.VIII *XhoI/EcoRI*
- Figure 28 shows the source of fragment PCR.IX *BstEII/NheI*
- Figure 29 shows the source of fragment PCR.X *PstI/EcoRI*
- Figure 30 shows the construction of plasmid pGOSA.A
- Figure 31 shows the construction of plasmid pGOSA.B
- Figure 32 shows the construction of plasmid pGOSA.C
- Figure 33 shows the construction of plasmid pGOSA.D
- Figure 34 shows the construction of plasmid pGOSA.E
- Figure 35 shows the construction of plasmid pGOSA.V
- Figure 36 shows the construction of plasmid pGOSA.S
- Figure 37 shows the construction of plasmid pGOSA.T
- Figure 38 a/b shows the construction of plasmid pGOSA.G
- Figure 39 shows the construction of plasmid pGOSA.J
- Figure 40 shows the construction of plasmid pGOSA.Z
- Figure 41 shows the construction of plasmid pGOSA.AA

- Figure 42 shows the construction of plasmid pGOSA.AB
Figure 43 shows the construction of plasmid pGOSA.L
Figure 44 shows the construction of plasmid pGOSA.Y
Figure 45 shows the construction of plasmid pGOSA.X
5 Figure 46 shows the construction of plasmid pGOSA.AC
Figure 47 shows the construction of plasmid pGOSA.AD.

Table 1 shows the nucleotide sequence of the
oligonucleotides used to produce the constructs
10 described in this specification. Restriction sites
encoded by these primers are underlined.

Table 2 gives an overview of all GOSA constructs described
in this specification.

15 Table 2A describes intermediate constructs that
were not further tested.

Table 2B describes the dicistronic constructs.

Table 2C describes the monocistronic constructs.

20 Detailed description of the invention

In this specification the construction of an antibody
fragment analogue consisting of a two chain protein complex
is described, in which one of the chains consists of two
heavy chain V-domains and the other chain consists of the
25 two corresponding light chain V-domains in either order.
The variable domains are linked either directly or through
a polypeptide linker. Subsequent molecular modelling of
this combination suggested that the protein chains could
fold such that both binding sites are fully accessible,
30 provided that the connecting linkers are kept long enough
to span 30 to 35 Å.

Whereas in patent application WO 93/11161 it is explicitly
described that for the above described bispecific complexes
two flexible polypeptide linkers in the self assembling
35 complex are required, the present invention illustrated
here describes in particular the construction of a two
chain protein complex containing only one linker or no

linkers at all. The latter antibody fragment analogue thus consists of a two chain protein complex containing one polypeptide chain comprising heavy chain V-domains fused directly together and another polypeptide chain comprising the corresponding light chain V-domains fused together, both fusions in the absence of linkers. But also two chain protein complexes in which each chain comprises a linker between the two variable domains can be used as antibody fragment analogues according to the invention as described below with construct pGOSA.E. However, the two chain complexes containing only one linker or no linker at all are preferred. The abbreviation GOSA used in this specification relates to a combination of glucose oxidase and *Streptococcus sanguis*.

In this specification evidence is provided that these antibody fragment analogues ("double heads") contain both antigen binding specificities of the Fv's used to generate these bispecific antibody fragments. It is exemplified that these type of constructs according to the invention can be used to target the enzyme glucose oxidase to whole bacteria, using antibody fragments derived from hybridomas expressing antibodies directed against these antigens.

The present invention is now described by reference to some specific examples, which are included for purposes of illustration only and are not intended to limit the scope of the invention.

30 EXAMPLES

General experimental

Strains, Plasmids and Media

All cloning steps were performed in *E. coli* JM109 (*endA1*, *recA1*, *gyrA96*, *thi*, *hsdR17*(r_k , m_k^+), *relA1*, *supE44*, Δ (*lac-proAB*), [*F'*, *traD36*, *proAB*, *lacI'* Δ M15]). *E. coli* cultures were grown in 2xTY medium (16 g tryptone, 10 g yeast extract, 5 g NaCl per litre H₂O), where indicated

supplemented with 2% glucose and/or 100 µg/ml ampicillin. Transformations were plated out on SOBAG plates (20 g tryptone, 5 g yeast extract, 15 g agar, 0.5 g NaCl per litre H₂O plus 10 mM MgCl₂, 2% glucose, 100 µg/ml ampicillin) The expression vectors used are derivatives of pUC19. The oligonucleotide primers used in the PCR reactions were synthesized on an Applied Biosystems 381A DNA Synthesiser by the phosphoramidite method.

10 **Expression of GOSA constructs**

Colonies from freshly transformed JM109 plated onto SOBAG plates were used to inoculate 2xTY medium supplemented with 100 µg/ml ampicillin, 2% glucose. Cultures were shaken at 37°C to an OD₆₀₀ in the range of 0.5 to 1.0. Cells were pelleted by centrifugation and the supernatant was removed. The pelleted cells were resuspended in 2xTY medium with 100 µg/ml ampicillin, 1 mM IPTG, and grown for a further 18 hours at 25°C. Cells were pelleted by centrifugation and the supernatant, containing the secreted chains, used directly in an ELISA. The proteins in the periplasm of the pelleted cells were extracted by resuspending the cell pellet in 1/20 of the original culture volume of lysis buffer (20% sucrose, 200 mM Tris-HCl pH 7.5, 1 mM EDTA, 500 µg/ml lysozyme). After incubation at 25°C for 20 minutes an equal volume of H₂O was added and the incubation was continued for another 20 minutes. The suspension was spun at 10.000 g for 15 minutes and the supernatant containing the periplasmic proteins was used directly in an ELISA.

30 **ELISA**

96 well ELISA plates (Greiner HC plates) were activated overnight at 37°C with 200 µl/well of an 1/10 dilution of an over night culture of *Streptococcus* cells in 0.05 M sodium carbonate buffer at pH=9.5. Following one wash with PBST, the antigen sensitised plates were pre-blocked for 1 hour at 37°C with 200 µl/well blocking buffer (2% BSA, 0.15% Tween in PBS). Samples containing 50 µl blocking

buffer plus 50 μ l culture supernatants or periplasmic cell extracts (neat or diluted with PBS) were added to the *Streptococcus* sensitised plate and incubated for 2 hours at 37°C. Following 4 washes with PBS-T, 100 μ l of blocking
5 buffer containing glucose oxidase (50 μ g/ml) was added to every well. After incubation at 37°C for 1 hour unbound glucose oxidase was removed by 4 washes with PBS-T. Bound glucose oxidase was detected by adding 100 μ l substrate to each well (70 mM Na-citrate, 320 mM Na-phosphate, 27 mg/ml
10 glucose, 0.5 μ g/ml HRP, 100 μ g/ml TMB). The colour reaction was stopped after 1 hour by the addition of 35 μ l 2 M HCl and the A450 was measured (compare Figures 11/15).

Affinity purification of GOSA antibody fragments

15 GOSA.E, GOSA.V, GOSA.S and GOSA.T were partially purified by affinity chromatography. 100 ml periplasmic extract of each of these constructs was loaded onto a Glucose-oxidase-Sepharose column (CNBr-Sepharose, Pharmacia) prepared according to the manufacturer's instructions. After
20 extensive washes with PBS the bound GOSA antibody fragments were eluted in 0.1M glycine buffer at pH=2.8. The fractions were neutralised with Tris and analysed by polyacrylamide gel electrophoresis followed by silver staining and tested for the presence of double head activity.

25

EXAMPLE 1. Construction of the pGOSA double head expression vectors

30 In this Example the construction of a two chain protein complex is described, in which one of the chains consists of two heavy chain V-domains and the other chain consists of the two corresponding light chain V-domains. The variable domains are linked either directly or through a
35 polypeptide linker. The expression vectors used are derivatives of a pUC19 derived plasmid containing a HindIII-EcoRI fragment that in the case of plasmid

scFv.4715-myc contains a DNA fragment encoding one pelB signal sequence fused to the N-terminus of the V_H that is directly linked to the corresponding V_L of the antibody through a connecting flexible peptide linker, (Gly₄Ser)₃,
5 (present in SEQ ID NO: 2 as amino acids 109-123 and in SEQ ID NO: 10 as amino acids 121-135), thus generating a single-chain molecule (see Figure 7).

In the dual-chain Fv and the pGOSA expression vectors, the DNA fragments encoding both the V_H and V_L of the antibody
10 are preceded by a ribosome binding site and a DNA sequence encoding the pelB signal sequence in an artificial dicistronic operon under the control of a single inducible promoter (see Figures 5, 6 and 8). Expression of these constructs is driven by the inducible lacZ promoter. The
15 nucleotide sequence of the *HindIII*-*EcoRI* inserts of the plasmids pUR.4124, Fv.3418, Fv.4715-myc and scFv.4715-myc constructs used for the generation of the bispecific antibody fragments are given in Figures 4-7, respectively. Moreover, a culture of *E. coli* cells harbouring plasmid
20 scFv.4715-myc and a culture of *E. coli* cells harbouring plasmid Fv.3418 were deposited under the Budapest Treaty at the National Collection of Type Cultures (Central Public Health Laboratory) in London (United Kingdom) with deposition numbers NCTC 12916 and NCTC 12915, respectively.
25 In agreement with Rule 28 (4) EPC, or a similar arrangement for a State not being a Contracting State of the EPC, it is hereby requested that a sample of such deposit, when requested, will be submitted to an expert only.

The construction of pGOSA.E (see Figure 8 for the *HindIII*-*EcoRI* insert of pUC19) involved several cloning steps. The
30 appropriate restriction sites in the various domains were introduced by PCR directed mutagenesis using the oligonucleotides listed in Table 1 below. The pGOSA.E derivatives pGOSA.V, pGOSA.S and pGOSA.T with only one or
35 no linker sequence are derived from the pGOSA.E construct by removing the linker sequences by means of PCR directed mutagenesis with oligonucleotides listed in Table 1 below.

Table 1.

| | | |
|--------|---|-------------------|
| DBL.1 | 5'-CAC CAT CTC CAG AGA CAA TGG CAA G-3' | (=SEQ ID NO: 14) |
| DBL.2 | 5'-GAG CGC GAG CTC <u>GGC CGA ACC GGC C</u> ¹ GA TCC GCC | |
| 5 | ACC GCC AGA GCC-3' | (=SEQ ID NO: 15) |
| DBL.3 | 5'-CAG GAT CCG GCC <u>GGT TCG GCC</u> ¹ CAG GTC CAG CTG | |
| | CAA CAG TCA GGA-3' | (=SEQ ID NO: 16) |
| DBL.4 | 5'-CTA CAT <u>GAA TTC</u> ² <u>GCT AGC</u> ³ TTA TTA TGA GGA GAC | |
| | GGT GAC GGT GGT CCC TTG GC-3' | (=SEQ ID NO: 17) |
| 10 | DBL.5 5'-TAA TAA <u>GCT AGC</u> ³ GGA GCT GCA TGC AAA TTC TAT | |
| | TTC-3' | (= SEQ ID NO: 18) |
| DBL.6 | 5'-ACC AAG <u>CTC GAG</u> ⁴ ATC AAA CGG GG-3' | (= SEQ ID NO: 19) |
| DBL.7 | 5'-AAT GTC <u>GAA TTC</u> ² <u>GTC GAC</u> ⁵ TCC GCC ACC GCC AGA | |
| | GCC-3' | (= SEQ ID NO: 20) |
| 15 | DBL.8 5'-ATT GGA <u>GTC GAC</u> ⁵ ATC GAA CTC ACT CAG TCT CCA | |
| | TTC TCC-3' | (= SEQ ID NO: 21) |
| DBL.9 | 5'-TGA AGT <u>GAA TTC</u> ² <u>GCG GCC GC</u> ⁶ T TAT TAC CGT TTG | |
| | ATT TCG AGC TTG GTC CC-3' | (= SEQ ID NO: 22) |
| DBL.10 | 5'-CGA ATT <u>CGG TCA CC</u> ¹ G TCT CCT CAC AGG TCC AGT | |
| 20 | TGC AAC AG-3' | (= SEQ ID NO: 23) |
| DBL.11 | 5'-CGA ATT <u>CTC GAG</u> ⁴ ATC AAA CGG GAC ATC GAA CTC | |
| | ACT CAG TCT CC-3' | (= SEQ ID NO: 24) |
| DBL.12 | 5'-CGA ATT <u>CGG TCA CC</u> ¹ G TCT CCT CAC AGG TGC AGT | |
| | TGC AGG AG-3' | (= SEQ ID NO: 25) |
| 25 | PCR.51 5'-AGG T(C/G)(A/C) A(C/A) <u>C</u> TGC AG ⁷ (C/G) AGT C(A/T)G | |
| | G-3' | (= SEQ ID NO: 26) |
| PCR.89 | 5'-TGA GGA GAC <u>GGT GAC C</u> ¹ GT GGT CCC TTG GCC CC-3' | |
| | | (= SEQ ID NO: 27) |
| PCR.90 | 5'-GAC ATT <u>GAG CTC</u> ⁹ ACC CAG TCT CCA-3' | (= SEQ ID NO: 28) |
| 30 | PCR.116 5'-GTT AGA <u>TCT CGA G</u> ⁴ CT TGG TCC C-3' | (= SEQ ID NO: 29) |

1=*Sfi*I, 2=*Eco*RI, 3=*Nhe*I, 4=*Xho*I, 5=*Sal*I, 6=*Nor*I, 7=*Pst*I, 8=*Bst*EII, 9=*Sac*I

These three constructs lack some of the restriction sites at the new joining points. The V_HA-V_HB gene fragment without a linker lacks the 5' V_HB *SfiI* site. The V_LB-V_LA gene fragment without a linker lacks the 5' V_LA *SalI* site.

- 5 The position of the oligonucleotides in the pGOSA constructs given in Table 1 are shown in Figure 9. The pGOSA expression vectors and the oligonucleotides in Table 1 have been designed to enable most specificities to be cloned into the pGOSA constructs. Figure 10 shows the amino
- 10 acid sequence of the junctions between the V_HA-V_HB and V_LB-V_LA fragments encoded by DNA present in pGOSA.E, pGOSA.V, pGOSA.S and pGOSA.T. A more detailed description of the preparation of pGOSA.E, pGOSA.V, pGOSA.S and pGOSA.T is given in Example 5.

15

EXAMPLE 2. Bifunctional binding activity of GOSA double heads

- 20 In this Example we provide evidence that the above described molecules ("double heads"), i.e. the two chain protein complexes, contain both antigen binding specificities of the Fv's used to generate these multi-functional antibody fragment analogues. Figure 12-15 show
- 25 that GOSA.E, GOSA.V, GOSA.S and GOSA.T can be used to specifically target the enzyme glucose oxidase to several *Streptococcus sanguis* strains using antibody fragments derived from hybridoma's expressing antibodies directed against these antigens.
- 30 Comparison of the binding specificity of the GOSA constructs (see Figures 12-15) and the binding specificity of the scFv.4715-myc (see Figure 11) shows that the fine specificity of the anti-*Streptococcus sanguis* scFv.4715 is preserved in the GOSA "double heads".

35

EXAMPLE 3. FPLC analysis of GOSA double heads

Partially purified GOSA.E, GOSA.V, GOSA.S and GOSA.T samples (estimated to be 50-80% pure by polyacrylamide gel electrophoresis) were analysed on a Pharmacia FPLC Superose 12 column. The analysis was performed using PBS at a flow rate of 0.3 ml/minute. Eluate was monitored at 280 nm and 0.3 ml fractions were collected and analysed by ELISA. Usually GOSA.E, GOSA.V, GOSA.S and GOSA.T samples only gave one GOSA double head activity peak as determined by ELISA (see Figures 16-19). The position of this peak in the elution pattern indicated that the molecular weight of the GOSA double head is 40-50 kD. Since this molecular weight corresponds to the expected molecular weight of the $V_H2 + V_L2$ double head dimer, the conclusion is justified that GOSA.E, GOSA.V, GOSA.S and GOSA.T are primarily produced as dimeric molecules. Occasionally an activity peak with an apparent molecular weight of ~200 kD was observed (see Figure 16). The presence of Glucose Oxidase activity in these fractions (data not shown) indicate that these fractions contain GOSA double head complexed with glucose oxidase that was eluted with the GOSA sample from the glucose oxidase-sepharose affinity matrix.

EXAMPLE 4. Production of other double heads

The methods described in the previous Examples were used to produce other double heads, which also appeared to be active against the antigens for which they were developed. These other double heads had the following specificities:

- anti-*S. sanguis* / anti-beta-HCG,
- anti-*S. sanguis* / anti-urease,
- anti-*S. sanguis* / anti-hen-egg-lysozyme,
- anti-beta-HCG / anti-hen-egg-lysozyme,
- anti-hen-egg-lysozyme / anti-glucose oxidase,
- anti-huIgG / anti-glucose oxidase,

anti-urease / anti-glucose oxidase,
anti-lacto-peroxidase / anti-glucose oxidase,
anti-alpha-HCG / anti-glucose oxidase, and
anti-reactive-Red-6 / anti-glucose oxidase.

5

EXAMPLE 5. Detailed description of the preparation of
intermediate constructs pGOSA.A, pGOSA.B
pGOSA.C and pGOSA.D and their use for the
preparation of plasmid pGOSA.E and its
derivatives pGOSA.V, pGOSA.S and pGOSA.T

10

Oligonucleotides and PCR

The primary structures of the oligonucleotide primers used
in the construction of the bispecific 'pGOSA' constructs
are shown in Table 1 above. Reaction mixture used for
amplification of DNA fragments were 10 mM Tris-HCl, pH 8.3,
2.5 mM MgCl₂, 50 mM KCl, 0.01% gelatin (w/v), 0.1% Triton X-
100, 400 mM of each dNTP, 5.0 units of Vent DNA polymerase
(New England Biolabs), 100 ng of template DNA, and 500 ng
of each primer (for 100 µl reactions). Reaction conditions
were: 94°C for 4 minutes, followed by 33 cycles of each 1
minute at 94°C, 1 minute at 55°C, and 1 minute 72°C.

25 Plasmid DNA\Vector\Insert preparation and ligation\transformation.

Plasmid DNA was prepared using the 'Qiagen P-100 Midi-DNA
Preparation' system. Vectors and inserts were prepared by
digestion of 10 µg (for vector preparation) or 20 µg (for
insert preparation) with the specified restriction
endonucleases under appropriate conditions (buffers and
temperatures as specified by suppliers). Modification of
the DNA ends with Klenow DNA polymerase and
dephosphorylation with Calf Intestine Phosphorylase were
performed according to the manufacturers instructions.
Vector DNAs and inserts were separated through agarose gel
electrophoresis and purified with DEAE-membranes NA45

30

35

(Schleicher & Schuell) as described by Maniatis et al. Ligations were performed in 20 μ l volumes containing 30 mM Tris-HCl pH 7.8, 10 mM $MgCl_2$, 10 mM DTT, 1 mM ATP, 300-400 ng vector DNA, 100-200 ng insert DNA and 1 Weiss unit T_4 DNA ligase. After ligation for 2-4 h at room temperature, $CaCl_2$ competent *E. coli* JM109 (Maniatis) were transformed using 7.5 μ l ligation reaction. The transformation mixtures were plated onto SOBAG plates and grown overnight at 37°C. Correct clones were identified by restriction analysis and verified by automated dideoxy sequencing (Applied Biosystems).

Restriction digestion of PCR products

Following amplification each reaction was checked for the presence of a band of the appropriate size by agarose gel electrophoresis. One or two 100 μ l PCR reaction mixtures of each of the PCR reactions PCR.I - PCR.X (Figure 20-29), together containing approximately 2-4 μ g DNA product were subjected to phenol-chloroform extraction, chloroform extraction and ethanol precipitation. The DNA pellets were washed twice with 70% ethanol and allowed to dry. Next, the PCR products were digested overnight (18 h) in the presence of excess restriction enzyme in the following mixes at the specified temperatures and volumes.

25 PCR.I:

50 mM Tris-HCl pH 8.0, 10 mM $MgCl_2$, 50 mM NaCl, 4 mM spermidine, 0.4 μ g/ml BSA, 4 μ l (= 40 U) *SacI*, 4 μ l (= 40 U) *BstEII*, in 100 μ l total volume at 37°C.

PCR.II:

30 10 mM Tris-Acetate pH 7.5, 10 mM $MgAc_2$, 50 mM KAc (1x "One-Phor-All" buffer ex Pharmacia), 4 μ l (= 48 U) *SfiI*, in 50 μ l total volume at 50°C under mineral oil. After overnight digestion, PCR.II-*SfiI* was digested with *EcoRI* (overnight at 37°C) by the addition of 16 μ l H_2O , 30 μ l 10x
35 "One-Phor-All" buffer (Pharmacia) (100 mM Tris-Acetate pH 7.5, 100 mM $MgAc_2$, 500 mM KAc) and 4 μ l (= 40 U) *EcoRI*.

PCR.III:

10 mM Tris-Acetate pH 7.5, 10 mM MgAc₂, 50 mM KAc (1x "One-Phor-All" buffer {Pharmacia}), 4 µl (= 40 U) *NheI*, 4 µl (= 40 U) *SacI*, in 100 µl total volume at 37°C.

5 PCR.IV:

20 mM Tris-Acetate pH 7.5, 20 mM MgAc₂, 100 mM KAc (2x "One-Phor-All" buffer {Pharmacia}), 4 µl (= 40 U) *XhoI*, 4 µl (= 40 U) *EcoRI*, in 100 µl total volume at 37°C.

PCR.V:

10 20 mM Tris-Acetate pH 7.5, 20 mM MgAc₂, 100 mM KAc (2x "One-Phor-All" buffer {Pharmacia}), 4 µl (= 40 U) *SalI*, 4 µl (= 40 U) *EcoRI*, in 100 µl total volume at 37°C.

PCR.VI:

10 mM Tris-Acetate pH 7.5, 10 mM MgAc₂, 50 mM KAc (1x "One-Phor-All" buffer {Pharmacia}), 4 µl (= 48 U) *SfiI*, in 50 µl total volume at 50°C under mineral oil. After overnight digestion, PCR.VI-*SfiI* was digested with *NheI* (overnight at 37°C) by the addition of 41 µl H₂O, 5 µl 10x "One-Phor-All" buffer (Pharmacia) (100 mM Tris-Acetate pH 20 7.5, 100 mM MgAc₂, 500 mM KAc) and 4 µl (= 40 U) *NheI*.

PCR.VII:

50 mM Tris-HCl, pH 8.0, 10 mM MgCl₂, 50 mM NaCl, 4 mM spermidine, 0.4 µg/ml BSA, 4 µl (= 40 U) *NheI*, 4 µl (= 40 U) *BstEII*, in 100 µl total volume at 37°C.

25 PCR.VIII:

20 mM Tris-Acetate pH 7.5, 20 mM MgAc₂, 100 mM KAc (2x "One-Phor-All" buffer {Pharmacia}), 4 µl (= 40 U) *EcoRI*, in 50 µl total volume at 37°C. After overnight digestion, PCR.VIII-*EcoRI* was digested with *XhoI* (overnight at 37°) by 30 the addition of 46 µl H₂O and 4 µl (= 40 U) *XhoI*.

PCR.IX:

25 mM Tris-Acetate, pH 7.8, 100 mM KAc, 10 mM MgAc, 1mM DTT (1x "Multi-Core" buffer {Promega}), 4 mM spermidine, 0.4 µg/ml BSA, 4 µl (= 40 U) *NheI*, 4 µl (= 40 U) *BstEII*, in 100 35 µl total volume at 37°C.

PCR.X:

50 mM Tris-HCl, pH 8.0, 10 mM MgCl₂, 50 mM NaCl, 4 mM spermidine, 0.4 µg/ml BSA, 4 µl (= 40 U) *Pst*I, 4 µl (= 40 U) *Eco*RI, in 100 µl total volume at 37°C.

5

The digested PCR fragments

| | |
|---|--|
| PCR.I- <i>Sac</i> I/ <i>Bst</i> EII, | PCR.II- <i>Sfi</i> I/ <i>Eco</i> RI, |
| PCR.III- <i>Nhe</i> I/ <i>Sac</i> I, | PCR.IV- <i>Xho</i> I/ <i>Eco</i> RI, |
| PCR.V- <i>Sal</i> I/ <i>Eco</i> RI, | PCR.VI- <i>Sfi</i> I/ <i>Nhe</i> I, |
| 10 PCR.VII- <i>Bst</i> EII/ <i>Nhe</i> I, | PCR.VIII- <i>Xho</i> I/ <i>Eco</i> RI, |
| PCR.IX- <i>Bst</i> EII/ <i>Nhe</i> I, and | PCR.X- <i>Pst</i> I/ <i>Eco</i> RI |

were purified on an 1.2% agarose gel using DEAE-membranes NA45 (Schleicher & Schuell) as described by Maniatis et al. The purified fragments were dissolved in H₂O at a
15 concentration of 100-150 ng/µl.

Construction of the pGOSA double head expression vectors.

The construction of pGOSA.E (see Figure 8) involved several cloning steps that produced 4 intermediate constructs

- 20 pGOSA.A to pGOSA.D (see Figure 30-34). The final expression vector pGOSA.E and the oligonucleotides in Table 1 above have been designed to enable most specificities to be cloned into the final pGOSA.E construct (Figure 9). The upstream V_H domain can be replaced by any *Pst*I-*Bst*EII V_H
25 gene fragment obtained with oligonucleotides PCR.51 and PCR.89 (see Table 1 above). The oligonucleotides DBL.3 and DBL.4 (see Table 1 above) were designed to introduce *Sfi*I and *Nhe*I restriction sites in the V_H gene fragments thus allowing cloning of those V_H gene fragments into the *Sfi*I-
30 *Nhe*I sites as the downstream V_H domain. All V_L gene fragments obtained with oligonucleotides PCR.116 and PCR.90 (see Table 1 above) can be cloned into the position of the V_L.3418 gene fragment as a *Sac*I-*Xho*I fragment. A complication here however is the presence of an internal
35 *Sac*I site in the V_H.3418 gene fragment. Oligonucleotides DBL.8 and DBL.9 (see Table 1 above) are designed to allow cloning of V_L gene fragments into the position of the

V_L.4715 gene fragment as a *Sal*I-*Not*I fragment. The pGOSA.E derivatives pGOSA.V, pGOSA.S and pGOSA.T with only one or no linker sequences contain some aberrant restriction sites at the new joining points. The V_HA-V_HB construct without a linker lacks the 5' V_HB *Sfi*I site. The V_HB fragment is cloned into these constructs as a *Bst*EII/*Nhe*I fragment using oligonucleotides DBL.10 or DBL.11 and DBL.4 (see Table 1 above). The V_LB-V_LA construct without a linker lacks the 5' V_LA *Sal*I site. The V_LA fragment is cloned into these constructs as a *Xho*I/*Eco*RI fragment using oligonucleotides DBL.11 and DBL.9 (see Table 1 above).

In the following part of the description the following linkers are mentioned which are also present in the sequence listing:

the (Gly₄Ser)₃ linker, present in SEQ ID NO: 2 as amino acids 109-123 and SEQ ID NO: 10 as amino acids 121-135, the (Gly₄Ser)₃AlaGlySerAla linker (= linkerA), present in SEQ ID NO: 12 as amino acids 121-139, and the (Gly₄Ser)₃Gly₄Val linker (= linkerV), present in SEQ ID NO: 13 as amino acids 108-122.

pGOSA.A

This plasmid is derived from both the Fv.4715-myc construct and the scFv.4715-myc construct.

An *Sfi*I restriction site was introduced between the DNA sequence encoding the (Gly₄Ser)₃ linker and the gene fragment encoding the V_L of the scFv.4715-myc construct (see Figure 30). This was achieved by replacing the *Bst*EII-*Sac*I fragment of the latter construct by the fragment PCR-I *Bst*EII/*Sac*I (Figure 20) that contains an *Sfi*I site between the DNA encoding the (Gly₄Ser)₃ linker and the V_L.4715 gene fragment. The introduction of the *Sfi*I site also introduced 4 additional amino acids (AlaGlySerAla) between the (Gly₄Ser)₃ linker and V_L.4715 resulting in a (Gly₄Ser)₃AlaGlySerAla linker (linkerA). The oligonucleotides used to produce PCR-I (DBL.1 and DBL.2,

see Table 1 above) were designed to match the sequence of the framework-3 region of V_H.4715 and to prime at the junction of the DNA encoding the (Gly,Ser), linker and the V_L.4715 gene fragment, respectively. Thus pGOSA.A can be indicated as:

5 **pelB-V_H4715-linkerA-(SfiI)-V_L4715-myc.**

pGOSA.B

This plasmid is derived from plasmid Fv.3418 (see Figure 31). The XhoI-EcoRI fragment of plasmid Fv.3418 comprising the 3' end of DNA encoding framework-4 of the V_L including the stop codon was removed and replaced by the fragment PCR-IV XhoI/EcoRI (Figure 23). The oligonucleotides used to produce PCR-IV (DBL.6 and DBL.7, see Table 1 above) were designed to match the sequence at the junction of the V_L and the (Gly,Ser), linker perfectly (DBL.6), and to be able to prime at the junction of the (Gly,Ser), linker and the V_H in pUR.4124 (DBL.7). DBL.7 removed the PstI site in the V_H (silent mutation) and introduced a SalI restriction site at the junction of the (Gly,Ser), linker and the V_H, thereby replacing the last Ser of the linker by a Val residue resulting in a (Gly,Ser),Gly,Val linker (linkerV). Thus pGOSA.B can be indicated as:

20

pelB-V_H3418 + pelB-V_L3418-linkerV-(SalI-EcoRI)

pGOSA.C

This plasmid contains DNA encoding V_H.4715 linked by the (Gly,Ser),AlaGlySerAla linker to V_H.3418 (see Figure 32), thus:

pelB-V_H4715-linkerA-V_H3418.

This construct was obtained by replacing the SfiI-EcoRI fragment from pGOSA.A encoding V_L.4715 by the fragment PCR-II SfiI/EcoRI containing the V_H.3418 gene (see Figure 21). The oligonucleotides used to produce PCR-II (DBL.3 and DBL.4, see Table 1 above) hybridize in the framework-1 and framework-4 region of the gene encoding V_H.3418, respectively. DBL.3 was designed to remove the PstI restriction site (silent mutation) and to introduce an SfiI restriction site upstream of the V_H gene. DBL.4 destroys

30

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the *Bst*EII restriction site in the framework-4 region and introduces an *Nhe*I restriction site downstream of the stopcodon.

5 pGOSA.D

This plasmid contains a dicistronic operon comprising the $V_H.3418$ gene and DNA encoding $V_L.3418$ linked by the (Gly,Ser),Gly,Val linker to $V_L.4715$ (see Figure 33), thus:

pelB-V_H3418 + pelB-V_L3418-linkerV-V_L4715.

- 10 This construct was obtained by digesting plasmid pGOSA.B with *Sal*I-*Eco*RI and inserting the fragment PCR-V *Sal*I/*Eco*RI (Figure 24) containing the $V_L.4715$ gene. The oligonucleotides used to obtain PCR-V (DBL.8 and DBL.9, see Table 1 above) were designed to match the nucleotide
- 15 sequence of the framework-1 and framework-4 regions of the $V_L.4715$ gene, respectively. DBL.8 removed the *Sac*I site from the framework-1 region (silent mutation) and introduced a *Sal*I restriction site upstream of the $V_L.4715$ gene. DBL.9 destroyed the *Xho*I restriction site in the framework-4
- 20 region of the $V_L.4715$ gene (silent mutation) and introduced a *Not*I and an *Eco*RI restriction site downstream of the stop codon.

pGOSA.E

- 25 This plasmid contains a dicistronic operon comprising DNA encoding $V_H.4715$ linked by the (Gly,Ser),AlaGlySerAla linker to $V_H.3418$ plus DNA encoding $V_L.3418$ linked by the (Gly,Ser),Gly,Val linker to $V_L.4715$ (see Figure 34), thus:
- pelB-V_H4715-linkerA-V_H3418 + pelB-V_L3418-linkerV-V_L4715.*
- 30 Both translational units are preceded by a ribosome binding site and DNA encoding a *pelB* leader sequence. This plasmid was obtained by a three-point ligation by mixing the vector resulting from pGOSA.D after removal of the $V_H.3418$ -encoding *Pst*I-*Sac*I insert with the *Pst*I-*Nhe*I pGOSA.C insert
- 35 containing $V_H.4715$ linked to $V_H.3418$ and the PCR-III *Nhe*I/*Sac*I fragment (see Figure 22). The remaining *Pst*I-*Sac*I pGOSA.D vector contains the 5' end of the framework-1

region of V_H.3418 upto the *Pst*I restriction site and V_L.3418 linked by the (Gly,Ser)₂Gly,Val linker to V_L.4715 starting from the *Sac*I restriction site in V_L.3418. The *Pst*I-*Nhe*I pGOSA.C insert contains V_H.4715 linked by the (Gly,Ser)₂-
 5 AlaGlySerAla linker to V_H.3418, starting from the *Pst*I restriction site in the framework-1 region in V_H.4715. The *Nhe*I-*Sac*I PCR-III fragment provides the ribosome binding site and DNA encoding the *pelB* leader sequence for the V_L.3418-(Gly,Ser)₂Gly,Val-V_L.4715 construct. The oligo-
 10 nucleotides DBL.5 and PCR.116 (see Table 1 above) used to generate PCR-III were designed to match the sequence upstream of the ribosome binding site of V_L.4715 in Fv.4715 and to introduce an *Nhe*I restriction site (DBL.5), and to match the framework-4 region of V_L.3418 (PCR.116).

15

pGOSA.V

This plasmid is derived from pGOSA.E (see Figure 35) from which the *Bst*EII/*Nhe*I fragment containing DNA encoding linkerA-V_H.3418 was excised and replaced by the fragment
 20 PCR-VII *Bst*EII/*Nhe*I containing the V_H.3418 gene (see Figure 26). The resulting plasmid pGOSA.V contains V_H.3418 linked directly to the framework-4 region of V_H.4715, plus V_L.4715 linked by the (Gly,Ser)₂Gly,Val linker to the framework-4 region of V_L.3418, thus:

25 $pelB-V_H4715*V_H3418 + pelB-V_L3418-linkerV-V_L4715.$

pGOSA.S

This plasmid is derived from pGOSA.E (see Figure 36) from which the (Gly,Ser)₂Gly,Val-V_L.4715 *Xho*I/*Eco*RI fragment was
 30 excised and replaced by the fragment PCR-VIII *Xho*I/*Eco*RI which contains V_L.4715 (see Figure 27). The resulting plasmid pGOSA.S contains V_H.4715 linked by the (Gly,Ser)₂-AlaGlySerAla linker to V_H.3418 plus V_L.3418 linked directly to the 5' end of the framework-1 region of V_L.4715, thus:

35 $pelB-V_H4715-linkerA-V_H3418 + pelB-V_L3418*V_L4715.$

pGOSA.T

This plasmid contains a dicistronic operon consisting of $V_H.3418$ directly to the framework-4 region of $V_H.4715$ plus $V_L.3418$ linked directly to the 5' end of the framework-1 region of $V_L.4715$ (see Figure 37). Both transcriptional units are preceded by a ribosome binding site and a *pelB* leader sequence, thus:

$pelB-V_H.4715*V_H.3418 + pelB-V_L.3418*V_L.4715.$

This construct was obtained by inserting the *NheI/EcoRI* fragment of pGOSA.S which contains $V_L.3418$ linked directly to the 5' end of the framework-1 region of $V_L.4715$, into the vector pGOSA.V from which the *NheI/EcoRI* fragment containing $V_L.3418$ linked by the (Gly,Ser)₂Gly,Val linker to $V_L.4715$ was removed.

EXAMPLE 6. Detailed description of the preparation of other dicistronic constructs pGOSA.G, and pGOSA.J, pGOSA.Z, pGOSA.AA and pGOSA.AB

pGOSA.G

This plasmid is an intermediate for the synthesis of pGOSA.J. It is derived from pGOSA.E from which the $V_H.4715$ *PstI/BstEII* fragment has been excised and replaced by the $V_H.3418$ *PstI/BstEII* fragment (excised from Fv.3418). The resulting plasmid pGOSA.G (see Figure 38) contains two copies of $V_H.3418$ linked by the (Gly,Ser)₂AlaGlySerAla linker, plus $V_L.4715$ linked by the (Gly,Ser)₂Gly,Val linker to the framework-4 region of $V_L.3418$, thus:

$pelB-V_H.3418-linkerA-V_H.3418 + pelB-V_L.3418-linkerV-V_L.4715.$

pGOSA.J

This plasmid contains a dicistronic operon consisting of $V_H.3418$ linked by the (Gly,Ser)₂AlaGlySerAla linker to $V_H.4715$ plus $V_L.3418$ linked by the (Gly,Ser)₂Gly,Val linker to $V_L.4715$. Both transcriptional units are preceded by a

ribosome binding site and a *pelB* leader sequence (see Figure 39), thus:

pelB-V_H.3418-linkerA-V_H.4715 + pelB-V_L.3418-linkerV-V_L.4715.

- 5 This construct was obtained by inserting the fragment PCR-VI *SfiI/NheI* which contains V_H4715 (Figure 25), into the vector pGOSA.G from which the *SfiI/NheI* V_H3418 fragment was removed.

10 **pGOSA.Z**

This plasmid is derived from pGOSA.G from which the (Gly₄Ser)₃AlaGlySerAla linker-V_H3418 *BstEII/NheI* fragment was excised and replaced by the fragment PCR-IX *BstEII/NheI* which contains V_H.4715 (Figure 28). The resulting plasmid

- 15 pGOSA.Z (see Figure 40) contains V_H.3418 linked directly to the framework-1 region of V_H.4715, plus V_L.4715 linked by the (Gly₄Ser)₂Gly₄Val linker to the framework-4 region of V_L.3418, thus:

*pelB-V_H.3418*V_H.4715 + pelB-V_L.3418-linkerV-V_L.4715.*

20

pGOSA.AA

This plasmid contains a dicistronic operon consisting of the V_H.3418 linked directly to the 5' end of the framework-1 region of V_H.4715 plus V_L.3418 linked directly to the 5'

- 25 end of the framework-1 region of V_L.4715. Both transcriptional units are preceded by a ribosome binding site and a *pelB* leader sequence (see Figure 41). This construct was obtained by inserting the *NheI/EcoRI* fragment of pGOSA.T which contains V_L.3418 linked directly to the 5' end of the framework-1 region of V_L.4715, into the vector pGOSA.Z from which the *NheI/EcoRI* fragment containing V_L.3418 linked by the (Gly₄Ser)₂Gly₄Val linker to V_L.4715 was removed, thus:

*pelB-V_H.3418*V_H.4715 + pelB-V_L.3418*V_L.4715.*

35

pGOSA.AB

This plasmid is derived from pGOSA.J by a three point ligation reaction (see Figure 42). The *SacI*/*EcoRI* insert, containing part of *V_H*.3418 and the full

- 5 (Gly,Ser),AlaGlySerAla linker-*V_H*.4715 and the *V_L*.3418-
(Gly,Ser),Gly,Val-*V_L*.4715 encoding sequences, was removed and replaced by the *SacI*/*SacI* pGOSA.J fragment containing the same part of *V_H*.3418 and the full (Gly,Ser),AlaGlySerAla linker-*V_H*.4715 and the *SacI*/*EcoRI* pGOSA.T fragment
10 containing *V_L*.3418 linked directly to the framework-1 region of *V_L*.4715 (see Figure 37). The resulting plasmid contains *V_H*.3418 linked by the (Gly,Ser),AlaGlySerAla linker to the 5' end of the framework-1 region of *V_H*.4715 plus *V_L*.3418 linked directly to the 5' end of the framework-1 region of
15 *V_L*.4715, thus:

pelB-V_H.3418-linkerA-*V_H*.4715 + *pelB-V_L*.3418**V_L*.4715.

- 20 **EXAMPLE 7.** Detailed description of the preparation of monocistronic constructs pGOSA.L and pGOSA.Y, and pGOSA.C, pGOSA.X, pGOSA.AC and pGOSA.AD

pGOSA.L

- This plasmid is derived from pGOSA.E from which the
25 *HindIII*/*NheI* fragment containing DNA encoding *V_H*.4715-
(Gly,Ser),AlaGlySerAla-*V_H*.3418 was removed (see Figure 43). The DNA ends of the vector were made blunt-end using Klenow DNA polymerase and ligated. The resulting plasmid pGOSA.L contains *V_L*.3418 linked by the (Gly,Ser),Gly,Val linker to
30 the 5' end of the framework-1 region of *V_L*.4715, thus:

pelB-V_L.3418-linkerV-*V_L*.4715.

pGOSA.Y

- This plasmid is derived from pGOSA.T from which the
35 *HindIII*/*NheI* fragment containing DNA encoding *V_H*.4715-
V_H.3418 was removed (see Figure 44). The DNA ends of the vector were made blunt-end using Klenow DNA polymerase and

ligated. The resulting plasmid pGOSA.Y contains $V_L.3418$ linked directly to 5' end of the framework-1 region of $V_L.4715$, thus:

$pelB-V_L.3418*V_L.4715.$

5

The preparation of pGOSA.C was given in Example 5 above; it can be indicated with: $pelB-V_H.4715-linkerA-V_H.3418.$

pGOSA.X

- 10 This plasmid is derived from pGOSA.T from which the *NheI/EcoRI* fragment containing DNA encoding $V_L.3418-V_L.4715$ was removed. The DNA ends of the vector were made blunt-end using Klenow DNA polymerase and ligated. The resulting plasmid pGOSA.X (see Figure 45) contains $V_H.4715$ linked
15 directly to 5' end of the framework-1 region of $V_H.3418$, thus:

$pelB-V_H.4715*V_H.3418.$

pGOSA.AC

- This plasmid is derived from pGOSA.Z from which the
20 *NheI/EcoRI* fragment containing DNA encoding $V_L.3418-(Gly,Ser),Gly,Val-V_L.4715$ was removed (see Figure 46). The DNA ends of the vector were made blunt-end using Klenow DNA polymerase and ligated. The resulting plasmid pGOSA.AC contains $V_H.3418$ linked directly to 5' end of the
25 framework-1 region of $V_H.4715$, thus:

$pelB-V_H.3418*V_H.4715.$

pGOSA.AD

- This plasmid was obtained by inserting the *PstI/EcoRI* PCR.X
30 fragment containing DNA encoding $V_H.3418-(Gly,Ser),AlaGly-SerAla-V_H.4715$ (see Figure 29) into the Fv.4715-myc vector from which the *PstI/EcoRI* Fv.4715-myc insert was removed (see Figure 47), thus: $pelB-V_H.3418-linkerA-V_H.4715.$

- 35 These monocistronic constructs can be used to transform the same host with two different plasmids or to transform two

different hosts, so that the two V_H 's in series can be produced separately from the two V_L 's in series.

Evaluation of the results obtained

5 Bifunctional binding activity of GOSA double heads

In this specification the construction of a two chain protein complex is described, in which one of the chains consists of two heavy chain V-domains and the other chain consists of the two corresponding light chain V-domains.

10 The variable domains are linked either directly or through a polypeptide linker. In this specification evidence is provided that these type of molecules ("double heads") contain both antigen binding specificities of the Fv's used to generate these multi-functional antibody fragments.

15 Figure 12 shows that GOSA.E can be used to specifically target the enzyme glucose oxidase to several *Streptococcus sanguis* strains, using antibody fragments derived from hybridomas expressing antibodies directed against these antigens. Figure 12 further shows that the fine specificity
20 of the anti-*Streptococcus sanguis* scFv 4715 is preserved in the GOSA.E double head.

Effect of linkers and relative position of V-domains on double head activity

25 After it was shown that the "cross-over double-head" approach ($V_{HA}-V_{HB} + V_{LB}-V_{LA}$) yields active bispecific molecules, the importance of the relative position of the V-domains in these constructs was investigated. Both possible positional orientations ($GOSA.E = V_{HA}-LinkerA-V_{HB} +$
30 $V_{LB}-LinkerV-V_{LA}$ and $GOSA.J = V_{HB}-LinkerA-V_{HA} + V_{LB}-LinkerV-V_{LA}$) were constructed and tested for bispecific activity, despite the suggestion obtained by molecular modelling that the binding site formed by the second (downstream/C-terminal) V-domains in the configuration $V_{HB}-V_{HA} + V_{LB}-V_{LA}$
35 ($GOSA.J$) was in an unfavourable position for binding to large protein antigens on the surface of cells. Surprisingly however, it was found experimentally that the

- downstream binding site is in fact accessible. Although the relative position of the heavy chains and the light chains was found to have an effect on the observed reactivity both tested combinations show bispecific activity with the
- 5 "cross-over" combination ($GOSA.E = V_HA-V_HB + V_LB-V_LA$) exhibiting a higher level of reactivity compared to the combination $V_HB-V_HA + V_LB-V_LA$ ($= GOSA.J$) as demonstrated for A=anti-Strep and B=anti-Gox.
- 10 Molecular modelling of the $V_HB-V_HA + V_LB-V_LA$ ($= GOSA.J$) configuration further suggested that, only when the connecting linkers are kept long enough (to span 30 to 35 Å), the protein chains could fold such that both binding sites are fully accessible.
- 15 The "cross-over" configuration: $V_HA-V_HB + V_LB-V_LA$ ($GOSA.E$) wherein linker length was not critical, was predicted to result in a complex with both binding sites facing in opposite directions, without the restraints suggested for the configuration $V_HB-V_HA + V_LB-V_LA$ ($GOSA.J$).
- 20 Removing the flexible polypeptide linker from the V_HA-V_HB chain only had a minimal effect on the ability of the double head in the "cross-over" configuration ($GOSA.V = V_HA*V_HB + V_LB-V_LA$) to bind both *S. sanguis* and Glucose oxidase. However, removing the flexible polypeptide linker
- 25 from the V_HB-V_HA chain from the molecule in the $V_HB-V_HA + V_LB-V_LA$ configuration ($GOSA.Z = V_HB*V_HA + V_LB-V_LA$) resulted in a dramatic reduction of its ability to bind both *S. sanguis* and Glucose oxidase.
- In contrast with the double head in the "cross-over"
- 30 configuration without the flexible polypeptide linker between the two heavy chain domains ($GOSA.V$), where molecular modelling predicted the resulting molecule to be active, removal of the flexible linker from the V_LB-V_LA chain could not be modelled such that both binding sites
- 35 were fully accessible. ELISA results confirm that the double head in the $V_HB-V_HA + V_LB-V_LA$ configuration without a linker between the two light chain domains ($GOSA.AB$)

exhibits only minimal *S. sanguis* and glucose oxidase binding activity. Surprisingly, deletion of the flexible linker from the V_LB-V_LA chain from the double head in the "cross-over" configuration (GOSA.S) only had a small effect on the bispecific activity of the resulting molecule. As expected from the molecular modelling results from the double heads without a flexible linker between the two light chain domains, removal of both the flexible polypeptide linkers from the double head molecules, could not be modelled such that both binding sites were fully accessible. In agreement with the ELISA results obtained with the GOSA.AB construct, the double head in the $V_{HB}-V_{HA} + V_LB-V_LA$ configuration without any linkers (GOSA.AA) only exhibits minimal if any *S. sanguis* and glucose oxidase binding activity. Surprisingly, the double head in the "cross-over" configuration without any linkers ($GOSA.T = V_{HA}*V_{HB} + V_LB*V_LA$) still exhibited 25-50% of *S. sanguis* and glucose oxidase bispecific binding activity when compared to the activity of the double head in the "cross-over" configuration with two linkers (GOSA.E). Thus the conclusion of this work is that modelling can give some indications, but that the computer programmes cannot predict what is possible and what not. Several deviations from the modelling expectations were found. With a paraphrase on an old saying: theories are nice but the experiment is the ultimate proof.

Sensitivity of GOSA double heads

Using an ELISA format it was shown that the sensitivity of the GOSA.E double head is as least as sensitive as an IgG-glucose oxidase conjugate, as determined by the lowest concentration of *Streptococcus sanguis* antigen immobilised on a solid phase that is still detectable.

GOSA double heads are produced as dimers

FPLC analysis of partially affinity-purified GOSA.E, GOSA.V, GOSA.S and GOSA.T samples usually gave only one

GOSA double head activity peak as determined by ELISA (Figures 16-19). The position of this peak in the elution pattern indicated that the molecular weight of the GOSA double head is 40-50 kD. Since this molecular weight corresponds to the expected molecular weight of the $V_H2 + V_L2$ double head dimer, it was concluded that GOSA.E, GOSA.V, GOSA.S and GOSA.T are primarily produced as dimeric molecules. Occasionally an activity peak with an apparent molecular weight of ~200 kD was observed (Figure 16). The presence of glucose oxidase activity in these fractions indicate that these fractions contain GOSA double head complexed with glucose oxidase.

In vitro assembly of GOSA double heads

It was shown that bifunctionally active dimeric GOSA molecules together in one cell can be produced by translation from one dicistronic messenger (GOSA.E, GOSA.S, GOSA.T, GOSA.V, GOSA.J, GOSA.AB, GOSA.AA and GOSA.Z). In addition high levels of *S. sanguis* and glucose oxidase bispecific binding activity is formed when supernatants of cultures producing the separate GOSA subunits are mixed (see Example 7). The effects of linkers and the relative position of the individual V_H -domains on the *S. sanguis* and glucose oxidase bispecific binding activity observed in these mixing experiments are comparable to the dicistronic constructs.

The constructs described above are summarised in Table 2 below.

Table 2A describes intermediate constructs that were not further tested.

Table 2B describes the dicistronic constructs.

Table 2C describes the monocistronic constructs.

(LiA) stands for the V_H - V_H linker (Gly,Ser)₂AlaGlySerAla (= linkerA)

(LiV) stands for the V_L - V_L linker (Gly,Ser)₂Gly,Val (= linkerV)

(*) indicates that the two heavy chain domains or the two light chain domains are fused together without a connecting linker.

5

Table 2.

Table 2A

| | | |
|----|----------|---|
| | GOSA.A : | V _H .4715-LiA-(SfiI)-V _L .4715-myc |
| 10 | GOSA.B : | V _H .3418-LiV-V _L .3418-(SalI/EcoRI) |
| | GOSA.D : | V _H .3418 + V _L .3418-LiV-V _L .4715 |
| | GOSA.G : | V _H .3418-LiA-V _H .3418 + V _L .3418-LiV-V _L .4715 |

Table 2B

| | | |
|----|----------|---|
| 15 | GOSA.E : | V _H .4715-LiA-V _H .3418 + V _L .3418-LiV-V _L .4715 |
| | GOSA.S : | V _H .4715-LiA-V _H .3418 + V _L .3418*V _L .4715 |
| | GOSA.T : | V _H .4715*V _H .3418 + V _L .3418*V _L .4715 |
| | GOSA.V : | V _H .4715*V _H .3418 + V _L .3418-LiV-V _L .4715 |
| 20 | GOSA.J : | V _H .3418-LiA-V _H .4715 + V _L .3418-LiV-V _L .4715 |
| | GOSA.AB: | V _H .3418-LiA-V _H .4715 + V _L .3418*V _L .4715 |
| | GOSA.AA: | V _H .3418*V _H .4715 + V _L .3418*V _L .4715 |
| | GOSA.Z : | V _H .3418*V _H .4715 + V _L .3418-LiV-V _L .4715 |

25 Table 2C

| | | |
|----|----------|---|
| | GOSA.L : | V _L .3418-LiV-V _L .4715 |
| | GOSA.Y : | V _L .3418*V _L .4715 |
| | GOSA.AD: | V _H .3418-LiA-V _H .4715 |
| 30 | GOSA.AC: | V _H .3418*V _H .4715 |
| | GOSA.C : | V _H .4715-LiA-V _H .3418 |
| | GOSA.X : | V _H .4715*V _H .3418 |

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(B) COMPUTER: IBM PC compatible
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(D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(v) CURRENT APPLICATION DATA:

APPLICATION NUMBER: EP 95307332.7

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 737 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "cDNA domains with synthetic linker(s)"

(vii) IMMEDIATE SOURCE:

(B) CLONE: EcoRI-HindIII insert of pUR4124

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION:11..730

(D) OTHER INFORMATION:/product= "VLlys-GS-VHlys"

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION:11..334

(D) OTHER INFORMATION:/product= "VLlys"

(ix) FEATURE:

(A) NAME/KEY: misc_RNA

(B) LOCATION:335..379

(D) OTHER INFORMATION:/product= "(Gly4Ser)3 linker"

(ix) FEATURE:

(A) NAME/KEY: mat_peptide

(B) LOCATION:380..727

(D) OTHER INFORMATION:/product= "VHlys"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

| | |
|---|-----|
| GAATTCGGCC GAC ATC GAG CTC ACC CAG TCT CCA GCC TCC CTT TCT GCG | 49 |
| Asp Ile Glu Leu Thr Gln Ser Pro Ala Ser Leu Ser Ala | |
| 1 5 10 | |
| TCT GTG GGA GAA ACT GTC ACC ATC ACA TGT CGA GCA AGT GGG AAT ATT | 97 |
| Ser Val Gly Glu Thr Val Thr Ile Thr Cys Arg Ala Ser Gly Asn Ile | |
| 15 20 25 | |
| CAC AAT TAT TTA GCA TGG TAT CAG CAG AAA CAG GGA AAA TCT CCT CAG | 145 |
| His Asn Tyr Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys Ser Pro Gln | |
| 30 35 40 45 | |
| CTC CTG GTC TAT TAT ACA ACA ACC TTA GCA GAT GGT GTG CCA TCA AGG | 193 |
| Leu Leu Val Tyr Tyr Thr Thr Thr Leu Ala Asp Gly Val Pro Ser Arg | |
| 50 55 60 | |
| TTC AGT GGC AGT GGA TCA GGA ACA CAA TAT TCT CTC AAG ATC AAC AGC | 241 |
| Phe Ser Gly Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys Ile Asn Ser | |
| 65 70 75 | |
| CTG CAA CCT GAA GAT TTT GGG AGT TAT TAC TGT CAA CAT TTT TGG AGT | 289 |
| Leu Gln Pro Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His Phe Trp Ser | |
| 80 85 90 | |
| ACT CCT CGG ACG TTC GGT GGA GGG ACC AAG CTC GAG ATC AAA CGG GGT | 337 |
| Thr Pro Arg Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Gly | |
| 95 100 105 | |
| GGA GGC GGT TCA GGC GGA GGT GGC TCT GGC GGT GGC GGA TCG CAG GTG | 385 |
| Gly Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser Gly Ser Gln Val | |
| 110 115 120 125 | |
| CAG CTG CAG GAG TCA GGA CCT GGC CTG GTG GCG CCC TCA CAG AGC CTG | 433 |
| Gln Leu Gln Glu Ser Gly Pro Gly Leu Val Ala Pro Ser Gln Ser Leu | |
| 130 135 140 | |
| TCC ATC ACA TGC ACC GTC TCA GGG TTC TCA TTA ACC GGC TAT GGT GTA | 481 |
| Ser Ile Thr Cys Thr Val Ser Gly Phe Ser Leu Thr Gly Tyr Gly Val | |
| 145 150 155 | |

43

| | |
|---|-----|
| AAC TGG GTT CGC CAG CCT CCA GGA AAG GGT CTG GAG TGG CTG GGA ATG Asn Trp Val Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Leu Gly Met 160 165 170 | 529 |
| ATT TGG GGT GAT GGA AAC ACA GAC TAT AAT TCA GCT CTC AAA TCC AGA Ile Trp Gly Asp Gly Asn Thr Asp Tyr Asn Ser Ala Leu Lys Ser Arg 175 180 185 | 577 |
| CTG AGC ATC AGC AAG GAC AAC TCC AAG AGC CAA GTT TTC TTA AAA ATG Leu Ser Ile Ser Lys Asp Asn Ser Lys Ser Gln Val Phe Leu Lys Met 190 195 200 205 | 625 |
| AAC AGT CTG CAC ACT GAT GAC ACA GCC AGG TAC TAC TGT GCC AGA GAG Asn Ser Leu His Thr Asp Asp Thr Ala Arg Tyr Tyr Cys Ala Arg Glu 210 215 220 | 673 |
| AGA GAT TAT AGG CTT GAC TAC TGG GGC CAA GGG ACC ACG GTC ACC GTC Arg Asp Tyr Arg Leu Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val 225 230 235 | 721 |
| TCC TCA TGA TAAGCTT Ser Ser * 240 | 737 |

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 240 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Asp Ile Glu Leu Thr Gln Ser Pro Ala Ser Leu Ser Ala Ser Val Gly
 1 5 10 15
 Glu Thr Val Thr Ile Thr Cys Arg Ala Ser Gly Asn Ile His Asn Tyr
 20 25 30
 Leu Ala Trp Tyr Gln Gln Lys Gln Gly Lys Ser Pro Gln Leu Leu Val
 35 40 45
 Tyr Tyr Thr Thr Thr Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly
 50 55 60
 Ser Gly Ser Gly Thr Gln Tyr Ser Leu Lys Ile Asn Ser Leu Gln Pro
 65 70 75 80
 Glu Asp Phe Gly Ser Tyr Tyr Cys Gln His Phe Trp Ser Thr Pro Arg
 85 90 95
 Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Gly Gly Gly Gly
 100 105 110
 Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gln Val Gln Leu Gln
 115 120 125
 Glu Ser Gly Pro Gly Leu Val Ala Pro Ser Gln Ser Leu Ser Ile Thr
 130 135 140
 Cys Thr Val Ser Gly Phe Ser Leu Thr Gly Tyr Gly Val Asn Trp Val
 145 150 155 160

44

```

Arg Gln Pro Pro Gly Lys Gly Leu Glu Trp Leu Gly Met Ile Trp Gly
      165                      170                      175
Asp Gly Asn Thr Asp Tyr Asn Ser Ala Leu Lys Ser Arg Leu Ser Ile
      180                      185                      190
Ser Lys Asp Asn Ser Lys Ser Gln Val Phe Leu Lys Met Asn Ser Leu
      195                      200                      205
His Thr Asp Asp Thr Ala Arg Tyr Tyr Cys Ala Arg Glu Arg Asp Tyr
      210                      215                      220
Arg Leu Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser *
      225                      230                      235                      240

```

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 920 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "cDNA domains with synthetic linker(s)"

(vii) IMMEDIATE SOURCE:

- (B) CLONE: HindIII-EcoRI insert Fv.3418

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:36..443
- (D) OTHER INFORMATION:/product= "pelB-VH3418"

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide
- (B) LOCATION:36..101
- (D) OTHER INFORMATION:/product= "pectate lyase"

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION:102..440
- (D) OTHER INFORMATION:/product= "VH3418"

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION:495..884
- (D) OTHER INFORMATION:/product= "pelB-VL4318"

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide
- (B) LOCATION:495..560
- (D) OTHER INFORMATION:/product= "pectate lyase"

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION:561..881
- (D) OTHER INFORMATION:/product= "VL3418"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

```

AAGCTTGCAA ATTCTATTTC AAGGAGACAG TCATA ATG AAA TAC CTA TTG CCT
                        Met Lys Tyr Leu Leu Pro
                        -22      -20

```

53

45

| | |
|---|-----|
| ACG GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCG ATG GCC Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro Ala Met Ala -15 -10 -5 | 101 |
| CAG GTG CAG CTG CAG CAG TCA GGA CCT GAG CTG GTA AAG CCT GGG GCT Gln Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala 1 5 10 15 | 149 |
| TCA GTG AAG ATG TCC TGC AAG GCT TCT GGA TAC ACA TTC ACT AGC TAT Ser Val Lys Met Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr 20 25 30 | 197 |
| GTT ATG CAC TGG GTG AAA CAG AAG CCT GGG CAG GGC CTT GAG TGG ATT Val Met His Trp Val Lys Gln Lys Pro Gly Gln Gly Leu Glu Trp Ile 35 40 45 | 245 |
| GGA TAT ATT TAT CCT TAC AAT GAT GGT ACT AAG TAC AAT GAG AAG TTC Gly Tyr Ile Tyr Pro Tyr Asn Asp Gly Thr Lys Tyr Asn Glu Lys Phe 50 55 60 | 293 |
| AAA GGC AAG GCC ACA CTG ACT TCA GAC AAA TCC TCC AGC ACA GCC TAC Lys Gly Lys Ala Thr Leu Thr Ser Asp Lys Ser Ser Ser Thr Ala Tyr 65 70 75 80 | 341 |
| ATG GAG CTC AGC AGC CTG ACC TCT GAG GAC TCT GCG GTC TAT TAC TGT Met Glu Leu Ser Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys 85 90 95 | 389 |
| TCA AGA CGC TTT GAC TAC TGG GGC CAA GGG ACC ACG GTC ACC GTC TCC Ser Arg Arg Phe Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser 100 105 110 | 437 |
| TCA TAA TAAGAGCTAT GGGAGCTTGC ATGCAAATTC TATTTCAAGG AGACAGTCAT Ser * | 493 |
| A ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu -22 -20 -15 -10 | 539 |
| GCT GCC CAA CCA GCG ATG GCC GAC ATC GAG CTC ACC CAG TCT CCA TCT Ala Ala Gln Pro Ala Met Ala Asp Ile Glu Leu Thr Gln Ser Pro Ser -5 1 5 | 587 |
| TCC ATG TAT GCA TCT CTA GGA GAG AGA ATC ACT ATC ACT TGC AAG GCG Ser Met Tyr Ala Ser Leu Gly Glu Arg Ile Thr Ile Thr Cys Lys Ala 10 15 20 25 | 635 |
| AGT CAG GAC ATT AAT ACC TAT TTA ACC TGG TTC CAG CAG AAA CCA GGG Ser Gln Asp Ile Asn Thr Tyr Leu Thr Trp Phe Gln Gln Lys Pro Gly 30 35 40 | 683 |
| AAA TCT CCC AAG ACC CTG ATC TAT CGT GCA AAC AGA TTG CTA GAT GGG Lys Ser Pro Lys Thr Leu Ile Tyr Arg Ala Asn Arg Leu Leu Asp Gly 45 50 55 | 731 |
| GTC CCA TCA AGG TTC AGT GGC AGT GGA TCT GGG CAA GAT TAT TCT CTC Val Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Gln Asp Tyr Ser Leu 60 65 70 | 779 |
| ACC ATC AGC AGC CTG GAC TAT GAA GAT ATG GGA ATT TAT TAT TGT CTA Thr Ile Ser Ser Leu Asp Tyr Glu Asp Met Gly Ile Tyr Tyr Cys Leu 75 80 85 | 827 |
| CAA TAT GAT GAG TTG TAC ACG TTC GGA GGG GGG ACC AAG CTC GAG ATC Gln Tyr Asp Glu Leu Tyr Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile 90 95 100 105 | 875 |

920

(2) INFORMATION FOR SEO ID NO: 4:

(i) **SEQUENCE CHARACTERISTICS:**

- SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 136 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

```

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala
-22      -20      -15      -10
Ala Gln Pro Ala Met Ala Gln Val Gln Leu Gln Gln Ser Gly Pro Glu
-5              1              5              10
Leu Val Lys Pro Gly Ala Ser Val Lys Met Ser Cys Lys Ala Ser Gly
15              20              25
Tyr Thr Phe Thr Ser Tyr Val Met His Trp Val Lys Gln Lys Pro Gly
30              35              40
Gln Gly Leu Glu Trp Ile Gly Tyr Ile Tyr Pro Tyr Asn Asp Gly Thr
45              50              55
Lys Tyr Asn Glu Lys Phe Lys Gly Lys Ala Thr Leu Thr Ser Asp Lys
60              65              70
Ser Ser Ser Thr Ala Tyr Met Glu Leu Ser Ser Leu Thr Ser Glu Asp
75              80              85              90
Ser Ala Val Tyr Tyr Cys Ser Arg Arg Phe Asp Tyr Trp Gly Gln Gly
95              100              105
Thr Thr Val Thr Val Ser Ser *
110

```

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 130 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala
-22 -20 -15 -10

Ala Gln Pro Ala Met Ala Asp Ile Glu Leu Thr Gln Ser Pro Ser Ser
-5 1 5 10

Met Tyr Ala Ser Leu Gly Glu Arg Ile Thr Ile Thr Cys Lys Ala Ser
15 20 25

Gln Asp Ile Asn Thr Tyr Leu Thr Trp Phe Gln Gln Lys Pro Gly Lys
30 35 40

47

Ser Pro Lys Thr Leu Ile Tyr Arg Ala Asn Arg Leu Leu Asp Gly Val
 45 50 55
 Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Gln Asp Tyr Ser Leu Thr
 60 65 70
 Ile Ser Ser Leu Asp Tyr Glu Asp Met Gly Ile Tyr Tyr Cys Leu Gln
 75 80 85 90
 Tyr Asp Glu Leu Tyr Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys
 95 100 105
 Arg *

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 999 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "cdna domains with synthetic linker(s)"
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: HindIII-EcoRI insert of Fv.4715-myc
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION:40..468
 - (D) OTHER INFORMATION:/product= "pelB-VH4715"
- (ix) FEATURE:
 - (A) NAME/KEY: sig_peptide
 - (B) LOCATION:40..105
 - (D) OTHER INFORMATION:/product= "pectate lyase"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION:106..465
 - (D) OTHER INFORMATION:/product= "VH4715"
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION:520..963
 - (D) OTHER INFORMATION:/product= "pelB-VL4715-myc"
- (ix) FEATURE:
 - (A) NAME/KEY: sig_peptide
 - (B) LOCATION:520..585
 - (D) OTHER INFORMATION:/product= "pectate lyase"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION:586..927
 - (D) OTHER INFORMATION:/product= "VL4715"
- (ix) FEATURE:
 - (A) NAME/KEY: misc_RNA
 - (B) LOCATION:928..960
 - (D) OTHER INFORMATION:/product= "myc-tag"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

| | |
|---|-----|
| AAGCTTGCAT GCAAATTCTA TTTCAAGGAG ACAGTCATA ATG AAA TAC CTA TTG | 54 |
| Met Lys Tyr Leu Leu | |
| -22 -20 | |
| CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCG ATG | 102 |
| Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala Ala Gln Pro Ala Met | |
| -15 -10 -5 | |
| GCC CAG GTG CAG CTG CAG GAG TCA GGG GGA GAC TTA GTG AAG CCT GGA | 150 |
| Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp Leu Val Lys Pro Gly | |
| 1 5 10 15 | |
| GGG TCC CTG ACA CTC TCC TGT GCA ACC TCT GGA TTC ACT TTC AGT AGT | 198 |
| Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly Phe Thr Phe Ser Ser | |
| 20 25 30 | |
| TAT GCC TTT TCT TGG GTC CGC CAG ACC TCA GAC AAG AGT CTG GAG TGG | 246 |
| Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp Lys Ser Leu Glu Trp | |
| 35 40 45 | |
| GTC GCA ACC ATC AGT AGT ACT GAT ACT TAT ACC TAT TAT TCA GAC AAT | 294 |
| Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr Tyr Ser Asp Asn | |
| 50 55 60 | |
| GTG AAG GGG CGC TTC ACC ATC TCC AGA GAC AAT GGC AAG AAC ACC CTG | 342 |
| Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Gly Lys Asn Thr Leu | |
| 65 70 75 | |
| TAC CTG CAA ATG AGC AGT CTG AAG TCT GAG GAC ACA GCC GTG TAT TAC | 390 |
| Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp Thr Ala Val Tyr Tyr | |
| 80 85 90 95 | |
| TGT GCA AGA CAT GGG TAC TAT GGT AAA GGC TAT TTT GAC TAC TGG GGC | 438 |
| Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr Phe Asp Tyr Trp Gly | |
| 100 105 110 | |
| CAA GGG ACC ACG GTC ACC GTC TCC TCA TAA TAAGAGCTAT GGGAGCTTGC | 488 |
| Gln Gly Thr Thr Val Thr Val Ser Ser * | |
| 115 120 | |
| ATGCAAATTC TATTTCAAGG AGACAGTCAT A ATG AAA TAC CTA TTG CCT ACG | 540 |
| Met Lys Tyr Leu Leu Pro Thr | |
| -22 -20 | |
| GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCG ATG GCC GAC | 588 |
| Ala Ala Ala Gly Leu Leu Leu Ala Ala Gln Pro Ala Met Ala Asp | |
| -15 -10 -5 1 | |
| ATC GAG CTC ACT CAG TCT CCA TTC TCC CTG ACT GTG ACA GCA GGA GAG | 636 |
| Ile Glu Leu Thr Gln Ser Pro Phe Ser Leu Thr Val Thr Ala Gly Glu | |
| 5 10 15 | |
| AAG GTC ACT ATG AAT TGC AAG TCC GGT CAG AGT CTG TTA AAC AGT GTA | 684 |
| Lys Val Thr Met Asn Cys Lys Ser Gly Gln Ser Leu Leu Asn Ser Val | |
| 20 25 30 | |
| AAT CAG AGG AAC TAC TTG ACC TGG TAC CAG CAG AAG CCA GGG CAG CCT | 732 |
| Asn Gln Arg Asn Tyr Leu Thr Trp Tyr Gln Gln Lys Pro Gly Gln Pro | |
| 35 40 45 | |
| CCT AAA CTG TTG ATC TAC TGG GCA TCC ACT AGG GAA TCT GGA GTC CCT | 780 |
| Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro | |
| 50 55 60 65 | |

49

| | |
|--|-----|
| GAT CGC TTC ACA GCC AGT GGA TCT GGA ACA GAT TTC ACT CTC ACC ATC Asp Arg Phe Thr Ala Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile | 828 |
| 70 75 80 | |
| AGC AGT GTG CAG GCT GAA GAC CTG GCA GTT TAT TAC TGT CAG AAT GAT Ser Ser Val Gln Ala Glu Asp Leu Ala Val Tyr Tyr Cys Gln Asn Asp | 876 |
| 85 90 95 | |
| TAT ACT TAT CCG TTC ACG TTC GGA GGG GGG ACC AAG CTC GAG ATC AAA Tyr Thr Tyr Pro Phe Thr Phe Gly Gly Thr Lys Leu Glu Ile Lys | 924 |
| 100 105 110 | |
| CGG GAA CAA AAA CTC ATC TCA GAA GAG GAT CTG AAT TAA TAAGATCAAA Arg Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn * | 973 |
| 115 120 125 | |
| CGGTAATAAG GATCCAGCTC GAATTC | 999 |

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 143 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

| | |
|---|-----------------|
| Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala | -22 -20 -15 -10 |
| Ala Gln Pro Ala Met Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp | -5 1 5 10 |
| Leu Val Lys Pro Gly Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly | 15 20 25 |
| Phe Thr Phe Ser Ser Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp | 30 35 40 |
| Lys Ser Leu Glu Trp Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr | 45 50 55 |
| Tyr Tyr Ser Asp Asn Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn | 60 65 70 |
| Gly Lys Asn Thr Leu Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp | 75 80 85 90 |
| Thr Ala Val Tyr Tyr Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr | 95 100 105 |
| Phe Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser * | 110 115 120 |

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 148 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

```

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
-22      -20              -15              -10

Ala Gln Pro Ala Met Ala Asp Ile Glu Leu Thr Gln Ser Pro Phe Ser
-5              1              5              10

Leu Thr Val Thr Ala Gly Glu Lys Val Thr Met Asn Cys Lys Ser Gly
              15              20              25

Gln Ser Leu Leu Asn Ser Val Asn Gln Arg Asn Tyr Leu Thr Trp Tyr
              30              35              40

Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser
              45              50              55

Thr Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Ala Ser Gly Ser Gly
              60              65              70

Thr Asp Phe Thr Leu Thr Ile Ser Ser Val Gln Ala Glu Asp Leu Ala
75              80              85              90

Val Tyr Tyr Cys Gln Asn Asp Tyr Thr Tyr Pro Phe Thr Phe Gly Gly
              95              100              105

Gly Thr Lys Leu Glu Ile Lys Arg Glu Gln Lys Leu Ile Ser Glu Glu
              110              115              120

Asp Leu Asn *
              125

```

(2) INFORMATION FOR SEQ ID NO: 9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 924 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "cDNA domains with synthetic linker(s)"
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: HindIII-EcoRI insert of scFv.4715-myc
- (ix) FEATURE:
 - (A) NAME/KEY: sig_peptide
 - (B) LOCATION:40..105
 - (D) OTHER INFORMATION:/product= "pectate lyase"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION:106..465
 - (D) OTHER INFORMATION:/product= "VH4715"
- (ix) FEATURE:
 - (A) NAME/KEY: misc RNA
 - (B) LOCATION:466..510
 - (D) OTHER INFORMATION:/product= "(Gly4Ser)3-linker"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION:511..852
 - (D) OTHER INFORMATION:/product= "VL4715"

(ix) FEATURE:

- (A) NAME/KEY: misc RNA
 (B) LOCATION:853..885
 (D) OTHER INFORMATION:/product= "myc-tag"

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION:40..888
 (D) OTHER INFORMATION:/product=
 "pelB-VH4715-(Gly4Ser)3-VL4715-myc"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

| | |
|---|-----|
| AAGCTTGCAT GCAAATTCTA TTTCAAGGAG ACAGTCATA ATG AAA TAC CTA TTG | 54 |
| Met Lys Tyr Leu Leu | |
| -22 -20 | |
| CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCG ATG | 102 |
| Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro Ala Met | |
| -15 -10 -5 | |
| GCC CAG GTG CAG CTG CAG GAG TCA GGG GGA GAC TTA GTG AAG CCT GGA | 150 |
| Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp Leu Val Lys Pro Gly | |
| 1 5 10 15 | |
| GGG TCC CTG ACA CTC TCC TGT GCA ACC TCT GGA TTC ACT TTC AGT AGT | 198 |
| Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly Phe Thr Phe Ser Ser | |
| 20 25 30 | |
| TAT GCC TTT TCT TGG GTC CGC CAG ACC TCA GAC AAG AGT CTG GAG TGG | 246 |
| Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp Lys Ser Leu Glu Trp | |
| 35 40 45 | |
| GTC GCA ACC ATC AGT AGT ACT GAT ACT TAT ACC TAT TAT TCA GAC AAT | 294 |
| Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr Tyr Ser Asp Asn | |
| 50 55 60 | |
| GTG AAG GGG CGC TTC ACC ATC TCC AGA GAC AAT GGC AAG AAC ACC CTG | 342 |
| Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Gly Lys Asn Thr Leu | |
| 65 70 75 | |
| TAC CTG CAA ATG AGC AGT CTG AAG TCT GAG GAC ACA GCC GTG TAT TAC | 390 |
| Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp Thr Ala Val Tyr Tyr | |
| 80 85 90 95 | |
| TGT GCA AGA CAT GGG TAC TAT GGT AAA GGC TAT TTT GAC TAC TGG GGC | 438 |
| Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr Phe Asp Tyr Trp Gly | |
| 100 105 110 | |
| CAA GGG ACC ACG GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GGC GGA | 486 |
| Gln Gly Thr Thr Val Thr Val Ser Ser Gly Gly Gly Gly Ser Gly Gly | |
| 115 120 125 | |
| GGT GGC TCT GGC GGT GGC GGA TCG GAC ATC GAG CTC ACT CAG TCT CCA | 534 |
| Gly Gly Ser Gly Gly Gly Ser Asp Ile Glu Leu Thr Gln Ser Pro | |
| 130 135 140 | |
| TTC TCC CTG ACT GTG ACA GCA GGA GAG AAG GTC ACT ATG AAT TGC AAG | 582 |
| Phe Ser Leu Thr Val Thr Ala Gly Glu Lys Val Thr Met Asn Cys Lys | |
| 145 150 155 | |
| TCC GGT CAG AGT CTG TTA AAC AGT GTA AAT CAG AGG AAC TAC TTG ACC | 630 |
| Ser Gly Gln Ser Leu Leu Asn Ser Val Asn Gln Arg Asn Tyr Leu Thr | |
| 160 165 170 175 | |

52

| | |
|---|-----|
| TGG TAC CAG CAG AAG CCA GGG CAG CCT CCT AAA CTG TTG ATC TAC TGG | 678 |
| Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile Tyr Trp | |
| 180 185 190 | |
| GCA TCC ACT AGG GAA TCT GGA GTC CCT GAT CGC TTC ACA GCC AGT GGA | 726 |
| Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg Phe Thr Ala Ser Gly | |
| 195 200 205 | |
| TCT GGA ACA GAT TTC ACT CTC ACC ATC AGC AGT GTG CAG GCT GAA GAC | 774 |
| Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Val Gln Ala Glu Asp | |
| 210 215 220 | |
| CTG GCA GTT TAT TAC TGT CAG AAT GAT TAT ACT TAT CCG TTC ACG TTC | 822 |
| Leu Ala Val Tyr Tyr Cys Gln Asn Asp Tyr Thr Pro Phe Thr Phe | |
| 225 230 235 | |
| GGA GGG GGG ACC AAG CTC GAG ATC AAA CGG GAA CAA AAA CTC ATC TCA | 870 |
| Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Glu Gln Lys Leu Ile Ser | |
| 240 245 250 255 | |
| GAA GAG GAT CTG AAT TAA TAAGATCAAA CGGTAATAAG GATCCAGCTC GAATTC | 924 |
| Glu Glu Asp Leu Asn * | |
| 260 | |

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 283 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

| | |
|---|--|
| Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala | |
| -22 -20 -15 -10 | |
| Ala Gln Pro Ala Met Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp | |
| -5 1 5 10 | |
| Leu Val Lys Pro Gly Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly | |
| 15 20 25 | |
| Phe Thr Phe Ser Ser Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp | |
| 30 35 40 | |
| Lys Ser Leu Glu Trp Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr | |
| 45 50 55 | |
| Tyr Tyr Ser Asp Asn Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn | |
| 60 65 70 | |
| Gly Lys Asn Thr Leu Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp | |
| 75 80 85 90 | |
| Thr Ala Val Tyr Tyr Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr | |
| 95 100 105 | |
| Phe Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Gly | |
| 110 115 120 | |
| Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Asp Ile Glu | |
| 125 130 135 | |
| Leu Thr Gln Ser Pro Phe Ser Leu Thr Val Thr Ala Gly Glu Lys Val | |
| 140 145 150 | |

53

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Thr Met Asn Cys Lys Ser Gly Gln Ser Leu Leu Asn Ser Val Asn Gln
155                               160                               165                               170

Arg Asn Tyr Leu Thr Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys
                               175                               180                               185

Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg
                               190                               195                               200

Phe Thr Ala Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser
                               205                               210                               215

Val Gln Ala Glu Asp Leu Ala Val Tyr Tyr Cys Gln Asn Asp Tyr Thr
                               220                               225                               230

Tyr Pro Phe Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Glu
235                               240                               245                               250

Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn *
                               255                               260

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(2) INFORMATION FOR SEQ ID NO: 11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1706 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "cDNA domains with synthetic linker(s)"
- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: HindIII-EcoRI insert of pGOSA.E
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 40..864
 - (D) OTHER INFORMATION: /product= "pelB-VH4715-LiA-VH3418"
- (ix) FEATURE:
 - (A) NAME/KEY: sig_peptide
 - (B) LOCATION: 40..105
 - (D) OTHER INFORMATION: /product= "pectate lyase"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 106..465
 - (D) OTHER INFORMATION: /product= "VH4715"
- (ix) FEATURE:
 - (A) NAME/KEY: misc_RNA
 - (B) LOCATION: 466..522
 - (D) OTHER INFORMATION: /product= "linkerA
(Gly4Ser)3AlaGlySerAla"
- (ix) FEATURE:
 - (A) NAME/KEY: mat_peptide
 - (B) LOCATION: 523..861
 - (D) OTHER INFORMATION: /product= "VH3418"
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 913..1689
 - (D) OTHER INFORMATION: /product= "pelB-VL3418-LiV-VL4715"

(ix) FEATURE:
 (A) NAME/KEY: sig_peptide
 (B) LOCATION:913..978
 (D) OTHER INFORMATION:/product= "pectate lyase"

(ix) FEATURE:
 (A) NAME/KEY: mat_peptide
 (B) LOCATION:979..1299
 (D) OTHER INFORMATION:/product= "VL3418"

(ix) FEATURE:
 (A) NAME/KEY: misc_RNA
 (B) LOCATION:1300..1344
 (D) OTHER INFORMATION:/product= "linker V
 (Gly4Ser)2Gly4Val"

(ix) FEATURE:
 (A) NAME/KEY: mat_peptide
 (B) LOCATION:1345..1686
 (D) OTHER INFORMATION:/product= "VL4715"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

| | |
|---|-----|
| AAGCTTGCAT GGAAATTCTA TTTCAAGGAG ACAGTCATA ATG AAA TAC CTA TTG | 54 |
| Met Lys Tyr Leu Leu | |
| -22 -20 | |
| CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA GCG ATG | 102 |
| Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro Ala Met | |
| -15 -10 -5 | |
| GCC CAG GTG CAG CTG CAG GAG TCA GGG GGA GAC TTA GTG AAG CCT GGA | 150 |
| Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp Leu Val Lys Pro Gly | |
| 1 5 10 15 | |
| GGG TCC CTG ACA CTC TCC TGT GCA ACC TCT GGA TTC ACT TTC AGT AGT | 198 |
| Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly Phe Thr Phe Ser Ser | |
| 20 25 30 | |
| TAT GCC TTT TCT TGG GTC CGC CAG ACC TCA GAC AAG AGT CTG GAG TGG | 246 |
| Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp Lys Ser Leu Glu Trp | |
| 35 40 45 | |
| GTC GCA ACC ATC AGT AGT ACT GAT ACT TAT ACC TAT TAT TCA GAC AAT | 294 |
| Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr Tyr Tyr Ser Asp Asn | |
| 50 55 60 | |
| GTG AAG GGG CGC TTC ACC ATC TCC AGA GAC AAT GGC AAG AAC ACC CTG | 342 |
| Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Gly Lys Asn Thr Leu | |
| 65 70 75 | |
| TAC CTG CAA ATG AGC AGT CTG AAG TCT GAG GAC ACA GCC GTG TAT TAC | 390 |
| Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp Thr Ala Val Tyr Tyr | |
| 80 85 90 95 | |
| TGT GCA AGA CAT GGG TAC TAT GGT AAA GGC TAT TTT GAC TAC TGG GGC | 438 |
| Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr Phe Asp Tyr Trp Gly | |
| 100 105 110 | |
| CAA GGG ACC ACG GTC ACC GTC TCC TCA GGT GGA GGC GGT TCA GGC GGA | 486 |
| Gln Gly Thr Thr Val Thr Val Ser Ser Gly Gly Gly Gly Ser Gly Gly | |
| 115 120 125 | |
| GGT GGC TCT GGC GGT GGC GGA TCG GCC GGT TCG GCC CAG GTC CAG CTG | 534 |
| Gly Gly Ser Gly Gly Gly Gly Ser Ala Gly Ser Ala Gln Val Gln Leu | |
| 130 135 140 | |

| | |
|---|------|
| CAA CAG TCA GGA CCT GAG CTG GTA AAG CCT GGG GCT TCA GTG AAG ATG Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly Ala Ser Val Lys Met 145 150 155 | 582 |
| TCC TGC AAG GCT TCT GGA TAC ACA TTC ACT AGC TAT GTT ATG CAC TGG Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser Tyr Val Met His Trp 160 165 170 175 | 630 |
| GTG AAA CAG AAG CCT GGG CAG GGC CTT GAG TGG ATT GGA TAT ATT TAT Val Lys Gln Lys Pro Gly Gln Gly Leu Glu Trp Ile Gly Tyr Ile Tyr 180 185 190 | 678 |
| CCT TAC AAT GAT GGT ACT AAG TAC AAT GAG AAG TTC AAA GGC AAG GCC Pro Tyr Asn Asp Gly Thr Lys Tyr Asn Glu Lys Phe Lys Gly Lys Ala 195 200 205 | 726 |
| ACA CTG ACT TCA GAC AAA TCC TCC AGC ACA GCC TAC ATG GAG CTC AGC Thr Leu Thr Ser Asp Lys Ser Ser Thr Ala Tyr Met Glu Leu Ser 210 215 220 | 774 |
| AGC CTG ACC TCT GAG GAC TCT GCG GTC TAT TAC TGT TCA AGA CGC TTT Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr Cys Ser Arg Arg Phe 225 230 235 | 822 |
| GAC TAC TGG GGC CAA GGG ACC ACC GTC ACC GTC TCC TCA TAA Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser * 240 245 250 | 864 |
| TAAGCTAGCG GAGCTGCATG CAAATTCTAT TTCAAGGAGA CAGTCATA ATG AAA TAC Met Lys Tyr -22 -20 | 921 |
| CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC GCT GCC CAA CCA Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala Ala Gln Pro -15 -10 -5 | 969 |
| GCG ATG GCC GAC ATC GAG CTC ACC CAG TCT CCA TCT TCC ATG TAT GCA Ala Met Ala Asp Ile Glu Leu Thr Gln Ser Pro Ser Ser Met Tyr Ala 1 5 10 | 1017 |
| TCT CTA GGA GAG AGA ATC ACT ATC ACT TGC AAG GCG AGT CAG GAC ATT Ser Leu Gly Glu Arg Ile Thr Ile Thr Cys Lys Ala Ser Gln Asp Ile 15 20 25 | 1065 |
| AAT ACC TAT TTA ACC TGG TTC CAG CAG AAA CCA GGG AAA TCT CCC AAG Asn Thr Tyr Leu Thr Trp Phe Gln Gln Lys Pro Gly Lys Ser Pro Lys 30 35 40 45 | 1113 |
| ACC CTG ATC TAT CGT GCA AAC AGA TTG CTA GAT GGG GTC CCA TCA AGG Thr Leu Ile Tyr Arg Ala Asn Arg Leu Leu Asp Gly Val Pro Ser Arg 50 55 60 | 1161 |
| TTC AGT GGC AGT GGA TCT GGG CAA GAT TAT TCT CTC ACC ATC AGC AGC Phe Ser Gly Ser Gly Ser Gly Gln Asp Tyr Ser Leu Thr Ile Ser Ser 65 70 75 | 1209 |
| CTG GAC TAT GAA GAT ATG GGA ATT TAT TAT TGT CTA CAA TAT GAT GAG Leu Asp Tyr Glu Asp Met Gly Ile Tyr Tyr Cys Leu Gln Tyr Asp Glu 80 85 90 | 1257 |
| TTG TAC ACG TTC GGA GGG GGG ACC AAG CTC GAG ATC AAA CGG GGT GGA Leu Tyr Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Gly Gly 95 100 105 | 1305 |
| GGC GGT TCA GGC GGA GGT GGC TCT GGC GGT GGC GGA GTC GAC ATC GAA Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Val Asp Ile Glu 110 115 120 125 | 1353 |

56

| | |
|---|------|
| CTC ACT CAG TCT CCA TTC TCC CTG ACT GTG ACA GCA GGA GAG AAG GTC | 1401 |
| Leu Thr Gln Ser Pro Phe Ser Leu Thr Val Thr Ala Gly Glu Lys Val | |
| 130 135 140 | |
| ACT ATG AAT TGC AAG TCC GGT CAG AGT CTG TTA AAC AGT GTA AAT CAG | 1449 |
| Thr Met Asn Cys Lys Ser Gly Gln Ser Leu Leu Asn Ser Val Asn Gln | |
| 145 150 155 | |
| AGG AAC TAC TTG ACC TGG TAC CAG CAG AAG CCA GGG CAG CCT CCT AAA | 1497 |
| Arg Asn Tyr Leu Thr Trp Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys | |
| 160 165 170 | |
| CTG TTG ATC TAC TGG GCA TCC ACT AGG GAA TCT GGA GTC CCT GAT CGC | 1545 |
| Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val Pro Asp Arg | |
| 175 180 185 | |
| TTC ACA GCC AGT GGA TCT GGA ACA GAT TTC ACT CTC ACC ATC AGC AGT | 1593 |
| Phe Thr Ala Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser | |
| 190 195 200 205 | |
| GTG CAG GCT GAA GAC CTG GCA GTT TAT TAC TGT CAG AAT GAT TAT ACT | 1641 |
| Val Gln Ala Glu Asp Leu Ala Val Tyr Tyr Cys Gln Asn Asp Tyr Thr | |
| 210 215 220 | |
| TAT CCG TTC ACG TTC GGA GGG GGG ACC AAG CTC GAA ATC AAA CGG TAA | 1689 |
| Tyr Pro Phe Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg * | |
| 225 230 235 | |
| TAAGCGGCCG CGAATTC | 1706 |

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 275 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

| | |
|---|--|
| Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Ala | |
| -22 -20 -15 -10 | |
| Ala Gln Pro Ala Met Ala Gln Val Gln Leu Gln Glu Ser Gly Gly Asp | |
| -5 1 5 10 | |
| Leu Val Lys Pro Gly Gly Ser Leu Thr Leu Ser Cys Ala Thr Ser Gly | |
| 15 20 25 | |
| Phe Thr Phe Ser Ser Tyr Ala Phe Ser Trp Val Arg Gln Thr Ser Asp | |
| 30 35 40 | |
| Lys Ser Leu Glu Trp Val Ala Thr Ile Ser Ser Thr Asp Thr Tyr Thr | |
| 45 50 55 | |
| Tyr Tyr Ser Asp Asn Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn | |
| 60 65 70 | |
| Gly Lys Asn Thr Leu Tyr Leu Gln Met Ser Ser Leu Lys Ser Glu Asp | |
| 75 80 85 90 | |
| Thr Ala Val Tyr Tyr Cys Ala Arg His Gly Tyr Tyr Gly Lys Gly Tyr | |
| 95 100 105 | |

57

Phe Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly Gly
 110 115 120
 Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Ala Gly Ser
 125 130 135
 Ala Gln Val Gln Leu Gln Gln Ser Gly Pro Glu Leu Val Lys Pro Gly
 140 145 150
 Ala Ser Val Lys Met Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Ser
 155 160 165 170
 Tyr Val Met His Trp Val Lys Gln Lys Pro Gly Gln Gly Leu Glu Trp
 175 180 185
 Ile Gly Tyr Ile Tyr Pro Tyr Asn Asp Gly Thr Lys Tyr Asn Glu Lys
 190 195 200
 Phe Lys Gly Lys Ala Thr Leu Thr Ser Asp Lys Ser Ser Ser Thr Ala
 205 210 215
 Tyr Met Glu Leu Ser Ser Leu Thr Ser Glu Asp Ser Ala Val Tyr Tyr
 220 225 230
 Cys Ser Arg Arg Phe Asp Tyr Trp Gly Gln Gly Thr Thr Val Thr Val
 235 240 245 250
 Ser Ser *

(2) INFORMATION FOR SEQ ID NO: 13:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 259 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

Met Lys Tyr Leu Leu Pro Thr Ala Ala Ala Gly Leu Leu Leu Leu Ala
 -22 -20 -15 -10
 Ala Gln Pro Ala Met Ala Asp Ile Glu Leu Thr Gln Ser Pro Ser Ser
 -5 1 5 10
 Met Tyr Ala Ser Leu Gly Glu Arg Ile Thr Ile Thr Cys Lys Ala Ser
 15 20 25
 Gln Asp Ile Asn Thr Tyr Leu Thr Trp Phe Gln Gln Lys Pro Gly Lys
 30 35 40
 Ser Pro Lys Thr Leu Ile Tyr Arg Ala Asn Arg Leu Leu Asp Gly Val
 45 50 55
 Pro Ser Arg Phe Ser Gly Ser Gly Ser Gly Gln Asp Tyr Ser Leu Thr
 60 65 70
 Ile Ser Ser Leu Asp Tyr Glu Asp Met Gly Ile Tyr Tyr Cys Leu Gln
 75 80 85 90
 Tyr Asp Glu Leu Tyr Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile Lys
 95 100 105
 Arg Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Val
 110 115 120

58

Asp Ile Glu Leu Thr Gln Ser Pro Phe Ser Leu Thr Val Thr Ala Gly
 125 130 135
 Glu Lys Val Thr Met Asn Cys Lys Ser Gly Gln Ser Leu Leu Asn Ser
 140 145 150
 Val Asn Gln Arg Asn Tyr Leu Thr Trp Tyr Gln Gln Lys Pro Gly Gln
 155 160 165 170
 Pro Pro Lys Leu Leu Ile Tyr Trp Ala Ser Thr Arg Glu Ser Gly Val
 175 180 185
 Pro Asp Arg Phe Thr Ala Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr
 190 195 200
 Ile Ser Ser Val Gln Ala Glu Asp Leu Ala Val Tyr Tyr Cys Gln Asn
 205 210 215
 Asp Tyr Thr Tyr Pro Phe Thr Phe Gly Gly Gly Thr Lys Leu Glu Ile
 220 225 230
 Lys Arg *
 235

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "synthetic DNA"

(vii) IMMEDIATE SOURCE:

- (B) CLONE: primer DBL.1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

CACCATCTCC AGAGACAATG GCAAG

25

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "synthetic DNA"

(vii) IMMEDIATE SOURCE:

- (B) CLONE: primer DBL.2

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

GAGCGCGAGC TCGGCCGAAC CGGCCGATCC GCCACCGCCA GAGCC

45

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 45 base pairs

59

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.3

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

CAGGATCCGG CCGGTTCCGC CCAGGTCCAG CTGCAACAGT CAGGA

45

- (2) INFORMATION FOR SEQ ID NO: 17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 53 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.4

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

CTACATGAAT TCGCTAGCTT ATTATGAGGA GACGGTGACG GTGGTCCCTT GGC

53

- (2) INFORMATION FOR SEQ ID NO: 18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.5

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

TAATAAGCTA GCGGAGCTGC ATGCAAATTC TATTTC

36

- (2) INFORMATION FOR SEQ ID NO: 19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.6

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

ACCAAGCTCG AGATCAAACG GGG

23

(2) INFORMATION FOR SEQ ID NO: 20:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 36 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "synthetic DNA"

(vii) IMMEDIATE SOURCE:

(B) CLONE: primer DBL.7

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

AATGTCGAAT TCGTCGACTC CGCCACCGCC AGAGCC

36

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 39 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "synthetic DNA"

(vii) IMMEDIATE SOURCE:

(B) CLONE: primer DBL.8

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

ATTGGAGTCG ACATCGAACT CACTCAGTCT CCATTCTCC

39

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 50 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "synthetic DNA"

(vii) IMMEDIATE SOURCE:

(B) CLONE: primer DBL.9

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

TGAAGTGAAT TCGCGGCCGC TTATTACCGT TTGATTTCGA GCTTGGTCCC

50

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 41 base pairs
- (B) TYPE: nucleic acid

61

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.10

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

CGAATTCGGT CACCGTCTCC TCACAGGTCC AGTTGCAACA G

41

- (2) INFORMATION FOR SEQ ID NO: 24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 44 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.11

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

CGAATTCTCG AGATCAAACG GGACATCGAA CTCACTCAGT CTCC

44

- (2) INFORMATION FOR SEQ ID NO: 25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 41 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer DBL.12

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 25:

CGAATTCGGT CACCGTCTCC TCACAGGTGC AGTTGCAGGA G

41

- (2) INFORMATION FOR SEQ ID NO: 26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 22 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "synthetic DNA"

- (vii) IMMEDIATE SOURCE:
 - (B) CLONE: primer PCR.51

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:
AGGTSMMAMCT GCAGSAGTCW GG 22
- (2) INFORMATION FOR SEQ ID NO: 27:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
- (vii) IMMEDIATE SOURCE:
(B) CLONE: primer PCR.89
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:
TGAGGAGACG GTGACCGTGG TCCCTTGCC CC 32
- (2) INFORMATION FOR SEQ ID NO: 28:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
- (vii) IMMEDIATE SOURCE:
(B) CLONE: primer PCR.90
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:
GACATTGAGC TCACCCAGTC TCCA 24
- (2) INFORMATION FOR SEQ ID NO: 29:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
- (vii) IMMEDIATE SOURCE:
(B) CLONE: primer PCR.116
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:
GTTAGATCTC GAGCTTGGTC CC 22
- (2) INFORMATION FOR SEQ ID NO: 30:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 13 amino acids
(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:

Thr Thr Val Thr Val Ser Ser Gln Val Gln Leu Gln Gln
1 5 10

(2) INFORMATION FOR SEQ ID NO: 31:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 12 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:

Lys Leu Glu Ile Lys Arg Asp Ile Glu Leu Thr Gln
1 5 10

DP/A/II/12
page 14

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

TO Dr. P van der Logt
Unilever Research
Colworth Laboratory
Biosciences Division
Colworth House, Sharnbrook
Bedford MK44 1LQ
NAME AND ADDRESS
OF DEPOSITOR

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
Issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

| | |
|--|---|
| I. IDENTIFICATION OF THE MICROORGANISM | |
| Identification reference given by the DEPOSITOR: Escherichia coli ScFy 4715.myc | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: NCTC 12916 |
| II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION | |
| The microorganism identified under I above was accompanied by: | |
| <input type="checkbox"/> a scientific description <input checked="" type="checkbox"/> a proposed taxonomic designation (Mark with a cross where applicable) | |
| III. RECEIPT AND ACCEPTANCE | |
| This International Depositary Authority accepts the microorganism identified under I above, which was received by it on 14th October 1995 (date of original deposit) ¹ | |
| IV. INTERNATIONAL DEPOSITARY AUTHORITY | |
| Name: National Collection of Type Cultures Address: Central Public Health Laboratory 61 Colindale Avenue London NW9 5HT | Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s): Date: 4/12/95 <i>Benny Holmes</i> B Holmes Clinical Scientist |

¹ Where Rule 6.4(d) applies, such date is the date on which the status of international depositary authority was acquired; where a deposit made outside the Budapest Treaty after the acquisition of the status of international depositary authority is converted into a deposit under the Budapest Treaty, such date is the date on which the microorganism was received by the international depositary authority.

BP/A/II/12
page 24BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Dr P van der Logt
Biliver Research
Elworth Laboratory
Cosciences Division
Elworth House, Sharnbrook
Bedford MK44 1LQ

VIABILITY STATEMENT
issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified on the following page

NAME AND ADDRESS OF THE PARTY
WHICH THE VIABILITY STATEMENT
IS ISSUED

| | |
|---|---|
| I. DEPOSITOR | II. IDENTIFICATION OF THE MICROORGANISM |
| Name: Dr P van der Logt Address: As above | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: NCTC 12916: Date of the deposit or of the transfer: 14th October 1995 |
| 3. VIABILITY STATEMENT | |
| 1. Viability of the microorganism identified under II above was tested 2. On that date, the said microorganism was <input type="checkbox"/> viable <input type="checkbox"/> no longer viable | |

Indicate the date of the original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).

In the cases referred to in Rule 10.2(a) (ii) and (iii), refer to the most recent viability test.

Mark with a cross the applicable box.

SP/A/II/12
page 25IV. CONDITIONS UNDER WHICH THE VIABILITY TEST HAS BEEN PERFORMED⁴

Nutrient Agar without additions (a)

Agar (17g Bacto-Tryptone, 10g Bacto-Yeast Extract, 5g NaCl, per litre) with 100ug/ml ampicillin and 1% glucose (b)

Aerobic incubation, 37°C, 24 hrs

(a) 8×10^5 cfu/ml(b) 6×10^5 cfu/ml

INTERNATIONAL DEPOSITARY AUTHORITY

Name:

National Collection of Type Cultures
Central Public Health Laboratory
61 Colindale Avenue, London NW9 5HT
Telephone: 0181-300 4400
Telex: 8933841 (CODED G)
Fax: 0181-300 7874

Address:

Signature(s) of person(s) having the power
to represent the International Depository
Authority or of authorized official(s):

Date: 4/12/95

Barry Holmes
B Holmes
Clinical Scientist

Fill in if the information has been requested and if the results of the test were negative.

BP/A/II/12
page 14

BUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

to Dr. P van der Logt
Unilever Research
Colworth Laboratory
Biosciences Division
Colworth House, Sharnbrook
Bedford MK44 1LQ

NAME AND ADDRESS
OF DEPOSITOR

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT
issued pursuant to Rule 7.1 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified at the bottom of this page

| | |
|---|---|
| I. IDENTIFICATION OF THE MICROORGANISM | |
| Identification reference given by the DEPOSITOR: Escherichia coli Fv 3418 | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: NCTC 12915 |
| II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DESIGNATION | |
| The microorganism identified under I above was accompanied by: | |
| <input type="checkbox"/> a scientific description <input checked="" type="checkbox"/> a proposed taxonomic designation (Mark with a cross where applicable) | |
| III. RECEIPT AND ACCEPTANCE | |
| This International Depositary Authority accepts the microorganism identified under I above, which was received by it on <u>14th October 1995 (date of original deposit)</u> ¹ | |
| IV. INTERNATIONAL DEPOSITARY AUTHORITY | |
| Name: National Collection of Type Cultures Address: Central Public Health Laboratory 61 Colindale Avenue London NW9 5HT | Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s): Date: 4/12/95 <i>Barry Holmes</i> B Holmes Clinical Scientist |

¹ Where Rule 6.4(d) applies, such date is the date on which the status of international depositary authority was acquired; where a deposit made outside the Budapest Treaty after the acquisition of the status of international depositary authority is converted into a deposit under the Budapest Treaty, such date is the date on which the microorganism was received by the international depositary authority.

UP/A/II/12
page 24

**HUDAPEST TREATY ON THE INTERNATIONAL
RECOGNITION OF THE DEPOSIT OF MICROORGANISMS
FOR THE PURPOSES OF PATENT PROCEDURE.**

INTERNATIONAL FORM

Dr P van der Logt
Biliver Research
Bilworth Laboratory
Biosciences Division
Bilworth House, Sharnbrook
Bedford MK44 1LQ

VIABILITY STATEMENT
issued pursuant to Rule 10.2 by the
INTERNATIONAL DEPOSITARY AUTHORITY
identified on the following page

NAME AND ADDRESS OF THE PARTY
WHICH THE VIABILITY STATEMENT
IS ISSUED

| | |
|--|--|
| I. DEPOSITOR | II. IDENTIFICATION OF THE MICROORGANISM |
| Name: Dr P van der Logt Address: As above | Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: NCTC 12915 Date of the deposit or of the transfer: 14th October 1995 |
| III. VIABILITY STATEMENT | |
| Viability of the microorganism identified under II above was tested 20th November 1995 ² . On that date, the said microorganism was <input checked="" type="checkbox"/> viable <input type="checkbox"/> no longer viable | |

Indicate the date of the original deposit or, where a new deposit or a transfer has been made, the most recent relevant date (date of the new deposit or date of the transfer).

(In the boxes referred to in Rule 10.2(a)(ii) and (iii), refer to the most recent viability test).

Mark with a cross the applicable box.

PCT/EP96/03605
page: 22IV. CONDITIONS UNDER WHICH THE VIABILITY TEST HAS BEEN PERFORMED⁴

Nutrient Agar without additions (a)
Agar (17g Bacto-Tryptone, 10g Bacto-Yeast Extract, 5g NaCl, per
litre) with 100ug/ml ampicillin and 1% glucose. (b)
Aerobic incubation, 37°C, 24 hrs.
(a) 2×10^6 cfu/ml
(b) 1×10^6 cfu/ml

INTERNATIONAL DEPOSITARY AUTHORITY

Address:

National Collection of Type Cultures
Central Public Health Laboratory
61 Colindale Avenue, London NW9 5HT
Telephone: 0181-300 4400
Telex: 883394Z (DHPEUK G)
Fax: 0181-300 7874

Signature(s) of person(s) having the power
to represent the International Depositary
Authority or of authorized official(s):

Date: 4/12/95

Bunny Holmes
B Holmes
Clinical Scientist

Fill in if the information has been requested and if the results of the test were negative.

C L A I M S

1. A bispecific or bivalent antibody fragment analogue, which comprises a binding complex containing two polypeptide chains, whereby one polypeptide chain comprises two times a variable domain of a heavy chain (V_H) **in series** and the other polypeptide chain comprises two times a variable domain of a light chain (V_L) **in series**, and the binding complex contains two pairs of variable domains (V_H -A// V_L -A and V_H -B// V_L -B).
2. An antibody fragment analogue according to claim 1, in which one polypeptide chain comprises a first V_H connected to a second V_H and the other polypeptide chain comprises a first V_L connected to a second V_L .
3. An antibody fragment analogue according to claim 2, in which the two V_H 's are directly connected to each other without an intermediate peptide linker.
4. An antibody fragment analogue according to claim 2, in which the two V_L 's are directly connected to each other without an intermediate peptide linker.
5. An antibody fragment analogue according to claim 3 or claim 4, in which one polypeptide chain comprises a first V_H directly connected to a second V_H , and the other polypeptide chain comprises a first V_L directly connected to a second V_L .
6. An antibody fragment analogue according to claim 2, in which the two V_H 's are connected to each other by a peptide linker and also the two V_L 's are connected to each other by a peptide linker, each peptide linker comprising at least one amino acid residue.

7. An antibody fragment analogue according to claim 6, in which one polypeptide chain comprises a first V_H (V_H -A) followed by a second V_H (V_H -B) and the other polypeptide chain comprises a first V_L (V_L -A) followed by a second V_L (V_L -B), and in which the two V_H 's are connected to each other by a peptide linker (Li_H), thus [V_H -A * Li_H * V_H -B], and also the two V_L 's are connected to each other by a peptide linker (Li_L), thus [V_L -A * Li_L * V_L -B], each peptide linker comprising at least 10 amino acid residues.

8. An antibody fragment analogue according to claim 2, in which one polypeptide chain comprises a first V_H (V_H -A) followed by a second V_H (V_H -B) with or without a connecting peptide linker (Li_H), thus [V_H -A * (Li_H) * V_H -B], and the other polypeptide chain comprises a first V_L (V_L -A) preceded by a second V_L (V_L -B) with or without a connecting peptide linker (Li_L), thus [V_L -B * (Li_L) * V_L -A].

9. An antibody fragment analogue according to claim 1, in which the two variable domains are different resulting in a **bispecific** antibody fragment analogue.

10. An antibody fragment analogue according to claim 1, in which the specificities A and B are the same resulting in a **bivalent** antibody fragment analogue.

11. Use of an antibody fragment analogue according to claim 1, in immunoassays including diagnostic techniques, in agglutination assays, in a purification method, for compositions suitable for therapy, or in other methods in which immunoglobulins or fragments thereof can be used.

12. A process for producing an antibody fragment analogue according to any one of claims 1-10, which comprises

(1) transforming a host by incorporating into that host a DNA encoding the two V_H 's in series with or without a

connecting peptide linker and a DNA encoding the two V_L 's in series with or without a connecting peptide linker,

(2) culturing such transformed host under conditions whereby the connected V_H 's and the connected V_L 's are formed, and

(3) allowing the two connected V_H 's and the two connected V_L 's to combine to each other under formation of a double head antibody fragment analogue, and

(4) optionally collecting the double head antibody fragment analogue.

13. A process for producing an antibody fragment analogue according to any one of claims 1-10, which comprises

(1) transforming a first host by incorporating into that first host a DNA encoding the two V_H 's in series with or without a connecting peptide linker,

(2) transforming a second host by incorporating into that second host a DNA encoding the two V_L 's in series with or without a connecting peptide linker,

(3) culturing the first and second transformed host under conditions whereby the connected V_H 's and the connected V_L 's, respectively, are formed,

(4) optionally collecting separately the two connected V_H 's and the two connected V_L 's, and

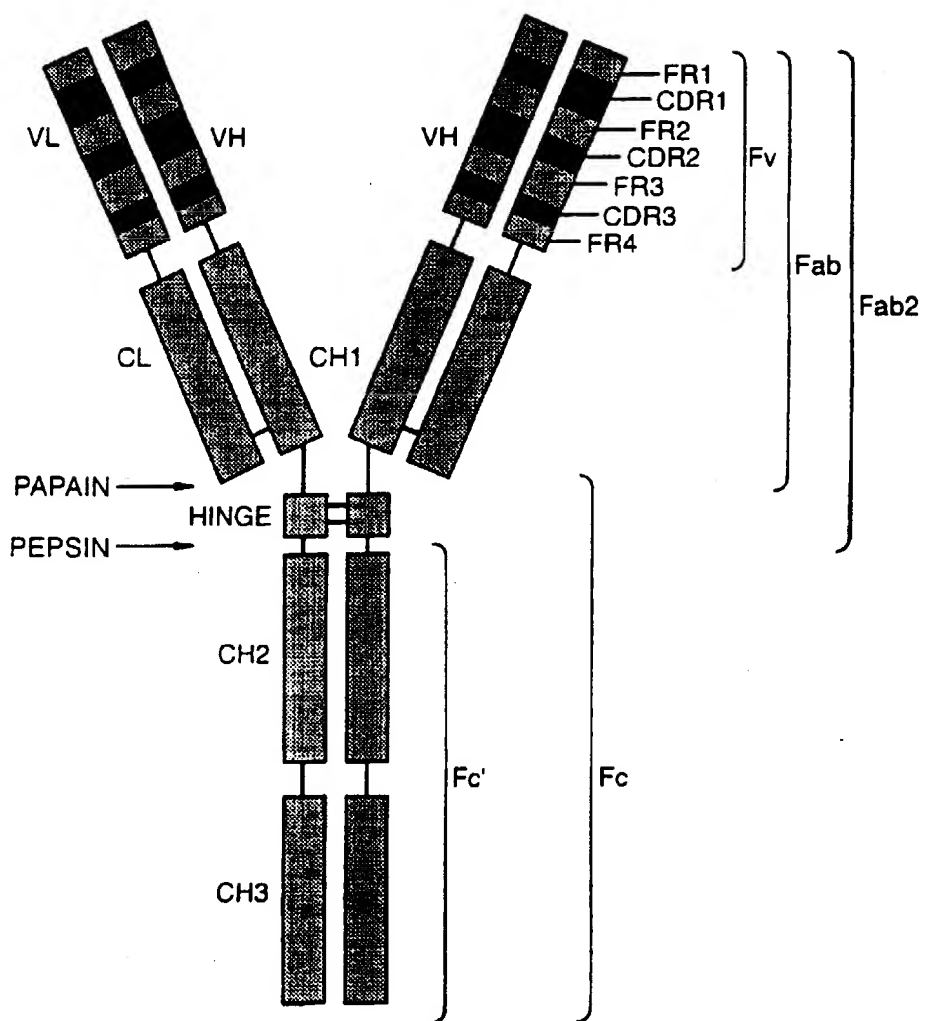
(5) combining the connected V_H 's and the connected V_L 's under conditions that they can form a double head antibody fragment analogue.

14. A process according to claim 12 or 13, in which the host is selected from the group consisting of prokaryotic micro-organisms comprising Gram-negative bacteria (e.g. *E. coli*) and Gram-positive bacteria (e.g. *B. subtilis* or lactic acid bacteria), lower eukaryotic microorganisms comprising yeasts (e.g. belonging to the genera *Saccharomyces*, *Kluyveromyces*, *Pichia*, and *Hansenula*) and moulds (e.g. belonging to the genera *Aspergillus*,

Neurospora or *Trichoderma*), and higher eukaryotic organisms (e.g. plants) or cell cultures thereof (e.g. hybridoma's).

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Fig.1.



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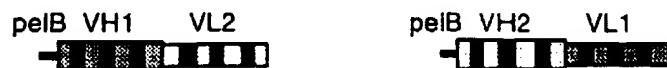
Fig.2A.



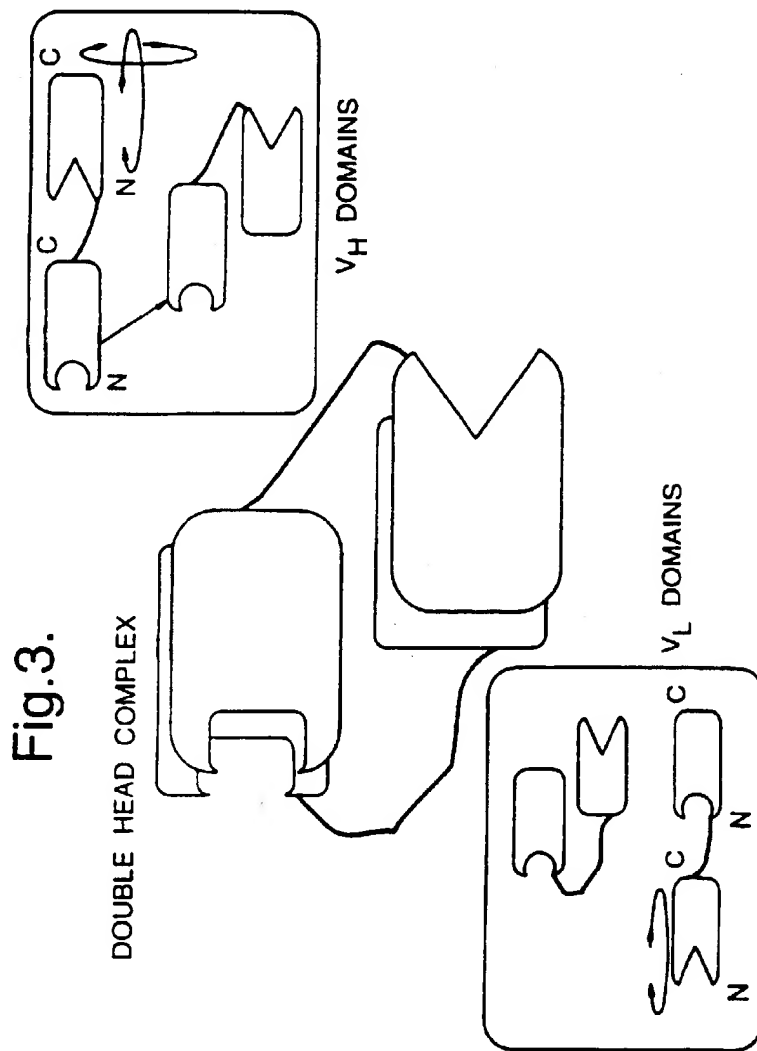
Fig.2B.



Fig.2C.



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Fig.4.

D I E L T Q S P A S L S A S V G E
 GAATTCGGCCGACATCGAGCTCAGCTCAGCTCCCTTTCTGCGTCTGTGGGAGA
 EcoRI SacI 60

T V T I T C R A S G N I H N Y L A W Y Q
 AACTGTCACCATCACATGTCGAGCAAGTGGGAATATTCACAATTATTTAGCATGGTATCA
 120

Q K Q G K S P Q L L V Y Y T T T L A D G
 GCAGAAACAGGGAAAATCTCCTCAGCTCCTGGTCTATTATACAACAACCTTAGCAGATGG
 180

V P S R F S G S G S G T Q Y S L K I N S
 TGTGCCATCAAGGTTTCAAGTGGCAGTGGATCAGGAACACAATATTCTCTCAAGATCAACAG
 240

L Q P E D F G S Y Y C Q H F W S T P R T
 CCTGCAACCTGAAGATTTTGGGAGTTATTACTGTCAACATTTTGGAGTACTCCTCGGAC
 300

F G G G T K L E I K R G G G G S G G G G
 GTTCGGTGGAGGGACCAAGCTCGAGATCAAACGGGGTGGAGGCGGTTAGGGCGGAGGTGG
 XhoI 360

S G G G G S Q V Q L Q E S Q P G L V A P
 CTCTGGCGGTGGCGGATCGCAGGTGCAGCTGCAGAGTCAAGACCTGGCCTGGTGGCGCC
 PstI 420

S Q S L S I T C T V S G F S L T G Y G V
 CTCACAGAGCCTGTCCATCACATGCACCGTCTCAGGGTTCTCATTAACCGGCTATGGTGT
 480

N W V R Q P P G K G L E W L G M I W G D
 AAAGTGGGTTCCGCGAGCTCCAGGAAAGGGTCTGGAGTGGCTGGGAATGATTTGGGGTGA
 540

G N T D Y N S A L K S R L S I S K D N S
 TGGAAACACAGACTATAATTCAGCTCTCAAATCCAGACTGAGCATCAGCAAGGACAATC
 600

K S Q V F L K M N S L H T D D T A R Y Y
 CAAGAGCCAAGTTTCTTAAAAATGAACAGTCTGCACACTGATGACACAGCCAGGTACTA
 660

C A R E R D Y R L D Y W G Q G T T V T V
 CTGTGCCAGAGAGAGAGATTATAGGCTTGACTACTGGGGCCAAGGGACCACGGTCACCGT
 720

S S *
 CTCCTCATGATAAGCTT
 HindIII 737

DIELTQSPAS---GGGTRLEIKR = VLLys
 GGGGSGGGGSGGGGS = Linker
 QVQLQESGGP---GGGTTTVTVSS = VHLys

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Fig.5.

M K Y L L P T A
 pelB AAGCTTGCAAATTCCTATTTCAAGGAGACAGTCATAATGAAATACCTATTGCCTACGGCAG
 LEADER A A G L L L L A A Q P A M A Q V Q L Q L
 CCGCTGGATTGTTATTACTCGCTGCCCAACCAGCGATGGCCCAGGTGCAGCTGCAGCAGT
 S G P E L V K P G A S V K M S E K A S G
 CAGGACCTGAGCTGGTAAAGCCTGGGGCTTCAGTGAAGATGTCCTGCAAGGCTTCTGGAT
 Y T F T S Y V M H W V K O K P G O G L R
 ACACATTCAGTAGCTATGTTATGCACTGGGTGAAACAGAAGCCTGGGCAGGGCCTTGAGT
 VH3418 W I G Y I Y P Y N D G T K Y N E K F K G
 GGATTGGATATATTTATCCTTACAATGATGGTACTAAGTACAATGAGAAGTTCAAAGGCA
 K A T L T S D K S S S T A Y M E L S S L
 AGGCCACACTGACTTCAGACAAATCCTCCAGCACAGCCTACATGGAGCTCAGCAGCCTGA
 T S E D S A V Y Y C S R R P D Y W G Q G
 CCTCTGAGGACTCTGCGGTCTATTACTGTTCAAGACGCTTTGACTACTGGGGCCAAGGGA
 T T V T V S S
 CCACGGTCACCGTCTCCTCATAATAAGAGCTATGGGAGCTTGCATGCAAATTCCTATTTC
 M K Y L L P T A A A G L L L L
 AGGAGACAGTCATAATGAAATACCTATTGCCTACGGCAGCCGCTGGATTGTTATTACTCG
 pelB A A Q P A M A D I E L T Q S P S S M Y A
 LEADER CTGCCCCAACCAGCGATGGCCGACATCGAGCTCAGCCAGTCTCCATCTTCCATGTATGCAT
 S L G E R I T I T C K A S Q D I N T Y L
 CTCTAGGAGAGAGAATCACTATCACTTGCAAGGCGAGTCAGGACATTAATACCTATTAA
 T W F Q Q K P G K S P K T L I Y R A N R
 CCTGGTTCCAGCAGAAACCAGGGAAATCTCCAAGACCCTGATCTATCGTGCAAACAGAT
 VL3418 L L D G V P S R F S G S G S G Q D Y S L
 TGCTAGATGGGGTCCCATCAAGGTTCAAGTGGCAGTGGATCTGGGCAAGATTATTCTCTCA
 T I S S L D Y E D M G I Y Y C L Q Y D E
 CCATCAGCAGCCTGGACTATGAAGATATGGGAATTTATTATTGCTCTACAATATGATGAGT
 L Y T F G G G T K L E I K R
 TGTACACGTTCCGAGGGGGGACCAAGCTCGAGATCAAACGGTAATAATGATCAAACGGT
 ATAAGGATCCAGCTCGAATTC

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Fig.6.

M K Y L L P T

pelB AAGCTTGCATGCAAATTCTATTTCAAGGAGACAGTCATAATGAAATACCTATTGCCTACG
 LEADER A A A G L L L L A A Q P A M A Q V R L D
 GCAGCCGCTGGATTGTTATTACTCGCTGCCCAACCAGCGATGGCCAGGTGCAGCTGCAG
 E S G G D L V K P G G S L T L S C A T S
 GAGTCAGGGGAGACTTAGTGAAGCCTGGAGGGTCCCTGACACTCTCCTGTGCAACCTCT
 G P T P E S Y A F S W V R D I S D R S L
 GGATTCACCTTCAGTAGTTATGCCTTTTCTTGGGTCCGCCAGACCTCAGACAAGAGTCTG
 E W V A T I S E T D T Y T Y Y S D N V K
 VH4715 GAGTGGGTGCGAACCATCAGTAGTACTGATACCTATACCTATTATTCAGACAATGTGAAG
 G R F T I S R D N G K N T L Y L Q M S S
 GGGCGCTTCACCATCTCCAGAGACAATGGCAAGAACCCTGTACCTGCAAATGAGCAGT
 L K S E D T A V Y Y C A R H G Y Y G K G
 CTGAAGTCTGAGGACACAGCCGTGTATTACTGTGCAAGACATGGGTACTATGGTAAAGGC
 Y F D Y W G O G T T V T V S S
 TATTTTGACTACTGGGGCCAAGGGACCACGGTCACCGTCTCCTCATAATAAGAGCTATGG

M K Y L L P T

pelB GAGCTTGCATGCAAATTCTATTTCAAGGAGACAGTCATAATGAAATACCTATTGCCTACG
 LEADER A A A G L L L L A A Q P A M A D I E L T
 GCAGCCGCTGGATTGTTATTACTCGCTGCCCAACCAGCGATGGCCGACATCGAGCTCACT
 Q S P F S L T V T A G E K V T M N C K S
 CAGTCTCCATTCTCCCTGACTGTGACAGCAGGAGAGAAGGTCACTATGAATTGCAAGTCC
 G Q S L L N S V N Q R N Y L T W Y Q Q K
 GGTCAGAGTCTGTAAACAGTGTAATCAGAGGAACCTTACCTGACCTGGTACCAGCAGAAG
 P G Q P P K L L I Y W A S T R E S G V P
 VL4715 CCAGGGCAGCCTCCTAAACTGTTGATCTACTGGGCATCCACTAGGGAATCTGGAGTCCCT
 D R F T A S G S G T D F T L T I S S V Q
 GATCGCTTCACAGCCAGTGGATCTGGAACAGATTCACTCTCACCATCAGCAGTGTGCAG
 A E D L A V Y Y C Q N D Y T Y P F T F G
 GCTGAAGACCTGGCAGTTTATTACTGTGCAAGATGATTATACTTATCCGTTACGTTTCGGA
 Myc-tag G G T K L E I K R E Q K L I S E E D L N
 GGGGGGACCAAGCTCGAGATCAAACGGGAACAAAACATCTCAGAAGAGGATCTGAAT
 TAATAAGATCAAACGGTAATAAGGATCCAGCTCGAATTC

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Fig.7.

M K Y L L P T

pelB AAGCTTGCATGCAAATTCTATTTCAAGGAGACAGTCATAATGAAATACCTATTGCCTACG

LEADER A A A G L L L L A A Q P A M A Q V Q L G
GCAGCCGCTGGATTGTTATTACTCGCTGCCCAACCAGCGATGGCCCAGGTGCAGCTGCAG
E S G G D L V K P G G S L T L S C A T S
GAGTCAGGGGGAGACTTAGTGAAGCCTGGAGGGTCCCTGACACTCTCCTGTGCAACCTCT
G F T P S S Y A F S W V R O T S D K S L
GGATTCACTTTTCAGTAGTTATGCCTTTTCTTGGGTCCGCCAGACCTCAGACAAGAGTCTG
VH4715 E W V A T I S S T D T Y T Y Y S D N V K
GAGTGGGTCGCAACCATCAGTAGTACTGATACCTTATACCTATTATTTCAGACAATGTGAAG
Q R F T I S R D N G K N T L Y L Q M S S
GGGCGCTTCACCATCTCCAGAGACAATGGCAAGAACACCCTGTACCTGCAAATGAGCAGT
L K S E D T A V Y Y C A R H G Y Y G K G
CTGAAGTCTGAGGACACAGCCGTGTATTACTGTGCAAGACATGGGTACTATGGTAAAGGC
Y F D Y W G Q G T T V T V S S G G G G S
LINKER TATTTTGACTACTGGGGCCAAGGGACCAGGTCAACGTCTCCTCAGGTGGAGGCGGTTCA
G G G G S G G G G S D I E L T Q S P F S
GGCGGAGGTGGCTCTGCCGGTGGCGGATCGGACATCGAGCTCACTCAGTCTCCATTCTCC
L T V T A G E K V T M N C K S G Q S L L
CTGACTGTGACAGCAGGAGAGAAGGTCACTATGAATTGCAAGTCCGGTCAGAGTCTGTTA
N S V K Q R N Y L T W Y Q Q K P G Q P P
AACAGTGTAATTCAGAGGAACACTTACCTGGTACCAGCAGAAGCCAGGGCAGCCTCCT
VL4715 K L L I Y W A S T R E S G V P D R F T A
AAACTGTTGATCTACTGGGCATCCACTAGGGAATCTGGAGTCCCTGATCGCTTCACAGCC
S G S G T D F T L T I S S V Q A E D L A
AGTGGATCTGGAACAGATTTCACTCTCACCATCAGCAGTGTGCAGGCTGAAGACCTGGCA
V Y Y C Q N D Y T Y P F T F G G G T K L
GTTTATTACTGTCAGAATGATTATACCTATCCGTTACGTTCCGAGGGGGGACCAAGCTC
Myc-tag E I K R E Q K L I S E E D L N
GAGATCAAACGGGAACAAAACTCATCTCAGAAGAGGATCTGAATTAATAAGATCAAACG
GTAATAAGGATCCAGCTCGAATTC

Fig.8.1(2)

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M K Y L L P T

AAGCTTGCATGCAAATTCTATTTCAAGGAGACAGTCATAATGAAATACCTATTGCCTACG

pelB A A A G L L L L A A Q P A M A Q V Q L Q

LEADER GCAGCCCTGGATTGTTATTACTCGCTGCCCAACCAGCGATGGCCCAGGTGCAGCTGCAG

S S G G D L V K P G G S L T L S C A T S

GAGTCAGGGGGAGACTTAGTGAAGCCTGGAGGGTCCCTGACACTCTCCTGTGCAACCTCT

VH4715 G T T P E S Y A F S W V R O T S D R S L

GGATTCACTTTCACTAGTTATGCCTTTTCTTGGGTCCGCCAGACCTCAGACAAGAGTCTG

E W V A T I S S T D T Y T Y Y S D N V T

GAGTGGGTGCAACCATCAGTAGTACTGATACTTATACCTATTATTTCAGACAATGTGAAG

G R F T I S R D N G K N T L Y L Q M S S

GGGCGCTTCACCATCTCCAGAGACAATGGCAAGAACCCTGTACCTGCAAATGAGCAGT

L K S E D T A V Y Y C A R H G Y Y G K G

CTGAAGTCTGAGGACACAGCCGTGTATTACTGTGCAAGACATGGGTACTATGGTAAAGGC

Y F D Y W G Q Q T T V T V S S G G G G S

TATTTTGACTACTGGGGCCAAGGGACCACGGTCACCGTCTCCTCAGGTGGAGGCGGTTC

LINKER G G G G S G G G G S A G S A Q V Q L Q Q

GGCGGAGGTGGCTCTGGCGGTGGCGGATCGGCCGGTTCGGCCAGGTCCAGCTGCAACAG

S G P E L V K P G A S V K M S C K A S G

TCAGGACCTGAGCTGGTAAAGCCTGGGCTTCAGTGAAGATGTCTGCAAGGCTTCTGGA

Y T F T S Y V M H W V K Q K P G Q Q L E

TACACATTCAGTATGTTATGCACTGGGTGAAACAGAAGCCTGGGCAGGGCCTTGAG

W I G Y I Y P Y N D G T K Y N P K F K G

TGGATTGGATATATTTATCCTTACAATGATGGTACTAAGTACAATGAGAAGTTCAAAGGC

VH3418 K A T L T S D K S S S T A Y M E L S S L

AAGGCCACACTGACTTCAGACAAATCCTCCAGCACAGCCTACATGGAGCTCAGCAGCCTG

T S E D S A V Y Y C S R R F D Y W G Q G

ACCTCTGAGGACTCTGCGGTCTATTACTGTTCAAGACGCTTTGACTACTGGGGCCAAGGG

T T V T V S S

ACCACCGTCACCGTCTCCTCATAATAAGCTAGCGGAGCTGCATGCAAATTCTATTTCAAG

pelB M K Y L L P T A A A G L L L L A

LEADER GAGACAGTCATAATGAAATACCTATTGCCTACGGCAGCCGCTGGATTGTTATTACTCGCT

A Q P A M A D I E L T Q S P S S M Y A S

GCCCAACCAGCGATGGCCGACATCGAGCTCAGCCAGTCTCCATCTTCCATGTATGCATCT

L G E R I T I T C K A S Q D I N T Y L T

CTAGGAGAGAGAATCACTATCACTTGCAAGGCGAGTCAGGACATTAATACCTATTTAACC

VH3418 W F Q Q K P G K S P F T L I Y R A N R L

TGGTTCAGCAGAAACCAGGGAAATCTCCCAAGACCCTGATCTATCGTGCAAACAGATTG

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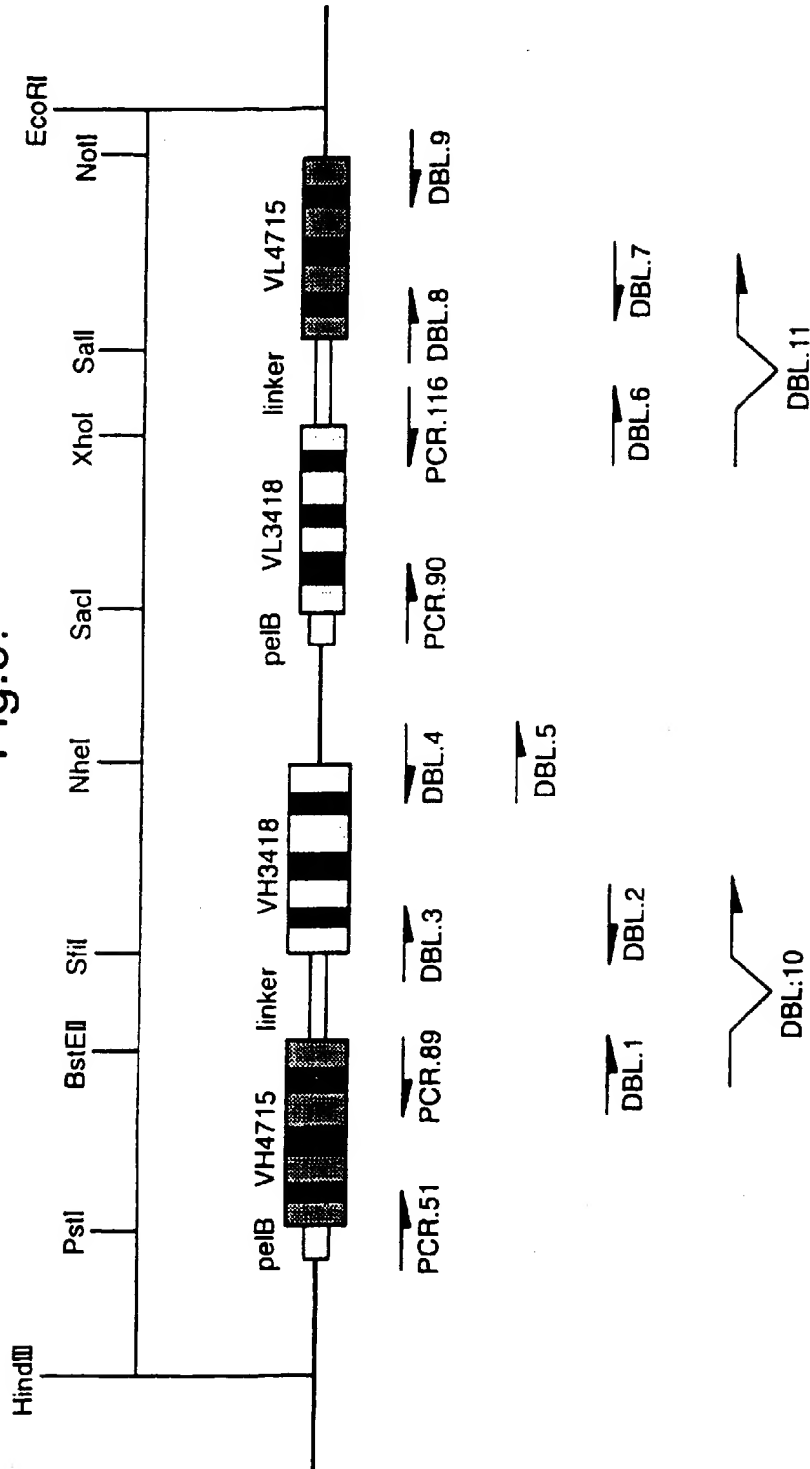
Fig.8. 2(2)

LINKER

VL4715

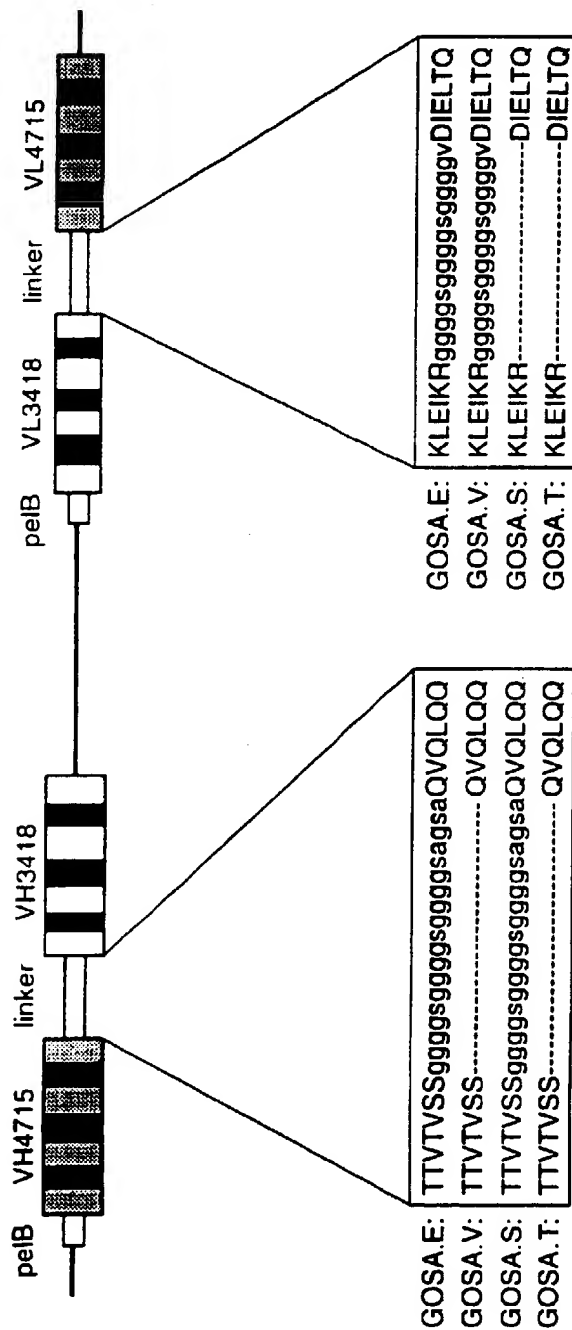
L D G V P S R F S G S G S G Q D Y S L T
 CTAGATGGGGTCCCATCAAGGTTTCAGTGGCAGTGGATCTGGGCAAGATTATCTCTCACC
 I S S L D Y E D M G I Y Y C L O Y D E L
 ATCAGCAGCCTGGACTATGAAGATATGGGAATTATTATTGTCTACAATATGATGAGTTG
 Y T F G G G T K L E I K R G G G G S G G
 TACACGTTCCGAGGGGGACCAAGCTCGAGATCAAACGGGGTGGAGGCGGTTTCAGGCGGA
 G G S G G G G V D I E L T Q S P F S L T
 GGTGGCTCTGGCGGTGGCGGAGTCGACATCGAACTCACTCAGTCTCCATTCTCCCTGACT
 V T A G E K V T M N C K S G Q S L L N S
 GTGACAGCAGGAGAGAAGGTCACTATGAATTGCAAGTCCGGTCAGAGTCTGTAAACAGT
 V N Q R N Y L T W Y Q Q K P G Q P P K L
 GTAAATCAGAGGAACTACTTGACCTGGTACCAGCAGAAGCCAGGGCAGCCTCCTAAACTG
 L I Y W A S T R E S G V P D R F T A S G
 TTGATCTACTGGGCATCCACTAGGGAATCTGGAGTCCCTGATCGCTTCACAGCCAGTGGGA
 S G T D F T L T I S S V Q A E D L A V Y
 TCTGGAACAGATTTCACCTCTCACCATCAGCAGTGTGCAGGCTGAAGACCTGGCAGTTTAT
 Y C Q N D Y T Y P F T F G G G T K L E I
 TACTGTCAATGATTATACTTATCCGTTACGTTCCGAGGGGGGACCAAGCTCGAAATC
 K R
 AAACGGTAATAAGCGGCCCTGAATTC

Fig.9.



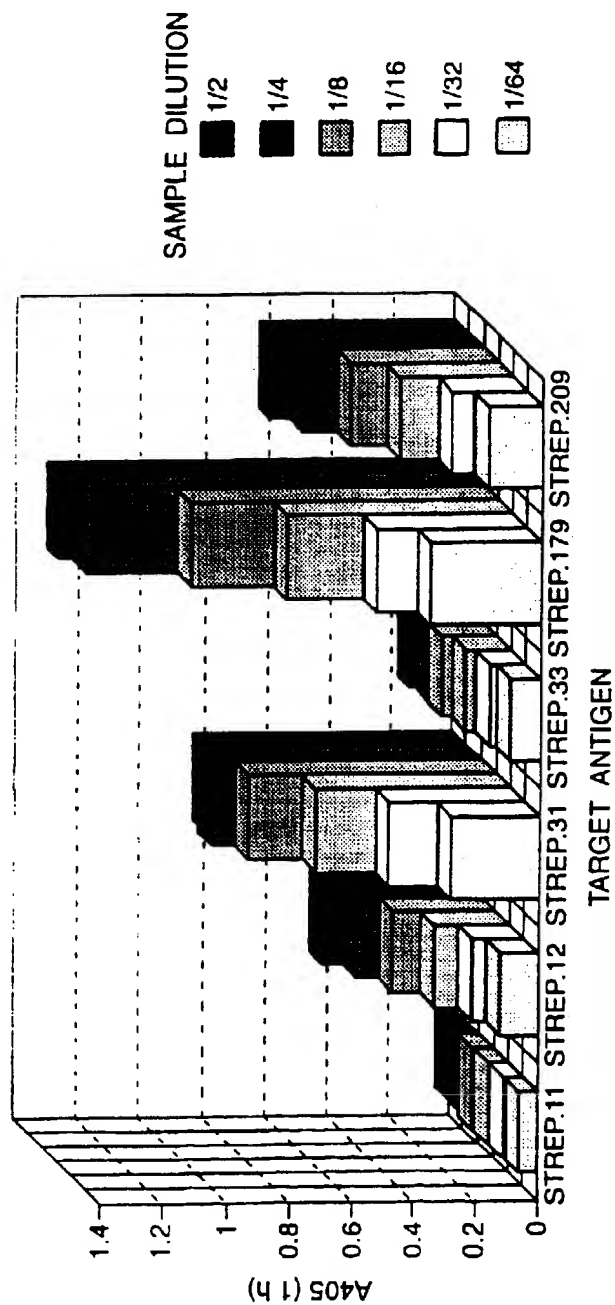
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Fig.10.



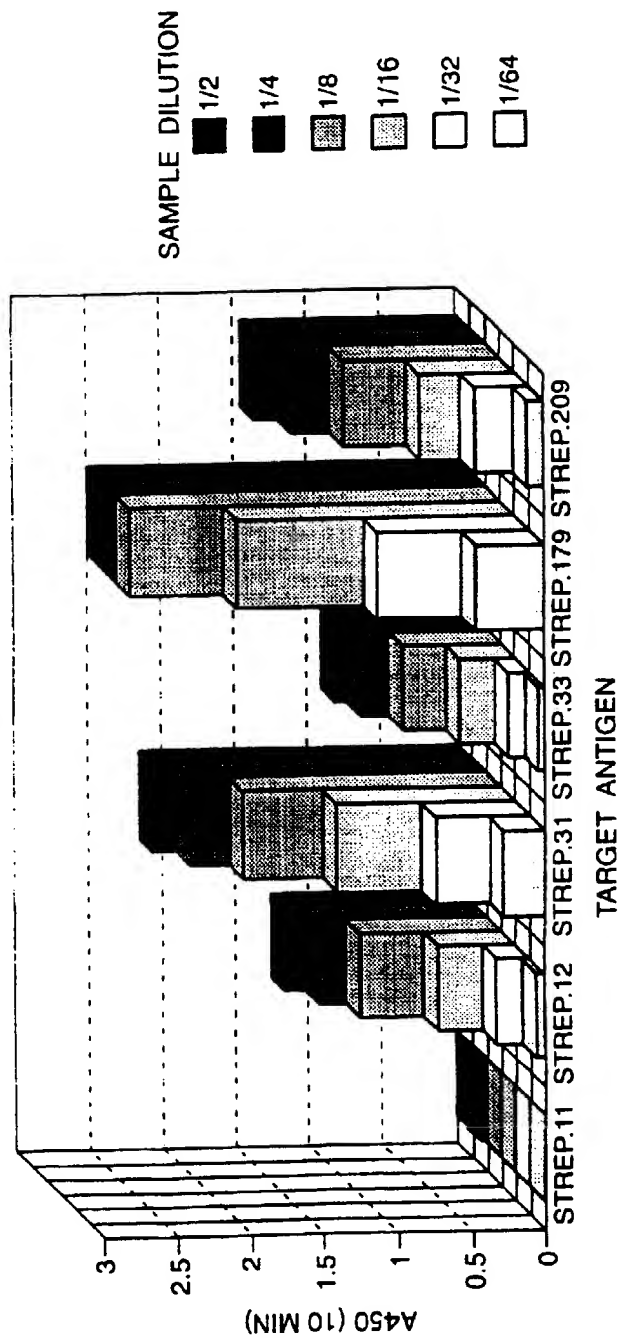
12/45

Fig.11.



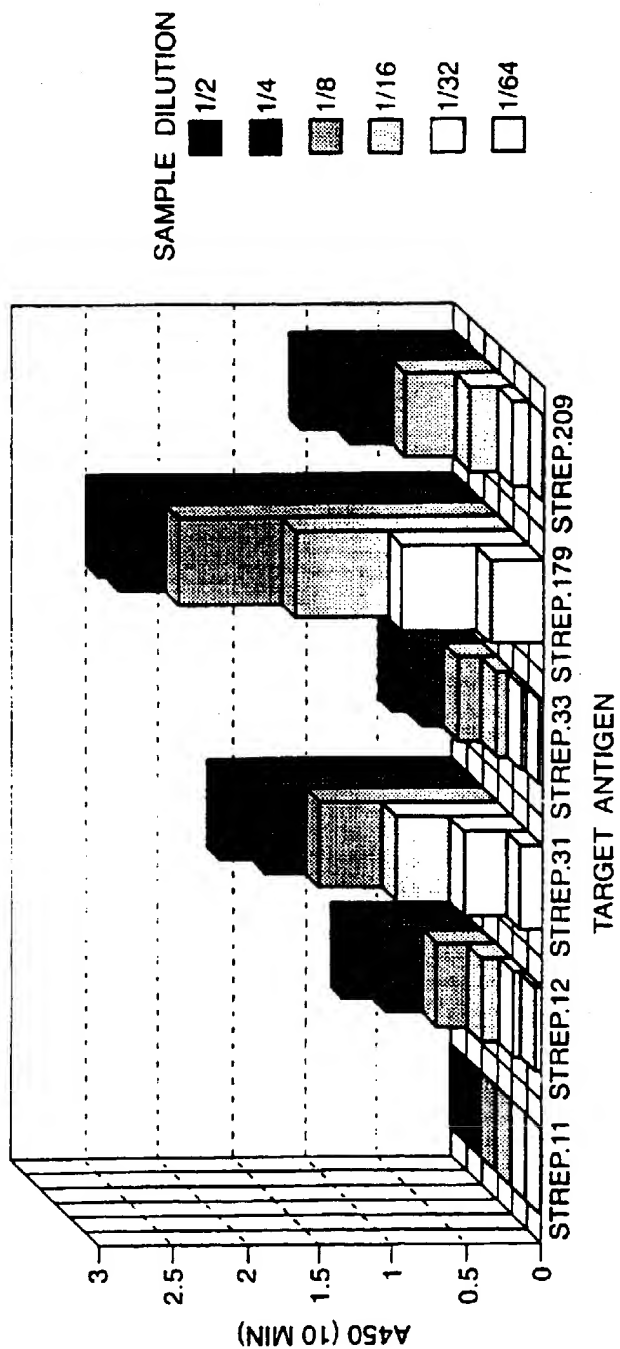
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Fig.12.



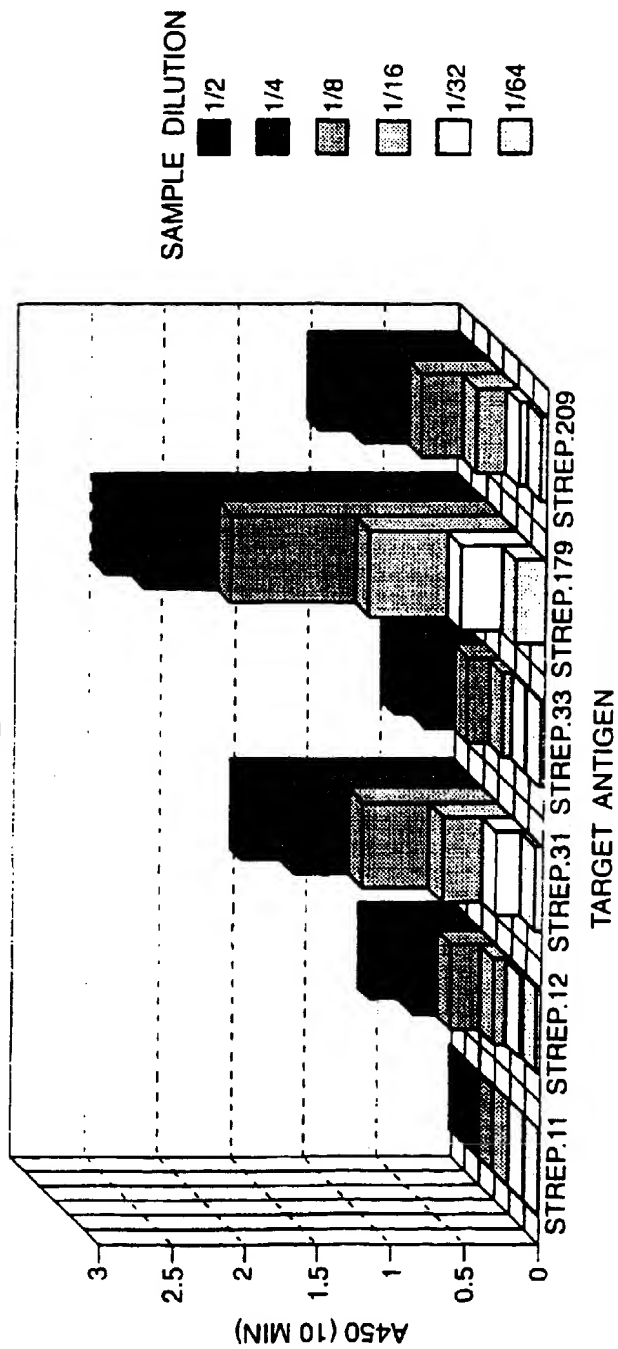
14/45

Fig.13.



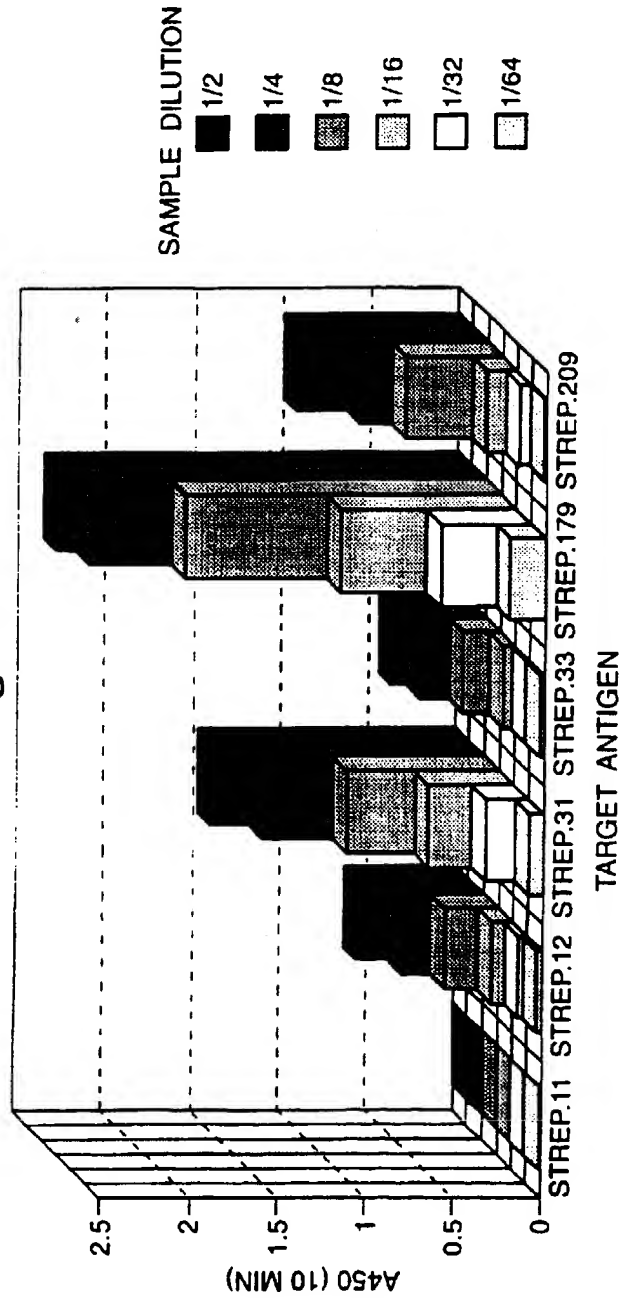
15/45

Fig.14.



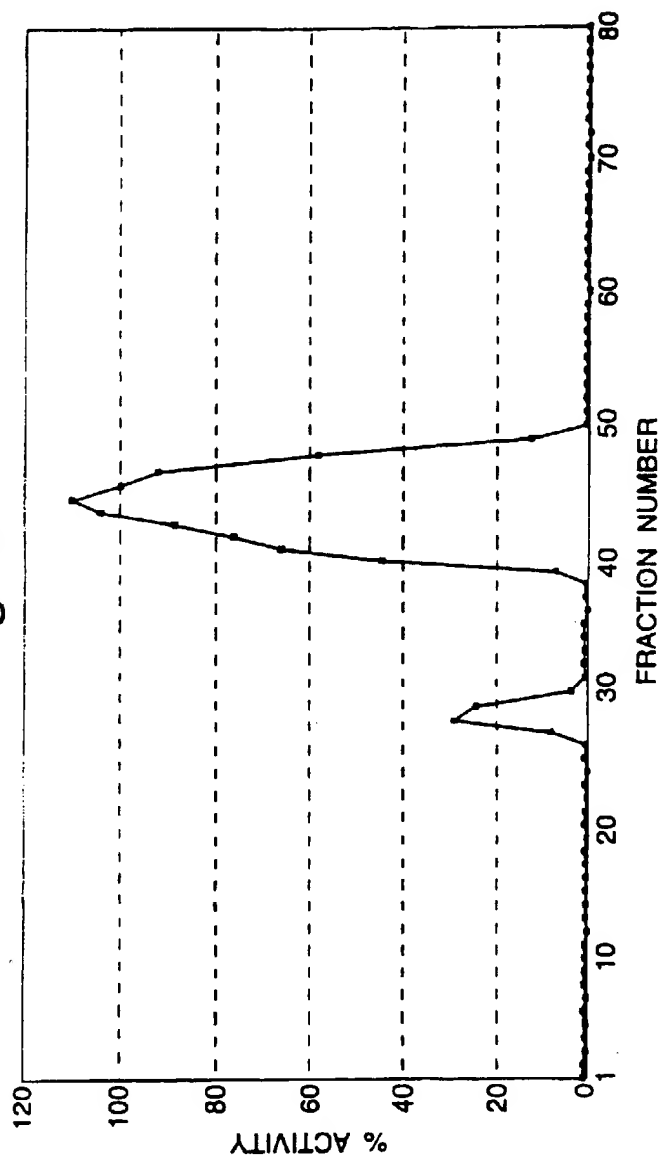
16/45

Fig.15.

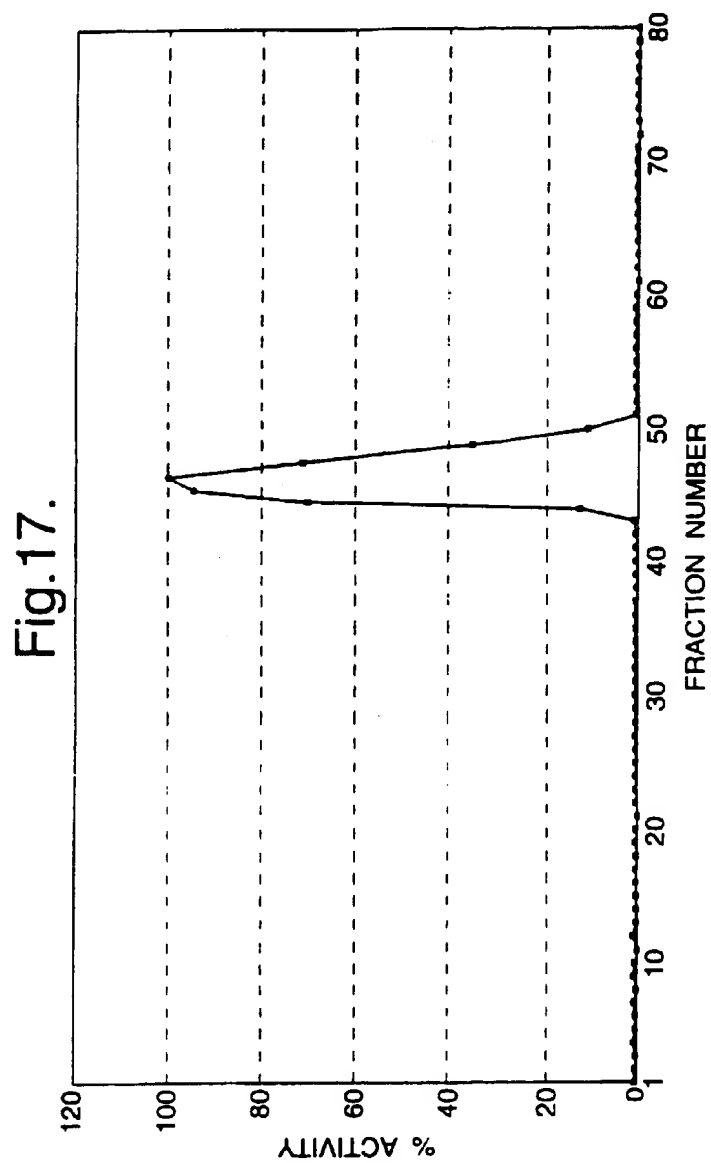


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Fig.16.



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NOT TO BE TAKEN
INTO CONSIDERATION
FOR THE PURPOSES
OF INTERNATIONAL PROCESSING

(See Section 310(d)(ii) of the Administrative Instructions)

20/45

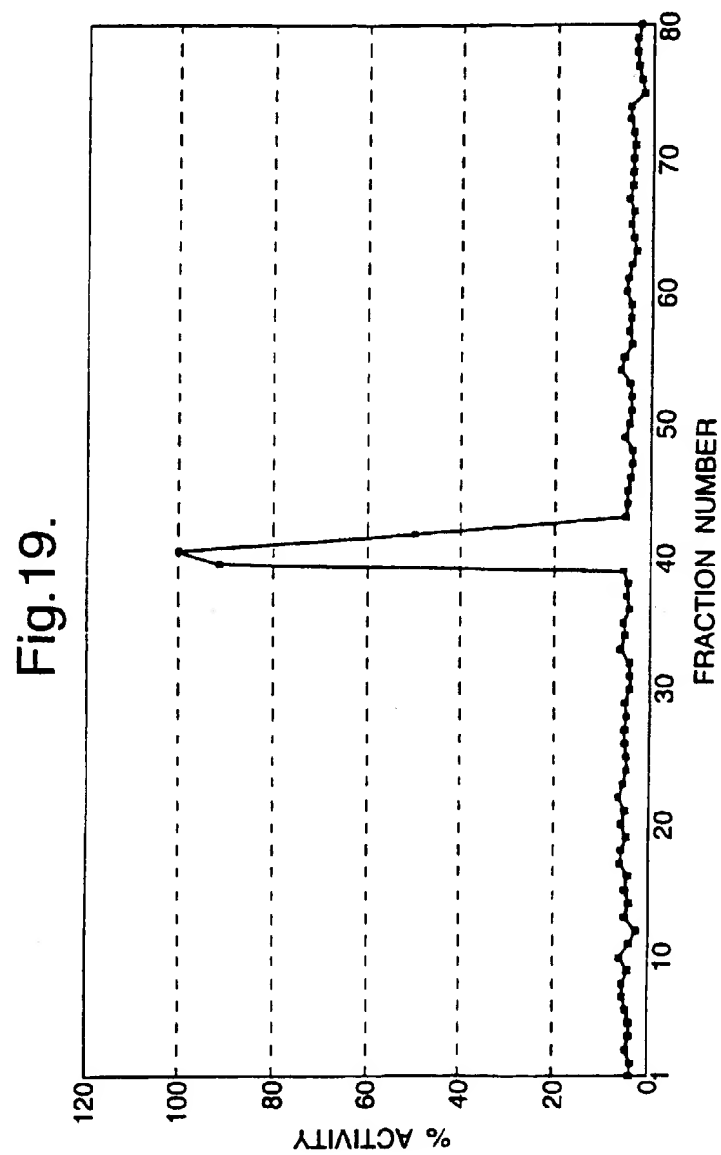


Fig.20.

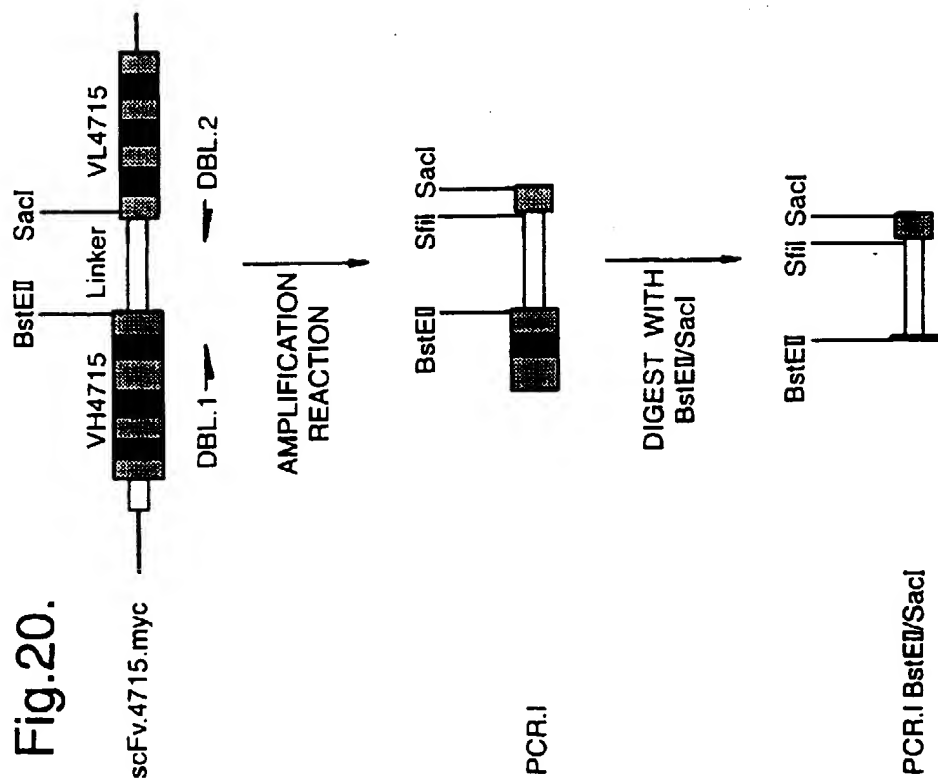
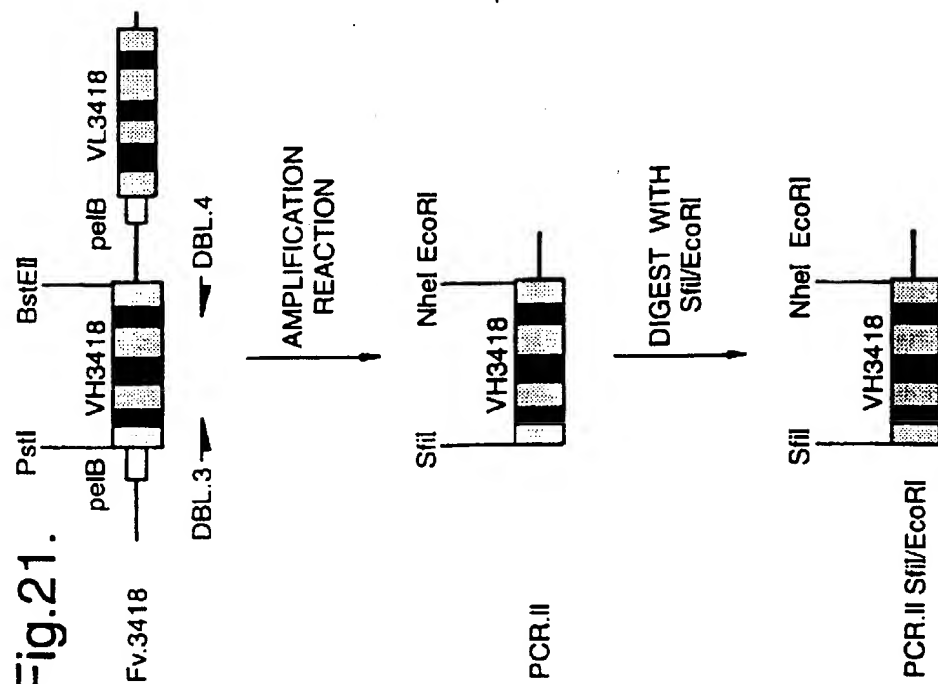
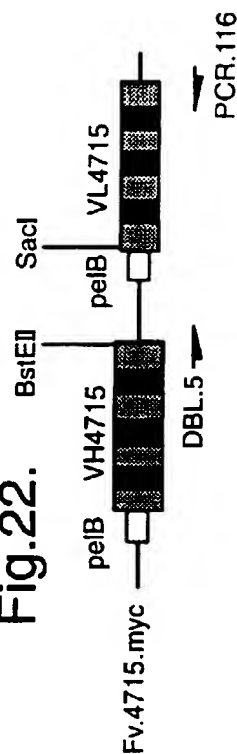


Fig.21.

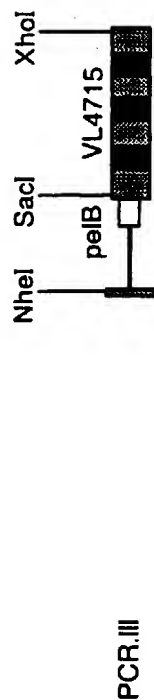


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Fig.22.



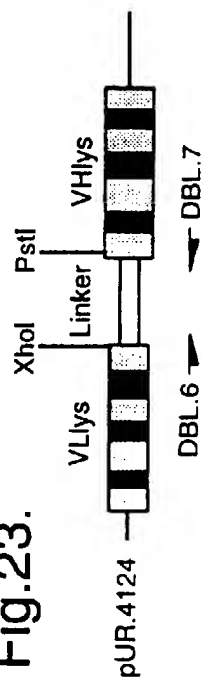
AMPLIFICATION
REACTION



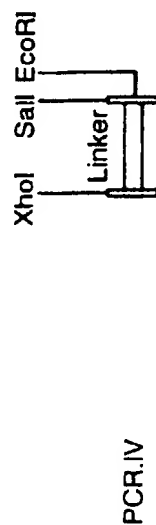
DIGEST WITH
NheI/SacI



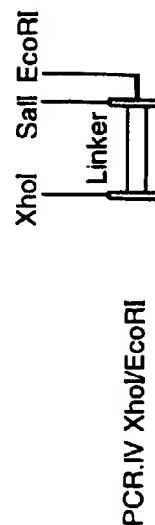
Fig.23.



AMPLIFICATION
REACTION



DIGEST WITH
XhoI/EcoRI



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Fig.24.

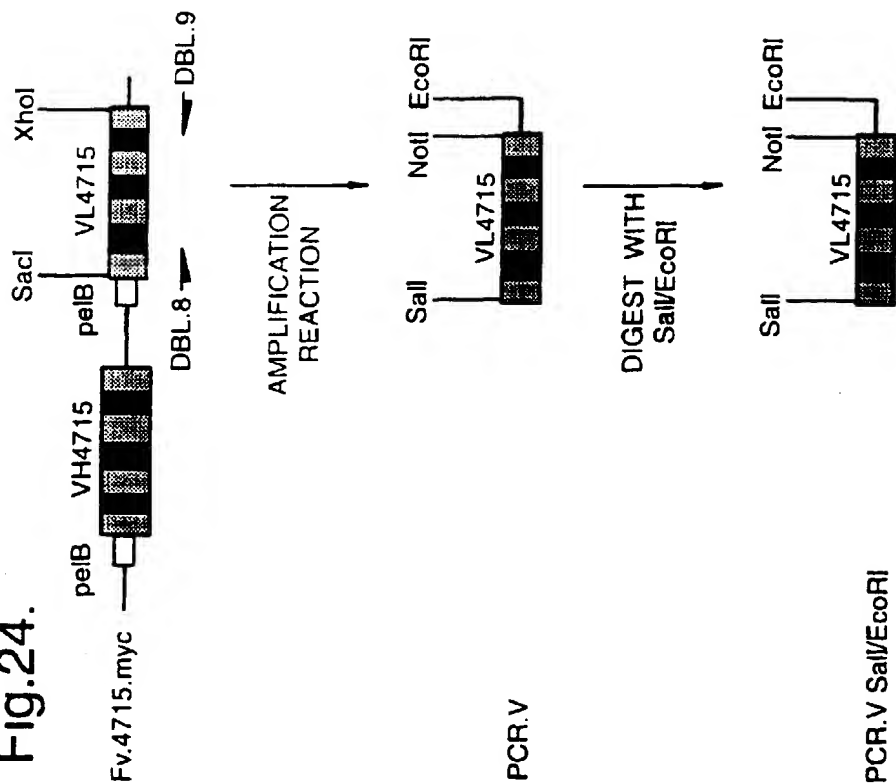


Fig.25.

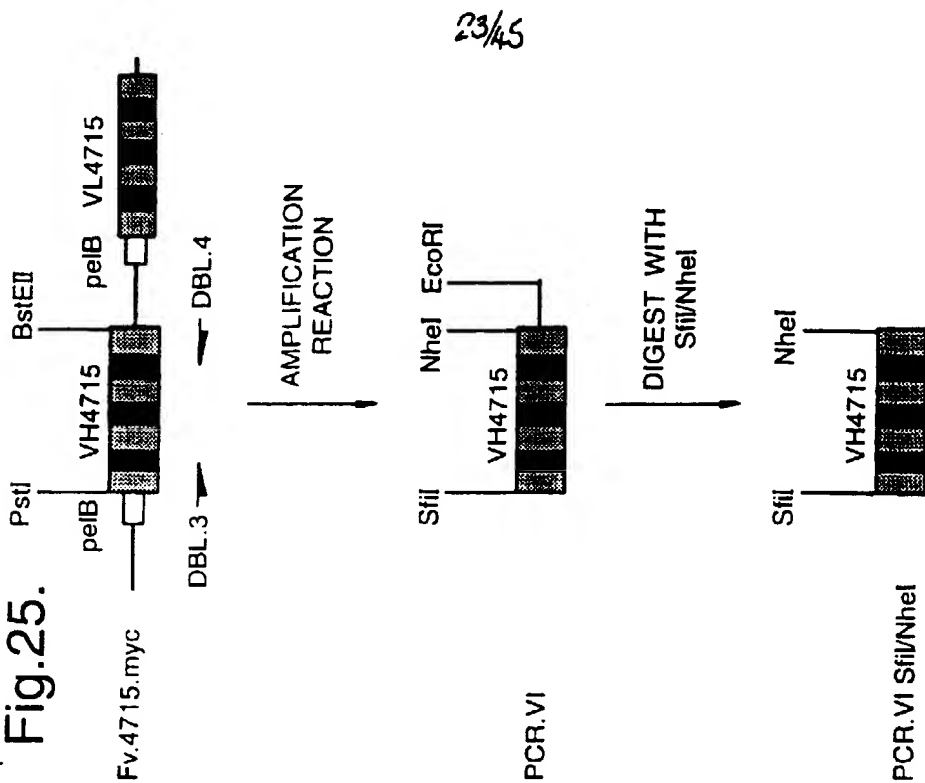


Fig.27.

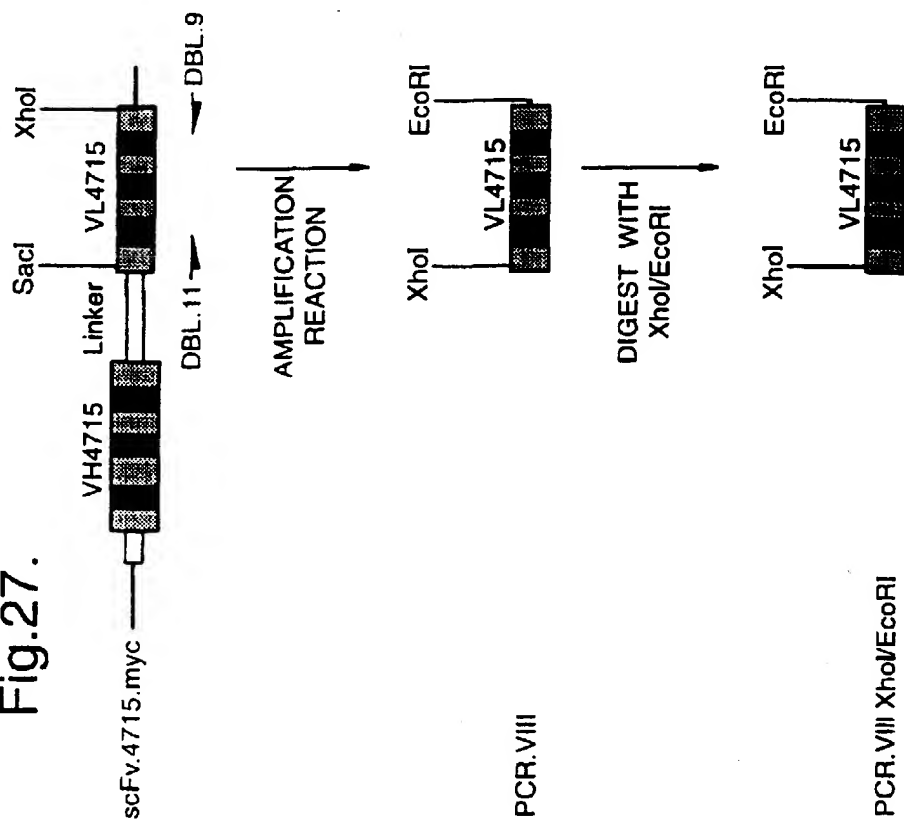
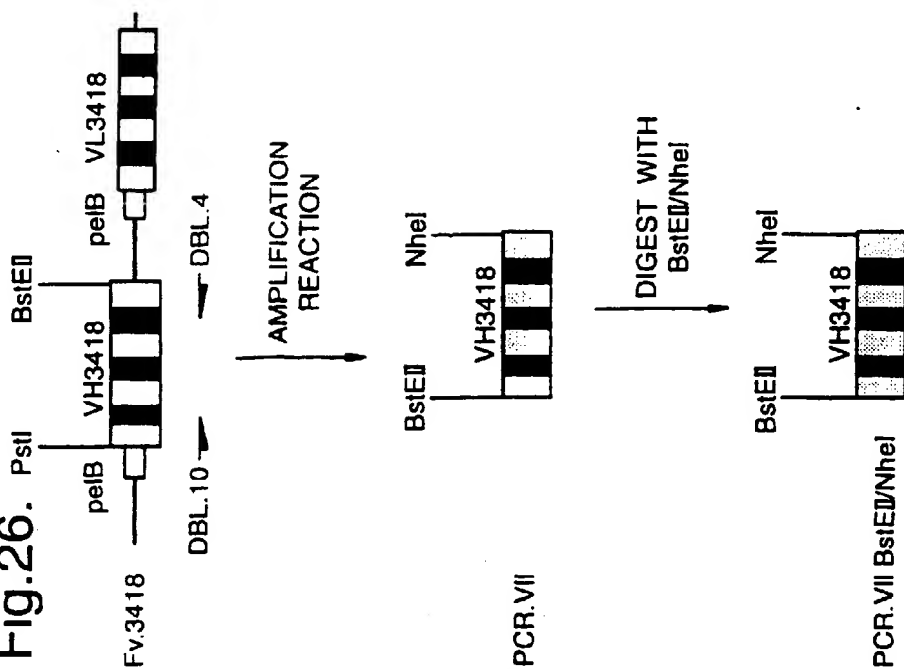
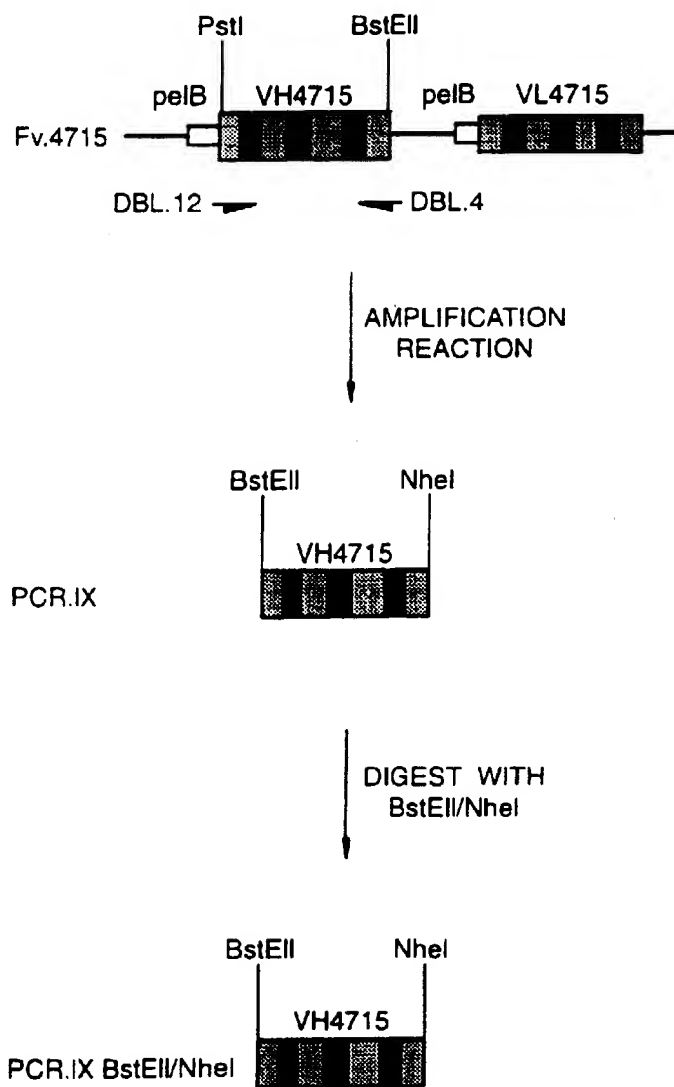


Fig.26.



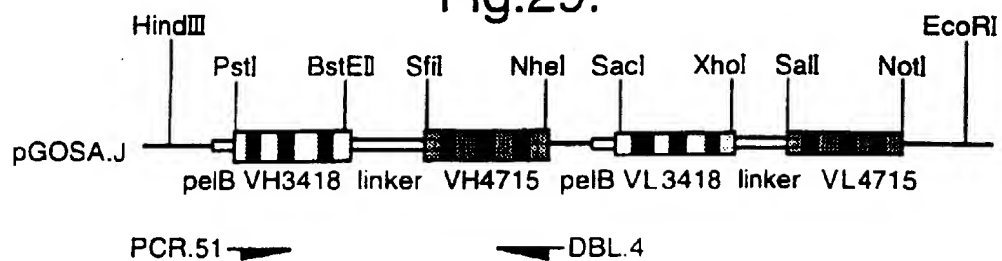
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Fig.28.



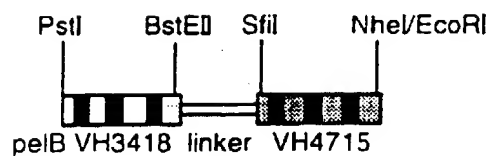
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Fig.29.



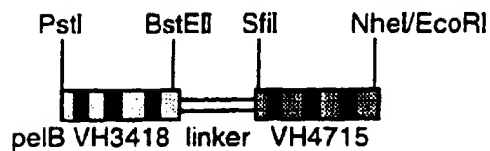
AMPLIFICATION
REACTION

PVR.X



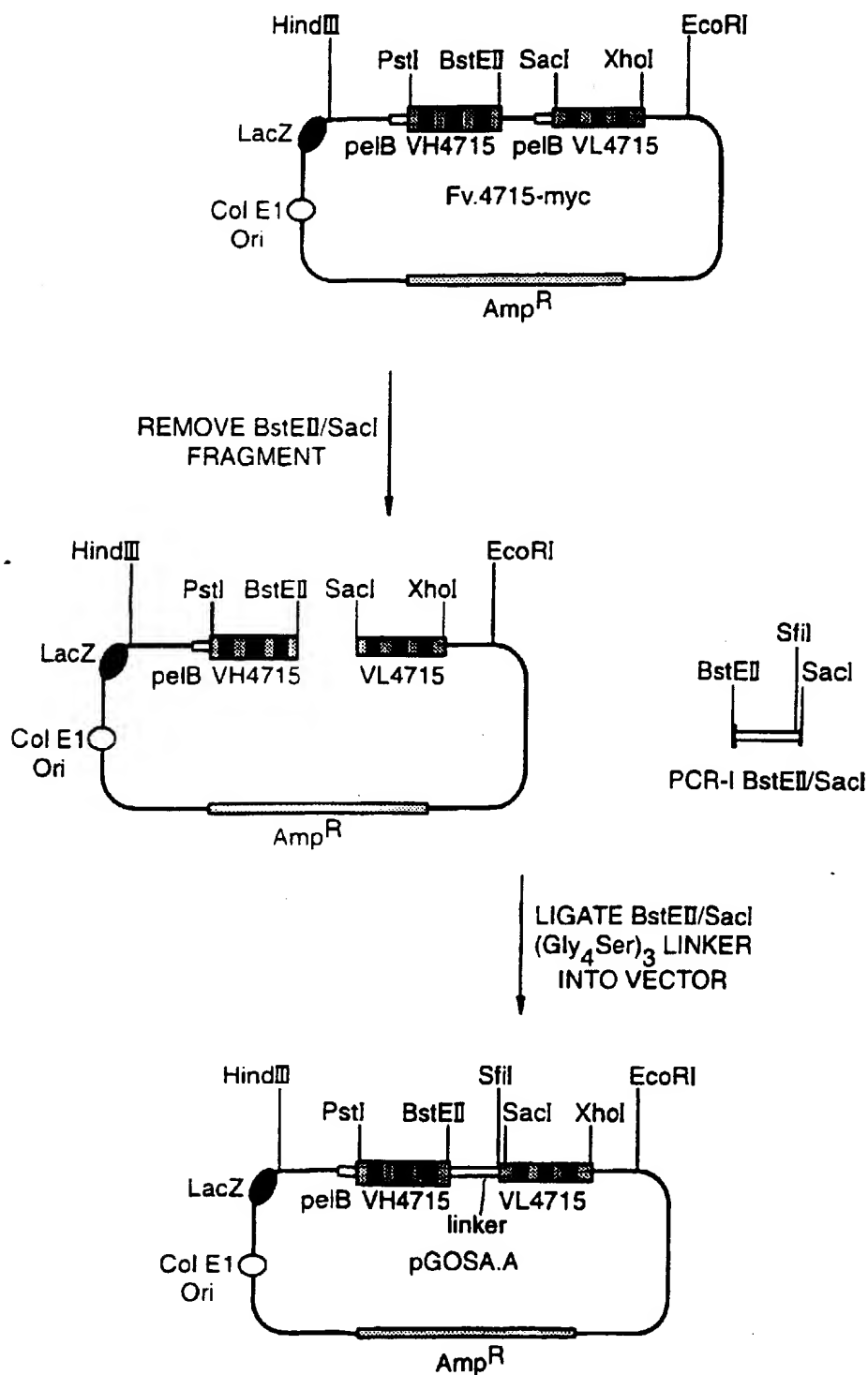
DIGEST PCR.X
WITH PstI/EcoRI

PCR.X PstI/EcoRI

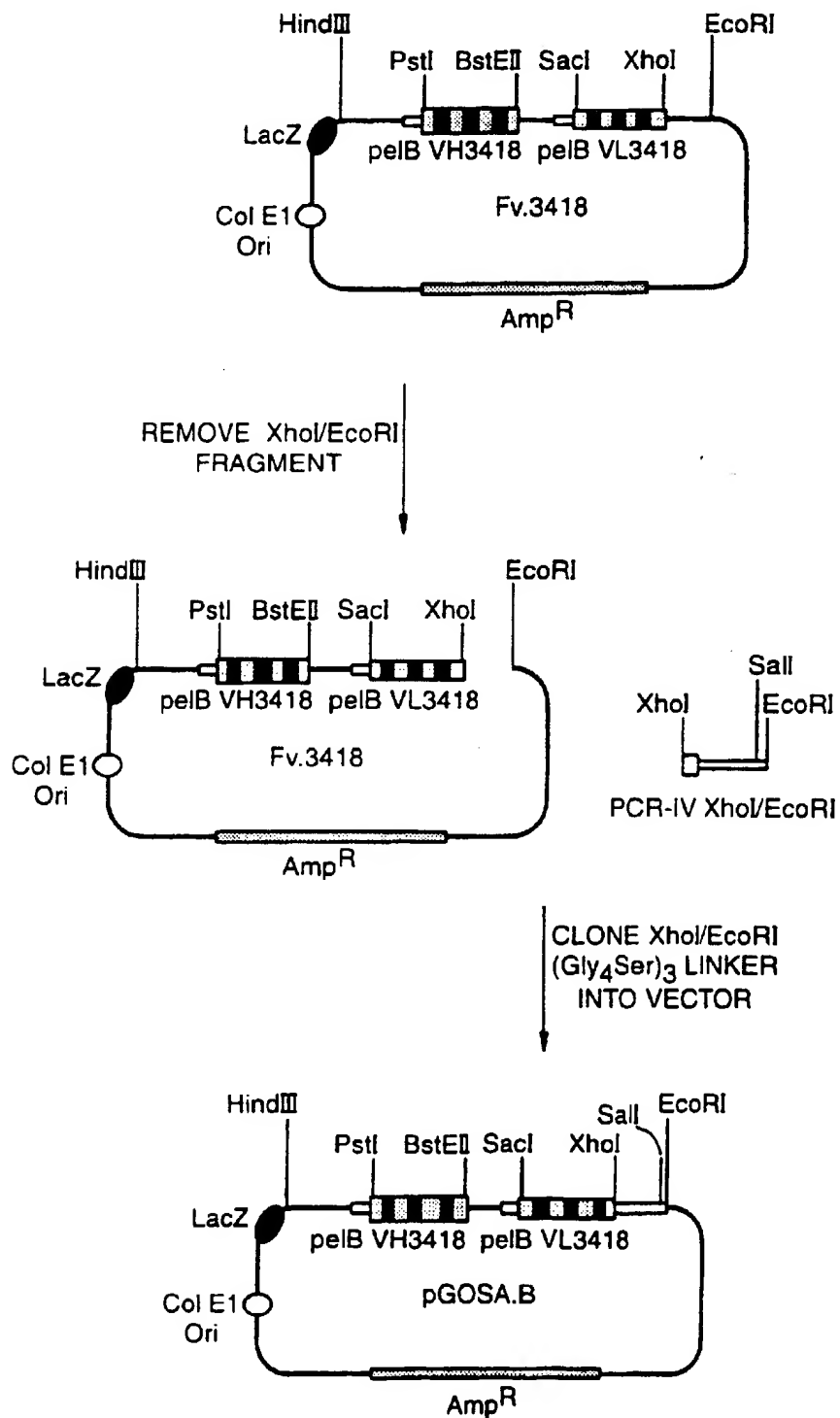


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Fig.30.

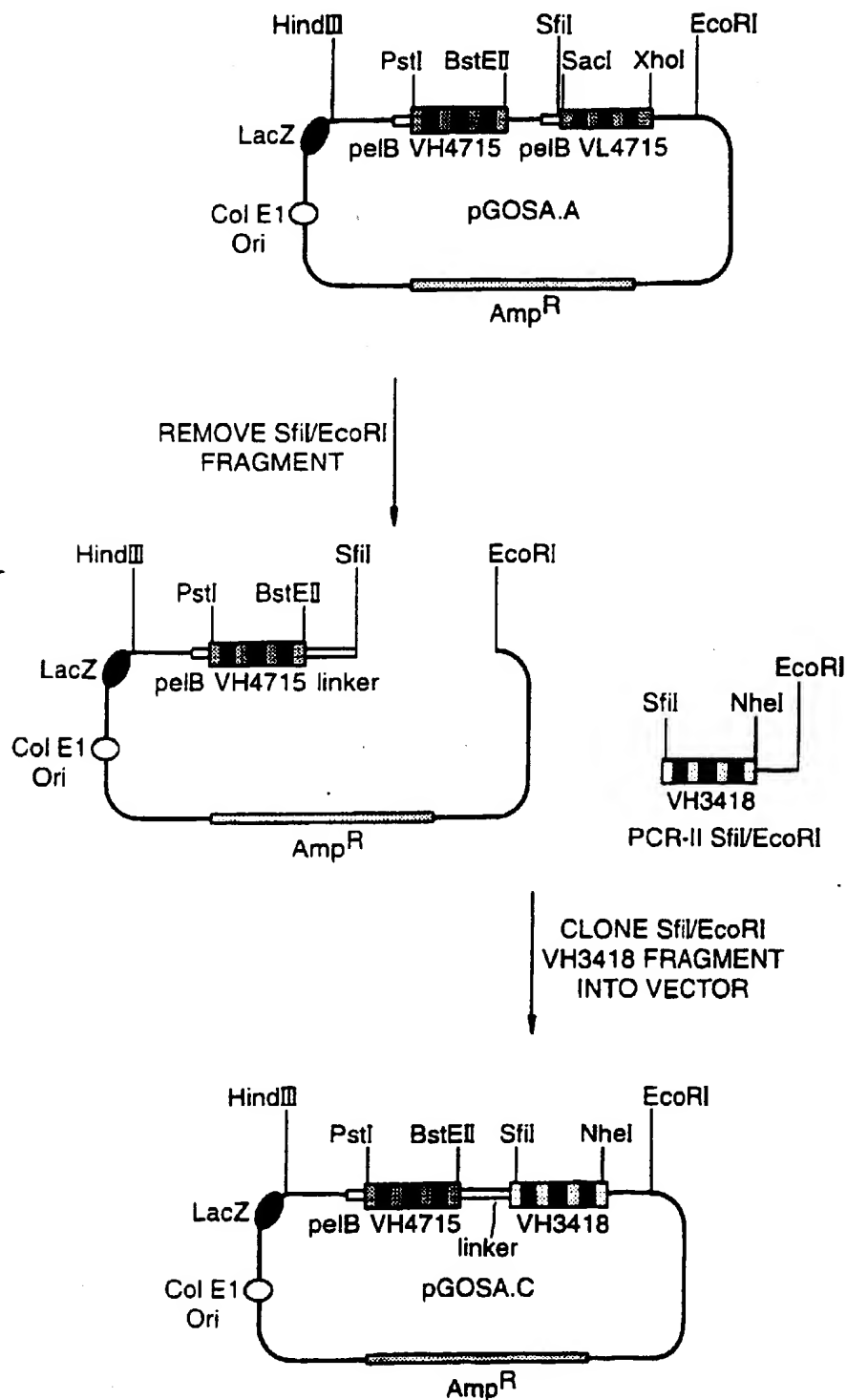


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Fig.31.



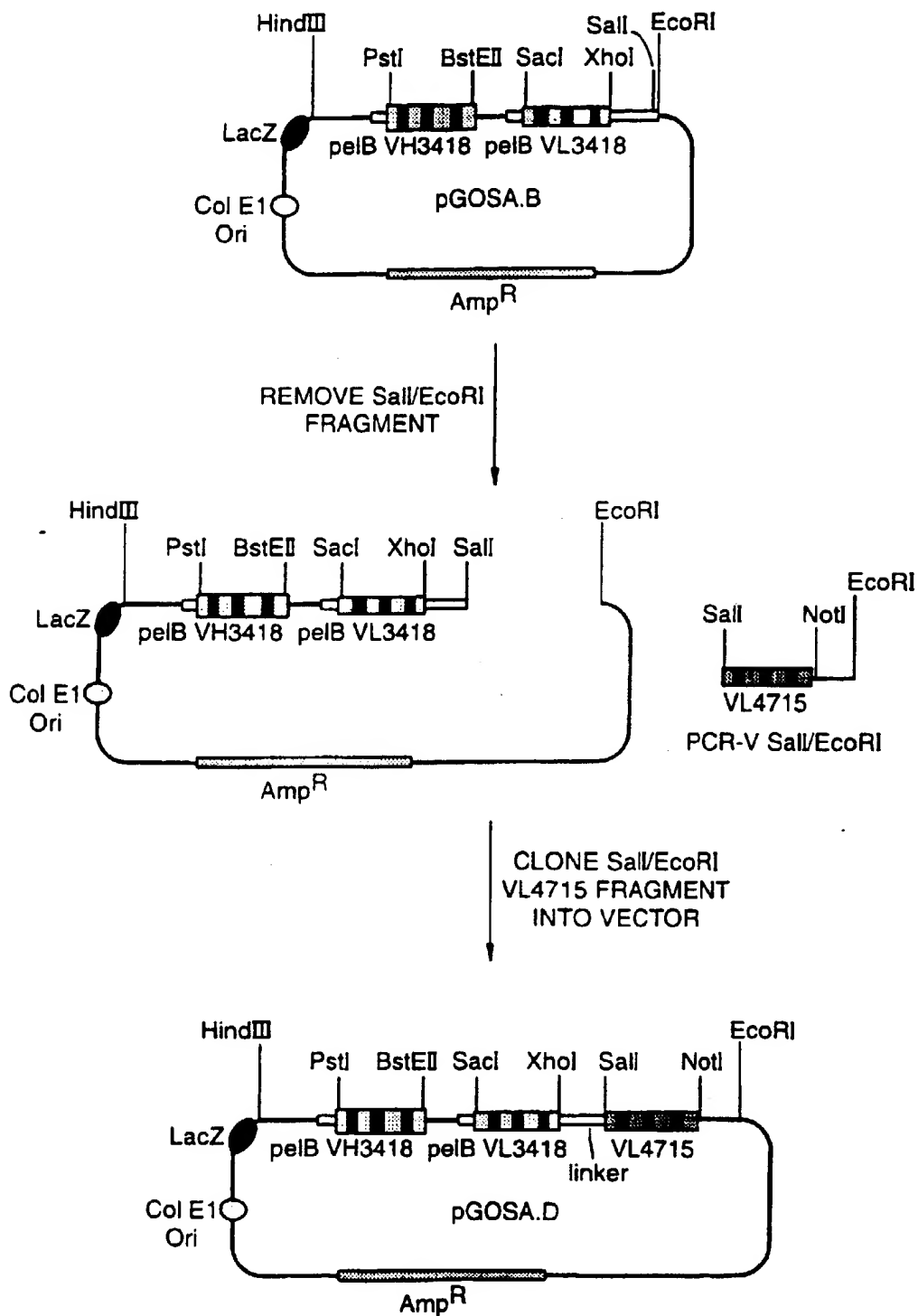
29/45

Fig.32.

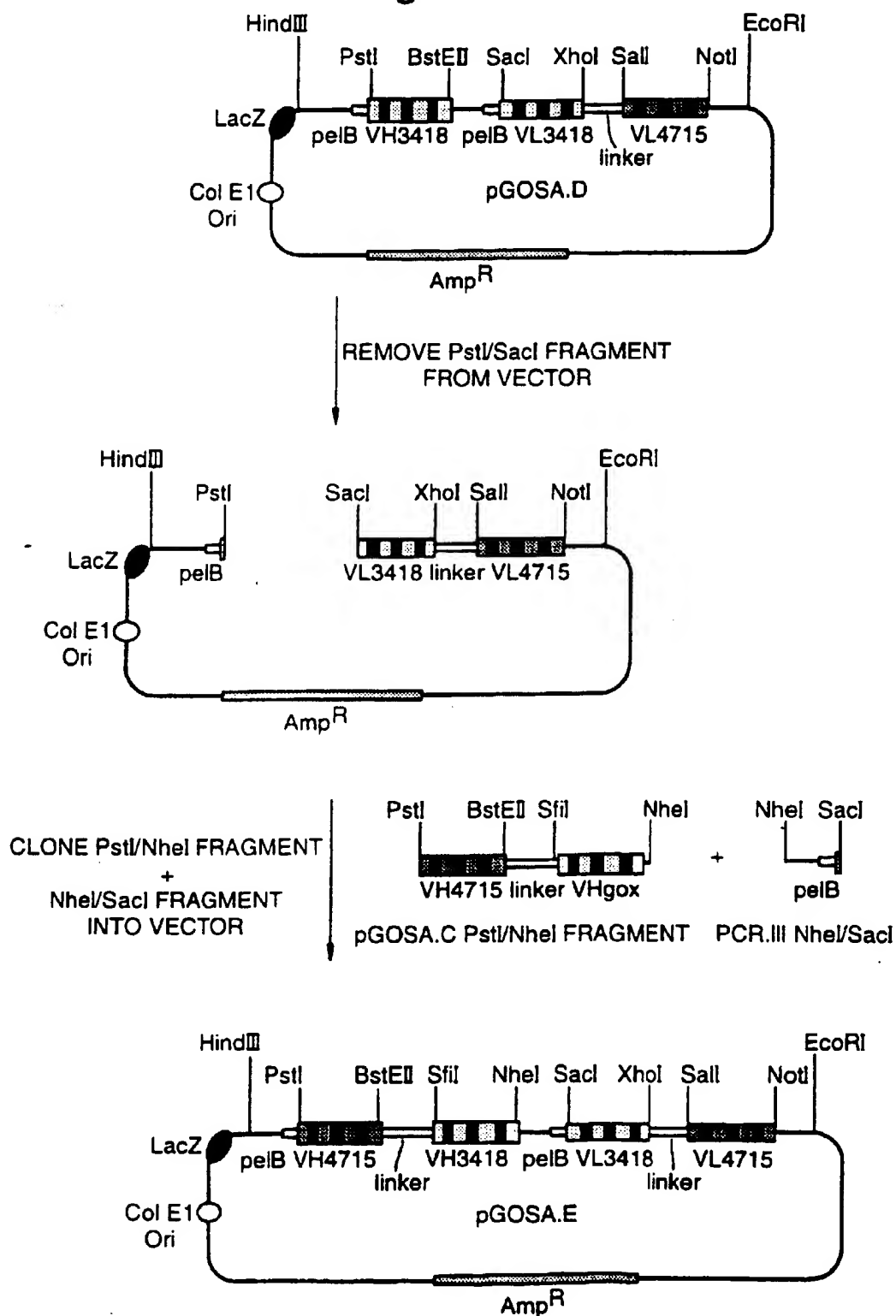


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Fig.33.



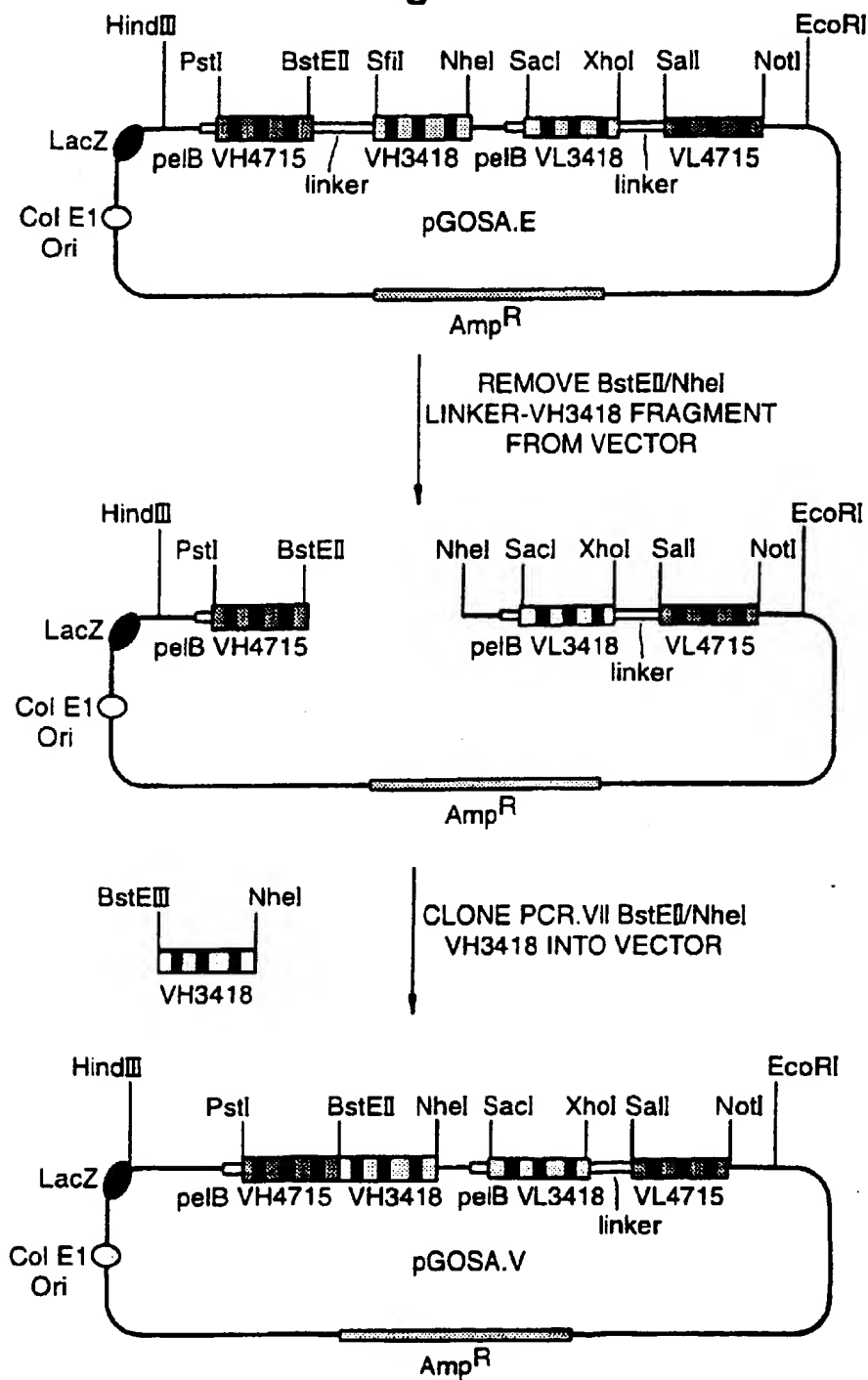
31/45
Fig.34.



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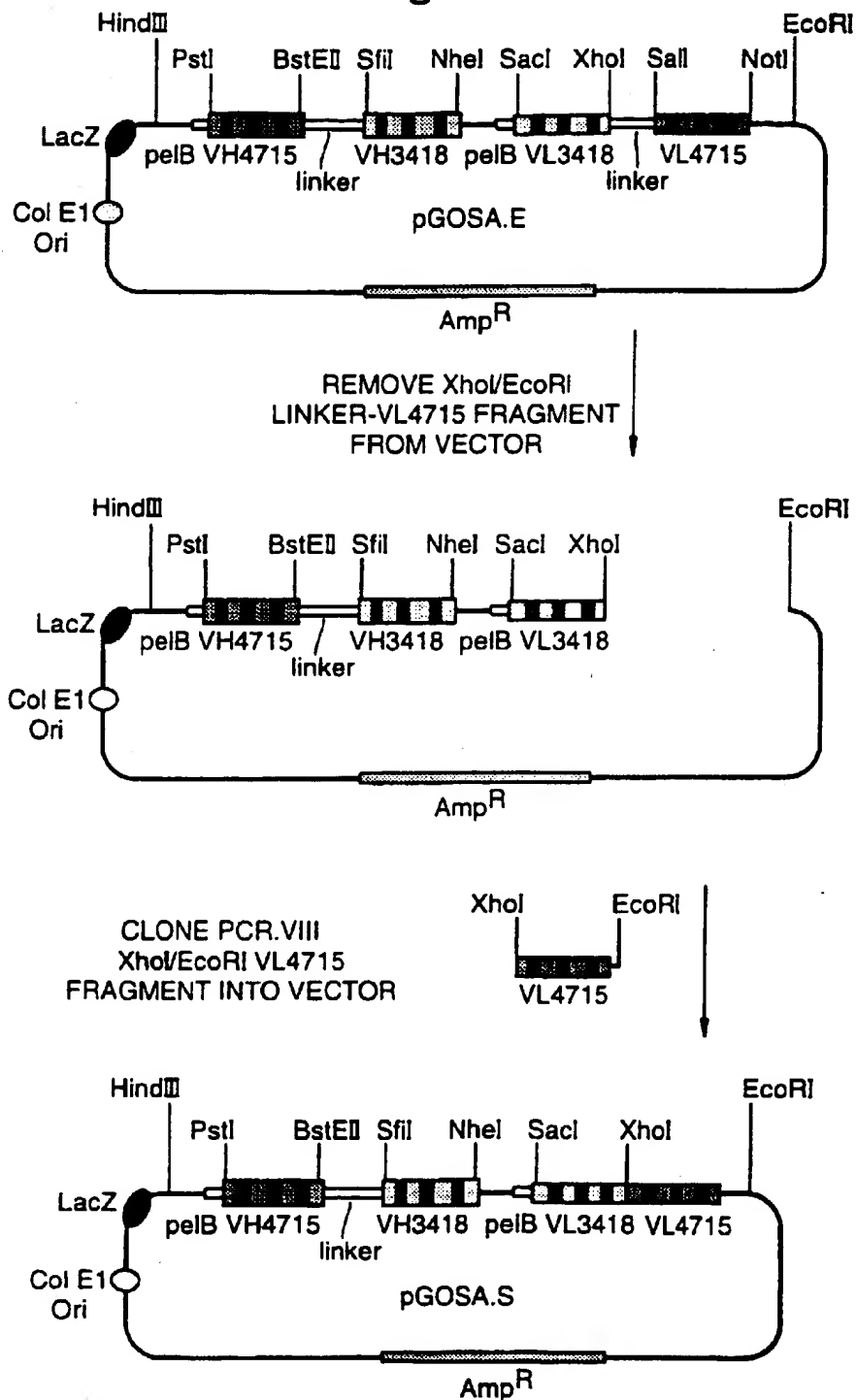
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Fig.35.

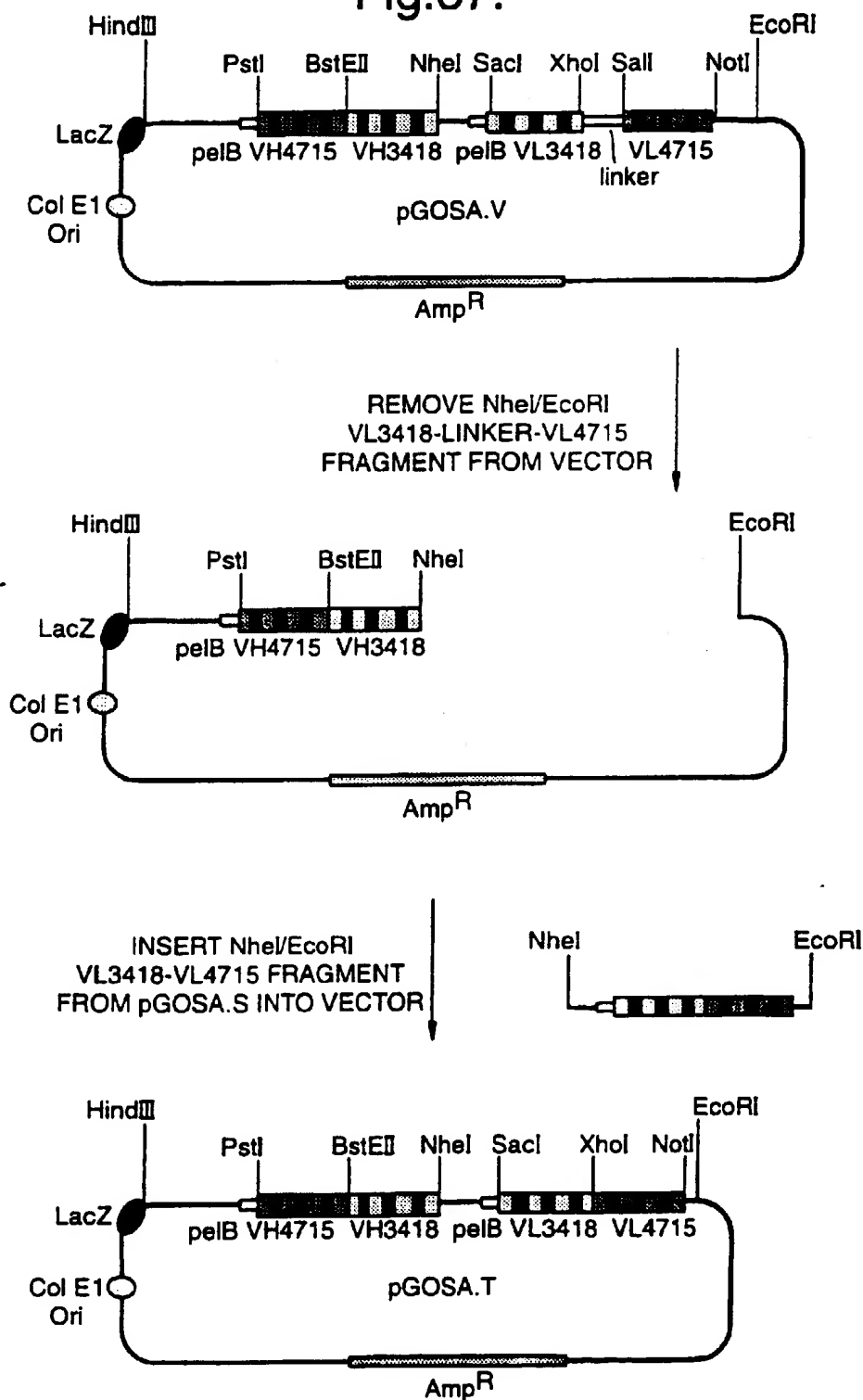


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Fig.36.

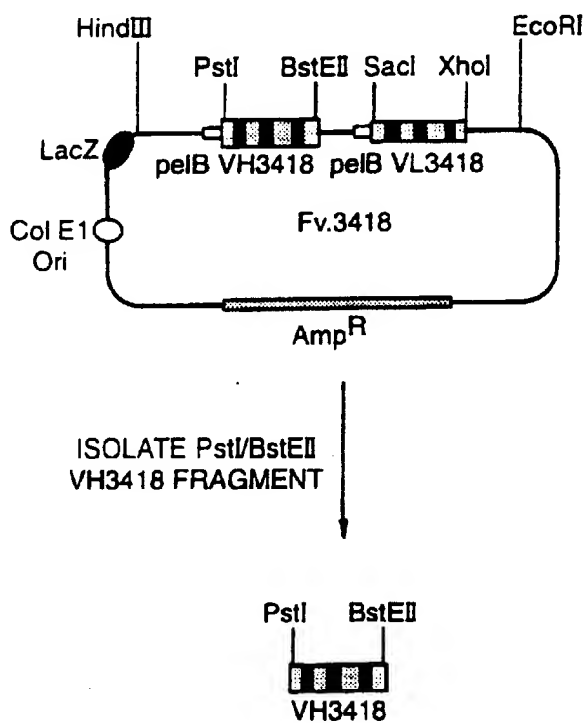


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Fig.37.



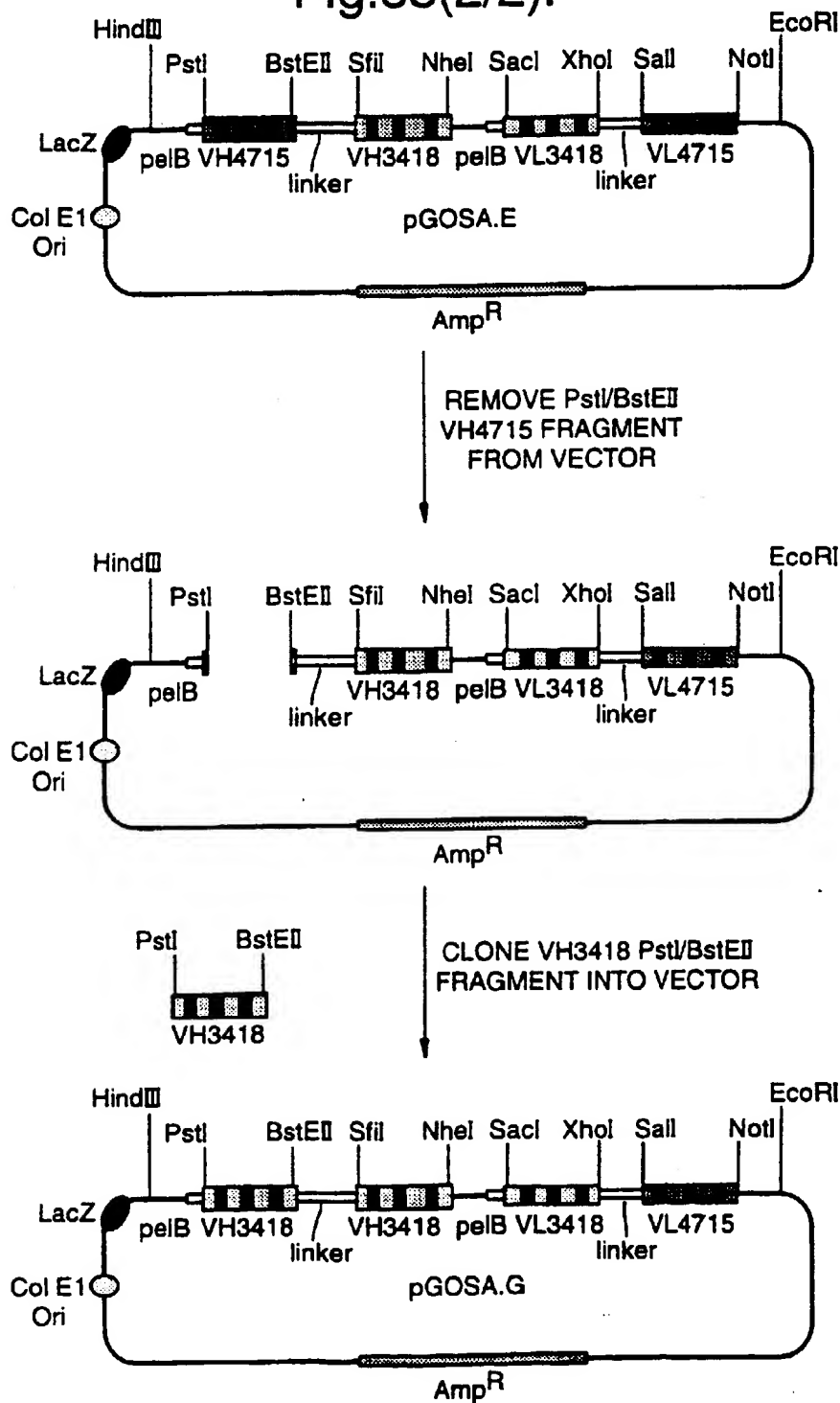
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Fig.38(1/2).



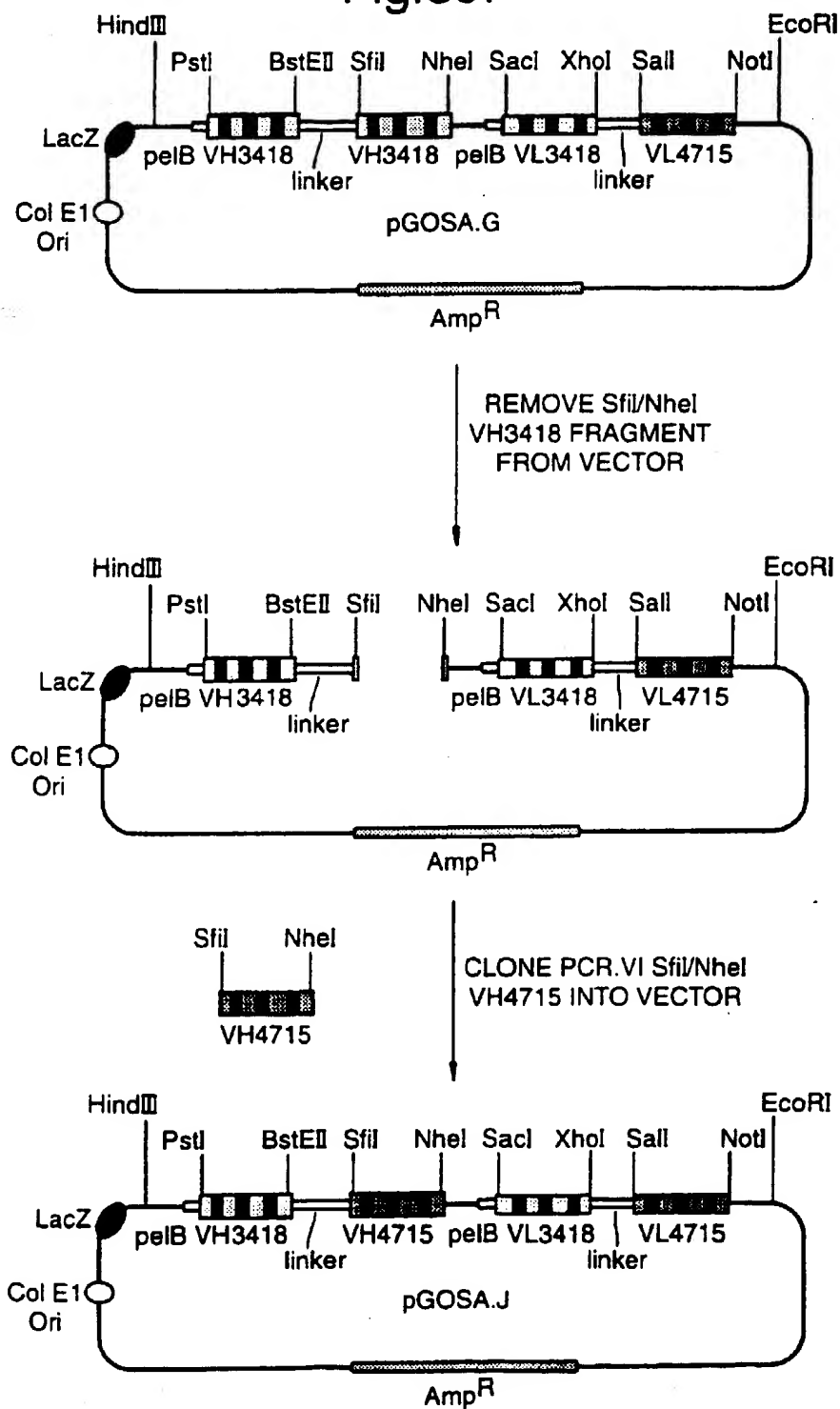
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Fig.38(2/2).



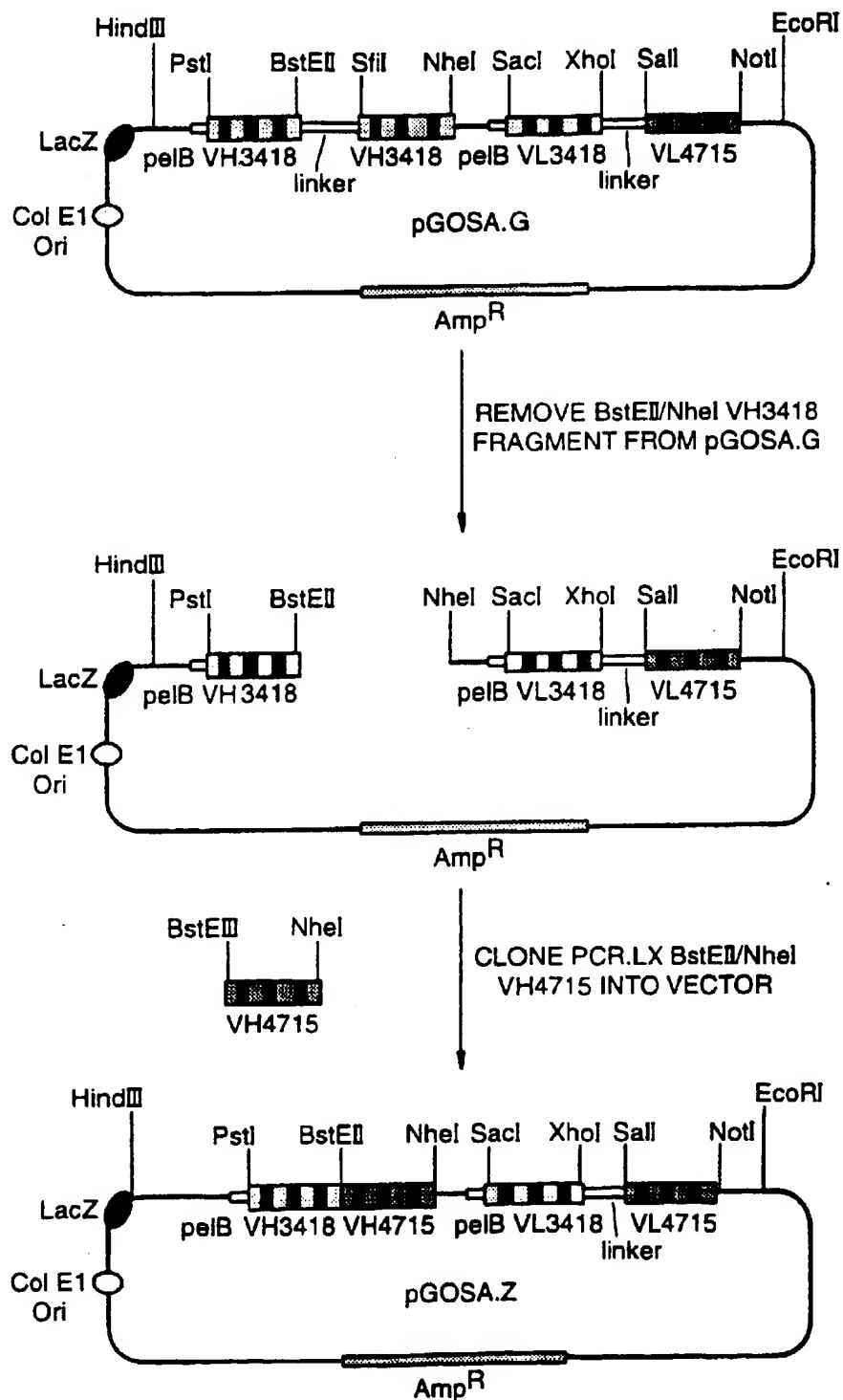
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Fig.39.



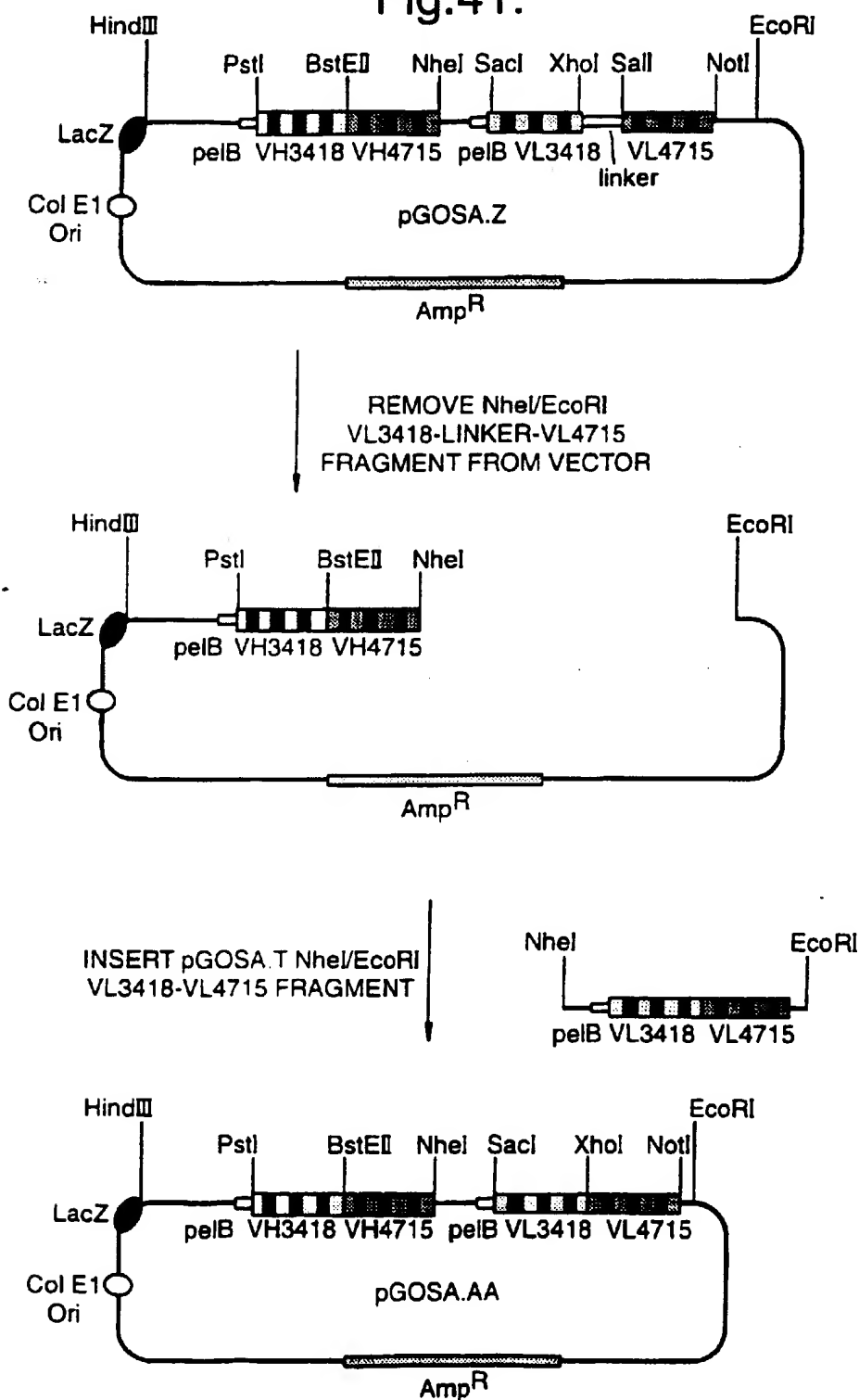
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Fig.40.

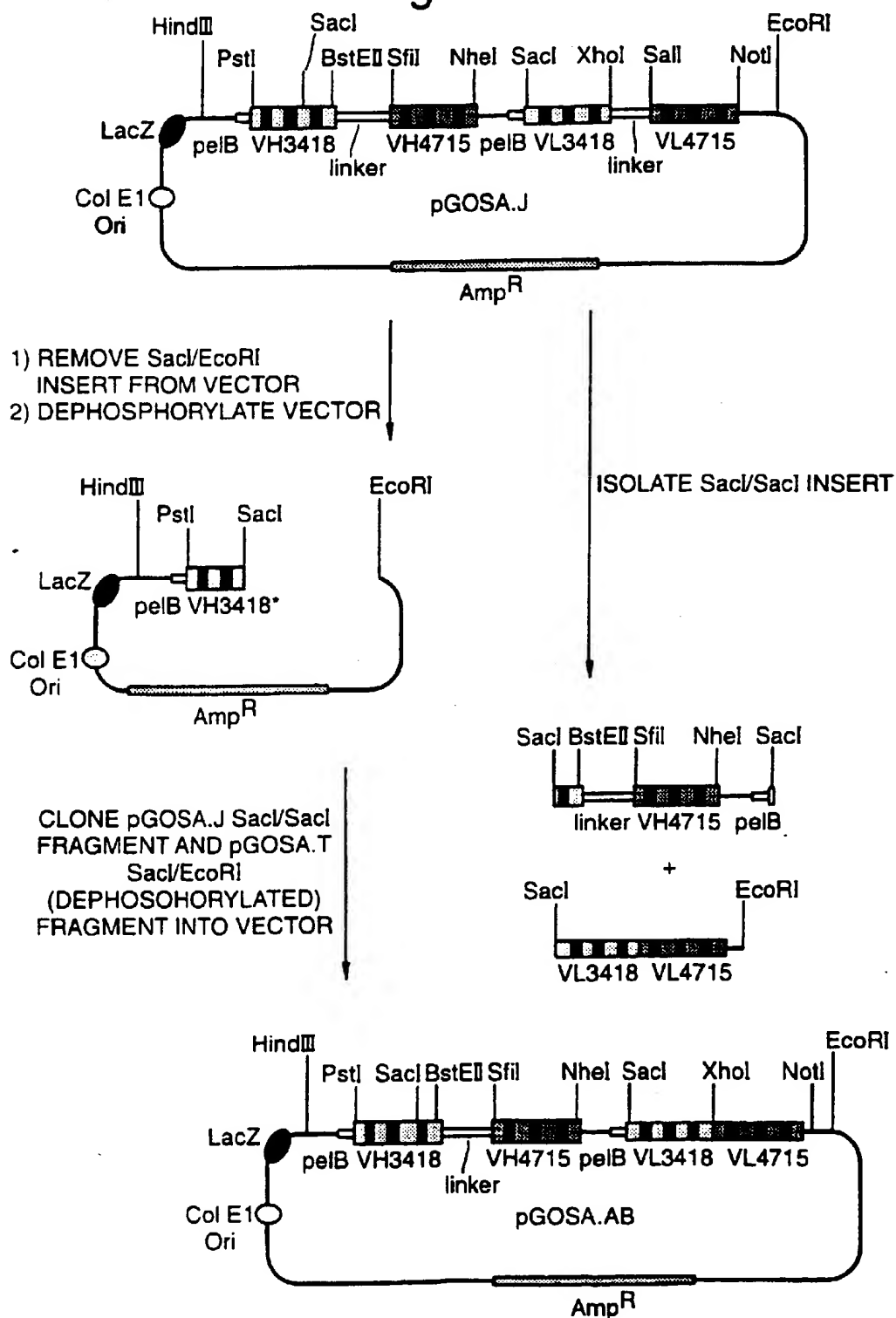


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Fig.41.

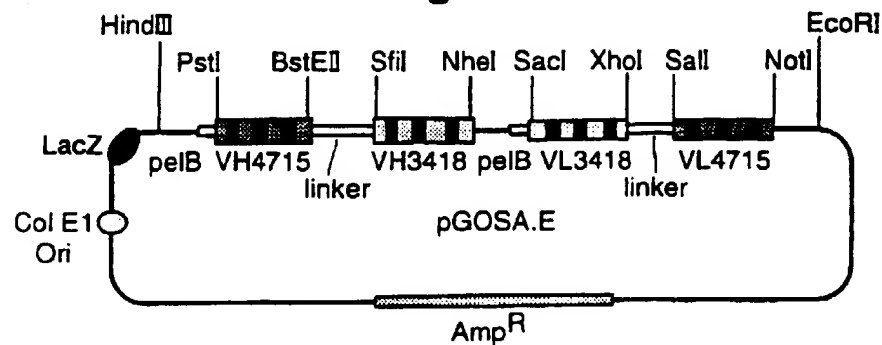


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Fig.42.

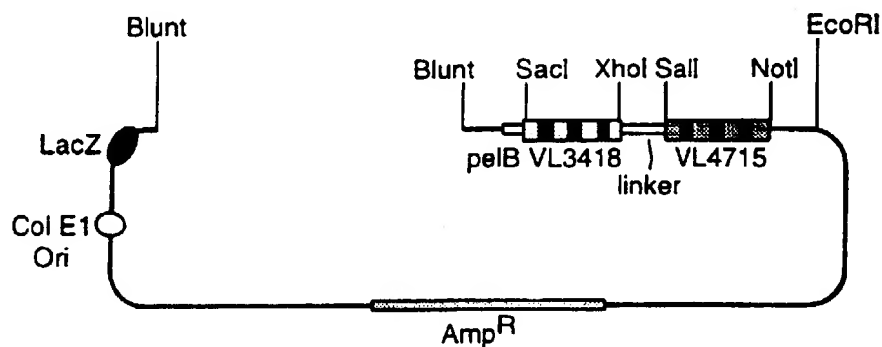


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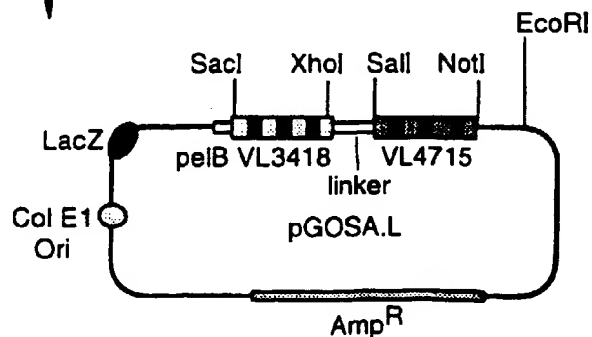
Fig.43.



- 1) REMOVE HindIII/NheI VH4715-LINKER-VH3418 FRAGMENT FROM VECTOR
- 2) FILL IN ENDS WITH KLENOW DNA POLYMERASE



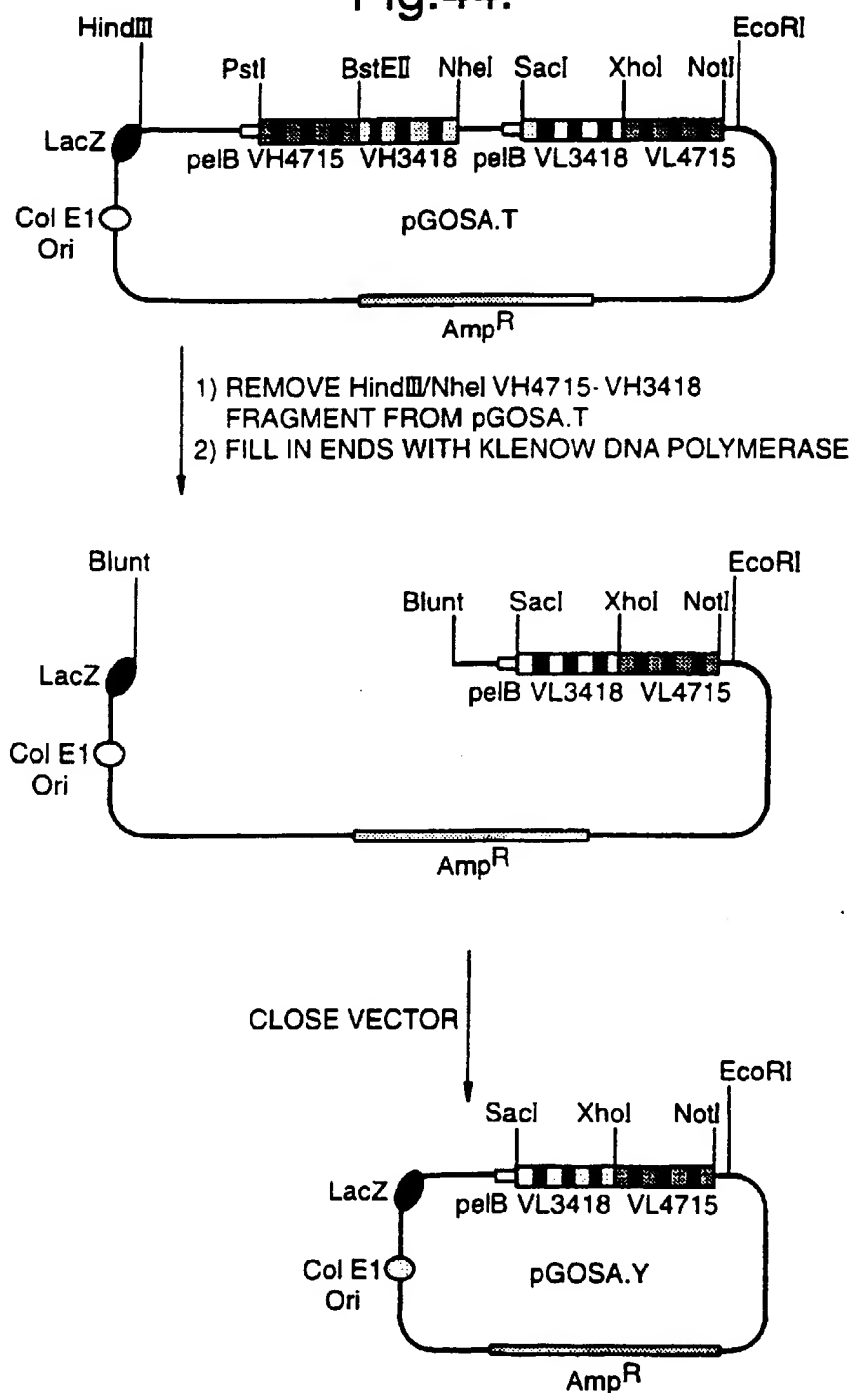
CLOSE VECTOR



SUBSTITUTE SHEET (RULE 26)

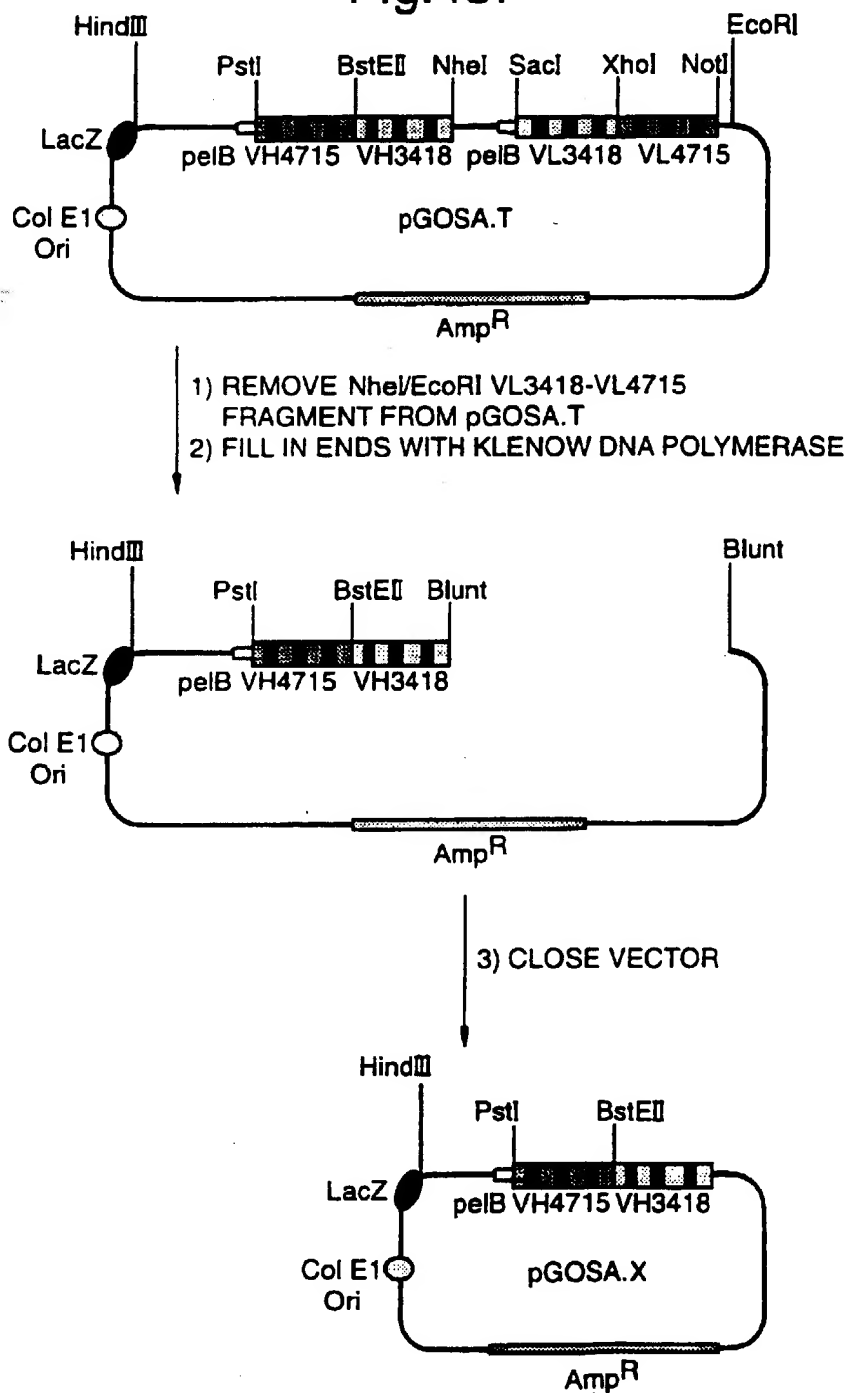
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Fig.44.



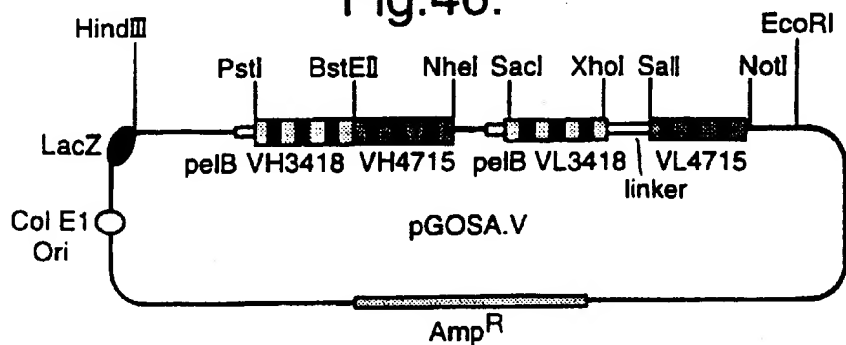
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Fig.45.

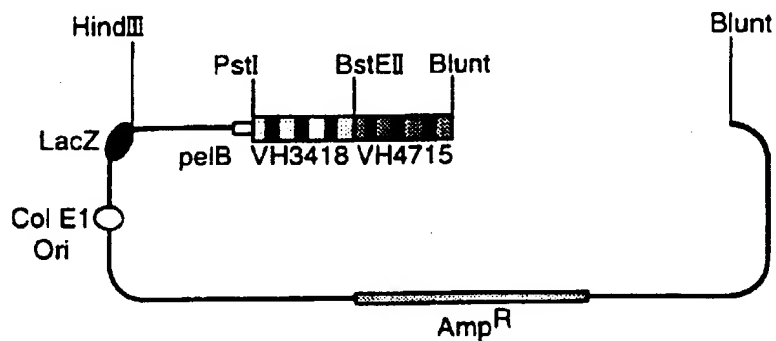


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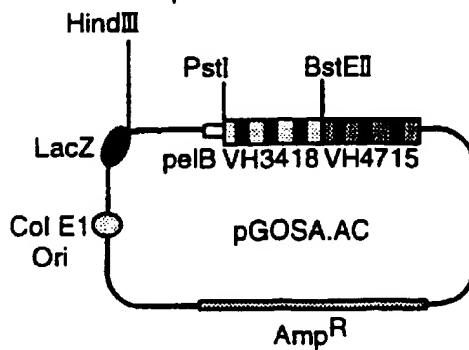
Fig.46.



- 1) REMOVE NheI/EcoRI VL3418-linker-VL4715
FRAGMENT FROM VECTOR
- 2) FILL IN ENDS WITH KLENOW DNA POLYMERASE

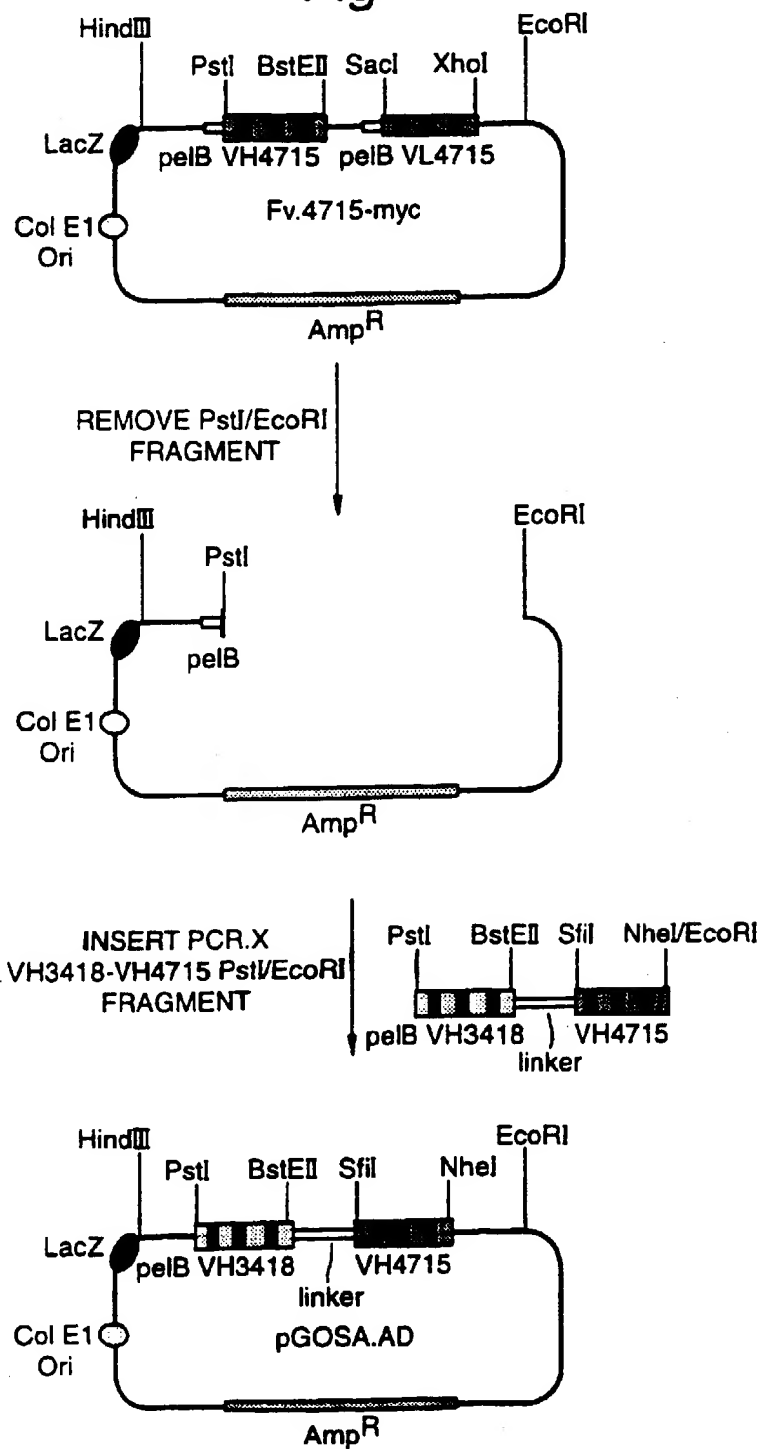


CLOSE VECTOR



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Fig.47.



INTERNATIONAL SEARCH REPORT

Inter- national Application No
PC1/EP 96/03605

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C07K16/46 C07K16/00 A61K39/395 G01N33/577

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C07K A61K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category * | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|------------|---|-----------------------|
| A,0 | BIOCHEMICAL SOCIETY TRANSACTIONS, vol. 23, no. 4, 18 - 21 July 1995, LONDON, GB, pages 1067-1073, XP000565752 M. VERHOEYEN ET AL.: "Antibody fragments for controlled delivery of therapeutic agents." see the whole document --- -/-- | 1-14 |

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

A document member of the same patent family

Date of the actual completion of the international search

16 December 1996

Date of mailing of the international search report

07.01.97

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx 31 651 epo nl,
Fax (+31-70) 340-3016

Authorized officer

Nooij, F

INTERNATIONAL SEARCH REPORT

International Application No

PC1/EP 96/03605

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

| Category * | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|------------|--|-----------------------|
| A | PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE USA, vol. 90, no. 14, 15 July 1993, WASHINGTON, DC, USA, pages 6444-6448, XP002021302 P. HOLLIGER ET AL.: "Diabodies": Small bivalent and bispecific antibody fragments." cited in the application see the whole document --- | 1-14 |
| A | THE JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 269, no. 1, 7 January 1994, BALTIMORE, MD, USA, pages 199-206, XP002021303 W. MALLENDER ET AL.: "Construction, expression, and activity of a bivalent bispecific single-chain antibody." cited in the application see abstract see figures 1,2 --- | 1-14 |
| A | WO 94 13806 A (THE DOW CHEMICAL COMPANY) 23 June 1994 cited in the application see figure 1 --- | 1-14 |
| A | WO 94 13804 A (CAMBRIDGE ANTIBODY TECHNOLOGY LTD. ET AL.) 23 June 1994 cited in the application see page 31, line 10 - line 12 see figure 1 --- | 1-14 |
| A | WO 93 11161 A (ENZON, INC.) 10 June 1993 cited in the application see page 22, line 1 - line 10 see claims ----- | 1-14 |

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PC1/EP 96/03605

| Patent document cited in search report | Publication date | Patent family member(s) | Publication date |
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| | | CA-A- 2117477 | 23-06-94 |
| | | EP-A- 0628078 | 14-12-94 |
| | | JP-T- 7503622 | 20-04-95 |
| ----- | | | |
| WO-A-9413804 | 23-06-94 | AU-A- 5654894 | 04-07-94 |
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| | | EP-A- 0720624 | 10-07-96 |
| WO-A- 9508577 | 30-03-95 | | |
| ----- | | | |
| WO-A-9311161 | 10-06-93 | AU-A- 3178993 | 28-06-93 |
| | | CA-A- 2122732 | 10-06-93 |
| | | EP-A- 0617706 | 05-10-94 |
| | | JP-T- 7501451 | 16-02-95 |
| ----- | | | |